

Patterns of Genetic Variation in
Rosette-*Brachyglottis*
are Inconsistent with
Current Species Delimitation

A thesis submitted in partial fulfilment
of the requirements for the Degree of
Master of Science in Plant Biology
at the University of Canterbury

by

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University of Canterbury

2014

Acknowledgements

I would like to thank the Miss E. L. Hellaby Indigenous Grasslands Research Trust for funding my field work and for hosting a highly enjoyable conference in Dunedin.

I would like to thank MBIE for core funding through Landcare Research and to all the staff at Landcare Research who made me feel very welcome.

Thank you to my Supervisors, Pieter Pelser and Rob Smissen, for their endless support in all aspects of this project and for introducing me to an area of science which I was previously unfamiliar with.

Thank you to David Drury for your advice and extensive morphological data on rosette-*Brachyglottis*.

Thank you to Mike Lusk, Kerry Ford and Peter Heenan for collecting rosette-*Brachyglottis* plants for me from locations I was unable to visit myself.

Thank you to Shannel Courtney for his detailed notes on rosette-*Brachyglottis* populations in the upper South Island that helped to get my field work underway.

Thank you to Nic Bolstridge, Dagmar Goeke, Caroline Mitchell and Katherine Trought for their help and advice in the lab on numerous occasions.

Thank you to Mary Korver, Ilse Breitwieser and Ines Schönberger for their help in the Allan Herbarium.

Thank you to Kate Boardman for recovering the identifications of so many CHR specimens.

Thank you to Maggie Tisch and Craig Galilee for their patience with genotyping my samples.

And finally thank you to all my friends and family for supporting me throughout my time at University.

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Abstract

Brachyglottis (Asteraceae) is a genus of approximately 30 species in the Brachyglottidinae, a recently recognised sub-tribe of tribe Senecioneae. Within *Brachyglottis* is a clade of five species of rosette-forming herbs: *B. bellidioides*, *B. haastii*, *B. lagopus*, *B. southlandica* and *B. traversii*. A sixth species, *B. saxifragoides*, has recently been synonymised with *B. lagopus*. The rosette-*Brachyglottis* have historically been recognised as a taxonomically problematic group because species overlap in both morphology and geographical distribution.

A recent molecular study of rosette-*Brachyglottis* using AFLP data indicated that genetic distances among populations of rosette-*Brachyglottis* in the South Island appear to be correlated with geographical distance between populations rather than taxonomic identification. This is problematic as the currently described rosette-*Brachyglottis* species have overlapping ranges which implicitly hypothesises reproductive barriers other than geographic distance.

We conducted an investigation into the species delimitation of rosette-*Brachyglottis* with the aim of answering two related questions: Does the current delimitation of rosette-*Brachyglottis* accurately reflect patterns of genetic similarity? Do the patterns of genetic structure in rosette-*Brachyglottis* support the presence of multiple biological species?

A total of 46 populations of rosette-*Brachyglottis* were represented in this study. Herbarium specimens collected from these populations were identified following the taxonomic treatment of Allan (1961). Twenty one discrete and numerical morphological characters were measured from herbarium specimens including those collected for this study and previously collected herbarium specimens. Morphological dissimilarity of 354 herbarium specimens was investigated by performing a PCoA on Gower's pairwise morphological distances among individuals. The pattern of genetic similarity was explored using DNA fragment length variation in nine markers for 273 individuals and this resulted in a total 177 unique alleles. Bayesian clustering analysis was performed on this data set using STRUCTURE, in addition, pairwise genetic distances were calculated among individuals and populations using Jaccard and Nei's dissimilarity coefficient's respectively. Jaccard genetic distances among individuals were analysed using PCoA and Nei's genetic distances among populations were analysed using a Neighbour-Net analysis. The relationship between pairwise genetic and geographic distances among populations was analysed using a combination of linear regression and a Mantel Test.

The pattern of morphological similarities among specimens was generally congruent with the currently delimited species in rosette-*Brachyglottis*. However, many morphologically intermediate specimens confound the recognition of distinct morphological entities. Comparison of patterns of genetic similarity and the current morphologically-based species delimitation showed that the delimitation does not accurately reflect the genetic structure of rosette-*Brachyglottis*. Furthermore, patterns of genetic dissimilarity did not indicate discrete genetic groups at the individual or population levels. The finding of incongruence between patterns of genetic and morphological similarity and absence of morphologically or genetically discrete groups suggests that rosette-*Brachyglottis* are best considered a single, yet morphologically diverse, biological species. In addition genetic structure within

this species appears to be primarily driven by geographical isolation.

Chapter 1

Introduction

1.1 Subtribe Brachyglottidinae

1.1.1 Overview of *Brachyglottis*

Brachyglottis J. R. & G. Forst. (Asteraceae) is a genus of approximately 30 species in Brachyglottidinae (Connor and Edgar 1987, Pelter et al. 2007, 2010). Brachyglottidinae is a sub-tribe of tribe Senecioneae which is named for the cosmopolitan genus *Senecio* (Pelter et al. 2007). *Brachyglottis* is a New Zealand endemic genus with the exception of a single Australian species, *B. brunonis* (Hook. f.) B. Nord. It is morphologically diverse and includes small rosette-forming herbs as well as secondarily-woody shrubs and trees (Allan 1961, Nordenstam 1978). *Brachyglottis* species occupy a wide variety of habitats growing from sea level to sub-alpine elevations. These include scrub and grasslands, rocky faces and riverbanks, and forest margins (Allan 1961). The morphological and ecological diversity of the *Brachyglottis* is hypothesised to be the result of rapid evolution and adaptation to a wide range of niches and the group has been cited as an example of adaptive radiation (Nordenstam 1978, Wagstaff and Breitwieser 2004).

Phylogenetic studies of *Brachyglottis* have raised multiple issues with the current treatment of the genus (Wagstaff and Breitwieser 2004, Pelser et al. 2007, Mennes et al. 2013). For example, they suggest that *Brachyglottis* is a paraphyletic genus because multiple other genera may nest within it including *Acrisione* B. Nord., *Bedfordia* DC., *Haastia* Hook. f., *Papuacalia* Veldkamp, and *Traversia* Hook. f., (Wagstaff and Breitwieser 2002, 2004, Pelser et al. 2007). Systematic delimitation of taxa aims to accurately portray evolutionary relationships among species by placing them into hierarchical, monophyletic groups. Paraphyletic classifications do not accurately depict this hierarchical structure and therefore, are not ideal. A further issue in the delimitation of *Brachyglottis* is that morphological and genetic analysis of the rosette-forming clade suggests that it may contain less species than previously thought (Wall 1918, Mennes et al. 2013).

The rosette-forming herbs of *Brachyglottis* form a clade of five species: *B. bellidioides* (Hook. f.) B. Nord., *B. haastii* (Hook. f.) B. Nord., *B. lagopus* (Raoul) B. Nord., *B. southlandica* (Cockayne) B. Nord., and *B. traversii* (F. Muell.) B. Nord. (Allan 1961, Wagstaff and Breitwieser 2004, Pelser et al. 2007). A sixth species, *B. saxifragoides* (Hook. f.) B. Nord., has recently been synonymised with *B. lagopus* (Mennes et al. 2013). The evolution of the rosette-forming group has been hypothesised to be an adaptive radiation into South Island temperate grasslands (Nordenstam 1978). Rosette-*Brachyglottis* are indeed usually found in exposed grassy or shrubby areas, ranging from coastal to sub-alpine environments. They are often associated with the exposed bedrock of cliffs, rock outcrops, and stream sides (Cockayne 1921, 1922, Allan 1961).

1.1.2 Historical Taxonomic Treatment of *Brachyglottis*

Most species currently included in *Brachyglottis* were initially included in *Senecio* (Nordenstam 1978, Connor and Edgar 1987). *Senecio* has historically been very

broadly defined and is one of the largest genera of flowering plants. It exhibits an exceptionally wide range in morphology and life histories (Pelser et al. 2007). *Senecio* is the largest genus of tribe Senecioneae, containing around one third of the approximately 3000 species of this tribe (Pelser et al. 2007). More recently, plant taxonomists and phylogeneticists have attempted to define *Senecio* more narrowly and identify subtribal clades in the Senecioneae (Nordenstam 1978, 1985, Pelser et al. 2007). Species currently assigned to *Brachyglottis* and many of its allied genera (*Acrisione*, *Dolichoglottis* B. Nord., *Papuacalia*, and *Traversia*) were formerly recognised in a much more broadly defined *Senecio* (Nordenstam 1978, 1985, Connor and Edgar 1987, Veldkamp 1991). However, more recently they were exuded from the genus based on morphological and cytological evidence (Nordenstam 1978, 1985, Connor and Edgar 1987, Veldkamp 1991). Subsequent phylogenetic analyses further increased the understanding of relationships among *Brachyglottis* and other genera of Senecioneae (Wagstaff and Breitwieser 2002, 2004, Pelser et al. 2007, 2010). The culmination of these analyses has lead to the recognition of the Brachyglottidinae, a sub-tribe of Senecioneae which includes *Acrisione*, *Bedfordia*, *Brachyglottis*, *Dolichoglottis*, *Haastia*, *Papuacalia*, and *Traversia* (Pelser et al. 2007, 2010).

Brachyglottis was first described for two species of small trees: *B. repanda* J. R. & G. Forst. and *B. rotundifolia* J. R. & G. Forst. (Forster et al. 1776). It was initially characterised by floral characters including the presence of large panicles of campanulate (bell shaped) capitula (flower heads), and white to creamy pistillate (female) ray-florets (Allan 1961). A third species, *B. arborescens* W. R. B. Oliver, was described from the Three Kings Islands. *Brachyglottis arborescens* was treated as a variety of *B. repanda* by Allan (1961) (*B. repanda* var. *arborescens* (W. R. B. Oliver) Allan), but was reinstated as a species by Drury (1973). Drury (1973) also reassigned *Senecio myrianthos* Cheeseman to

Brachyglottis (*B. myrianthos* (Cheeseman) Drury). *Brachyglottis* continued to grow as new species were described and existing species transferred to it from other genera on the basis of morphological similarities (Nordenstam 1978, Connor and Edgar 1987).

Nordenstam (1978) recognised two morphologically distinct groups of New Zealand *Senecio*, the ‘senecioid’ and the ‘cacalioid’ groups. Cacalioid *Senecio* species are generally distinguished from senecioid *Senecio* species by the following characters: chromosome number of $2n = 60$, style branches with undivided stigmatic areas, cylindrical filament collars that are no thicker than the filament, and polarised endothelial tissue of the anther wall (Nordenstam 1985). The majority of present day *Brachyglottis* species were originally classified as cacalioid *Senecio* (Nordenstam 1978, Connor and Edgar 1987). Morphological and cytological similarities between cacalioid *Senecio* and *Brachyglottis* species lead to the reassignment of most New Zealand cacalioid *Senecio* to *Brachyglottis* or the closely allied genera *Dolichoglottis*, and *Urostemon* B. Nord., (Nordenstam 1985).

Nordenstam (1978) transferred *S. lyallii* Hook. f., and *S. scorzonerooides* Hook. f. into the new genus *Dolichoglottis* as *D. lyallii* (Hook. f.) B. Nord., and *D. scorzonerooides* (Hook. f.) B. Nord. Nordenstam (1978) placed *S. kirkii* Hook fil. ex Kirk into the monotypic genus *Urostemon* based on “unusual” morphological characters: caudate (tail-like) anthers with papillate (protruding) asymmetric tails, T-shaped eglandular hairs, and the combination of white ray-florets with a smooth ligule (corolla) epidermis. *Urostemon kirkii* (Hook. f. ex Kirk) B. Nord. was later transferred to *Brachyglottis* as *B. kirkii* (Kirk) C. Webb on the basis that the morphological characters of *B. kirkii* sighted by Nordenstam (1978) were in fact also present in other *Brachyglottis* species (Jeffrey 1979, Connor and Edgar 1987). Excluding *S. turneri* Cheeseman, Nordenstam (1978) transferred all remaining New Zealand cacalioid *Senecio* species along with the Tasmanian

S. brunonis to *Brachyglottis*. *Senecio turneri* was not mentioned by Nordenstam (1978) but was later transferred to *Brachyglottis* as *B. turneri* (Cheeseman) C. Webb due to its cacalioid characters (Connor and Edgar 1987). Nordenstam (1978) also transferred the Tasmanian *Senecio brunonis* (Hook. fil.) J. H. Willis to *Brachyglottis* (as *B. brunonis* (Hook. f.) B. Nord.) due to similar, cacalioid, morphology. *Brachyglottis brunonis* was originally placed in the monotypic genus *Centropappus* Hook. f. (*C. brunonis* Hook. f.) (Hooker 1847). Connor and Edgar (1987) recognised a total of 30 New Zealand *Brachyglottis* species which, with the inclusion of the Tasmanian *Brachyglottis brunonis*, gives a total of 31 *Brachyglottis* species.

1.2 Phylogenetic Studies of Brachyglottidinae

1.2.1 Summary of Phylogenetic Methods

Early taxonomic treatments of the New Zealand Senecioneae were based on morphological characters and later a combination of morphological and cytological (i.e. chromosome numbers and cell morphology) characters (Hooker 1853, Allan 1961, Drury 1973, Beuzenberg and Groves 1974, Beuzenberg 1975, Nordenstam 1978). The development of modern genetic and statistical techniques has revolutionised the understanding of species relationships. These techniques have provided new sources of data and powerful analytical tools to explore these data. *Brachyglottis* and allies have been included in several phylogenetic studies (Wagstaff and Breitwieser 2002, 2004, Pelsner et al. 2007, 2010).

Phylogenetic analyses use tree-like diagrams as hypotheses of evolutionary history and relationships (Page and Holmes 1998). A phylogenetic analysis can hypothetically test a data set against all possible trees for a given number of taxa to find the tree that has the highest value of the chosen optimality criterion,

which is the tree that is assumed to show their correct evolutionary history (Page and Holmes 1998, Wiley and Lieberman 2011). However, the number of potential trees increases exponentially relative to the number of taxa in an analysis and therefore calculating the optimality ‘score’ of all possible individual trees is not feasible in most cases (Page and Holmes 1998). To avoid this issue, phylogenetic methods use algorithms to perform a heuristic search through parameter space (the hypothetical multidimensional space of all possible trees and associated parameter values such as branch lengths) and select candidate trees in a manner that increases the probability of finding the most optimal tree (Holder and Lewis 2003, Knowles and Kubatko 2011). Many of these algorithms work by slightly altering the current candidate tree until it produces a more optimal candidate tree (Holder and Lewis 2003, Wiley and Lieberman 2011). In this manner the algorithm can move through parameter space towards ‘peaks’ of optimal trees (Holder and Lewis 2003, Wiley and Lieberman 2011). For each candidate tree, the number and or nature of character changes (evolutionary steps) along all edges of the candidate tree are assessed against the optimality criterion and the most optimal tree(s) can be determined (Holder and Lewis 2003, Wiley and Lieberman 2011).

The core assumption of a maximum parsimony analysis is that evolutionary change is rare and therefore that the tree requires us to assume the fewest character state changes is the best estimate of the true evolutionary history (Goloboff 2003, Wiley and Lieberman 2011). Statistical support for the clades of each tree is commonly calculated using methods of estimating the sampling error with pseudo-replicate sampling including the bootstrap and jackknife techniques (Page and Holmes 1998, Wiley and Lieberman 2011). Bootstrapping is a process that involves random re-sampling with replacement from the observed data (Mooney and Duval 1993, Wiley and Lieberman 2011). The ‘new’ data is used to re-run the

analysis and this process is repeated thousands of times (Mooney and Duval 1993). Bootstrap values for each clade of the tree are calculated based on the number of replicates that recover the same clade (Page and Holmes 1998). Jackknifing is a method of testing subsets of the observed data and is usually implemented by resampling without replacement (Wiley and Lieberman 2011). Maximum parsimony is a widely used method because it makes few explicit assumptions and it is relatively quick when compared to some other phylogenetic methods (Page and Holmes 1998, Goloboff 2003). Maximum parsimony does not always cope well with homoplasy especially when branches are of vastly uneven lengths which can cause long branches to be interpreted as a single branch (long branch attraction) (Page and Holmes 1998). Because long branch attraction is more likely when terminal branches are of uneven length, it can be minimised by the inclusion of many taxa to ‘break up’ long terminal branches (Heath et al. 2008).

Other phylogenetic techniques, such as maximum likelihood, use an evolutionary model to infer the likelihood that each tree explains the observed data (i.e. likelihood is the optimality criterion) (Page and Holmes 1998, Knowles and Kubatko 2011). An evolutionary model incorporates the nature of evolutionary steps as well as the number of steps (Page and Holmes 1998). For example, phylogenetic hypotheses that require a number of rare evolutionary changes to have happened can be considered less likely than alternative hypotheses that assume the same number of more common evolutionary changes (Page and Holmes 1998). Models of the evolution of DNA sequence data often treat nucleotide transversions and less probable than transitions (Page and Holmes 1998). In a maximum likelihood analysis a model of evolution is used to calculate the likelihood of each candidate tree explaining the observed data (Knowles and Kubatko 2011, Wiley and Lieberman 2011). The likelihood value is a measure of how likely a tree is to have given rise to the observed data under the model of

evolution that it was calculated with (Holder and Lewis 2003). Unlike maximum parsimony, a maximum likelihood analysis can allow for multiple evolutionary changes of a single character along the same branch (Page and Holmes 1998). This allows a maximum likelihood analysis to incorporate the possibility of unobserved ancestral character states (Page and Holmes 1998). The use of a evolutionary model is one of the strengths of maximum likelihood, however the use of a model with inaccurate assumptions may produce a less accurate tree (Page and Holmes 1998, Goloboff 2003). Therefore, the selection of an appropriate evolutionary model is critical for maximum likelihood analyses (Goloboff 2003). Bootstrapping and or jackknifing can be used to assess confidence for each clade in a maximum likelihood tree in the same manner as it is used in maximum parsimony (Wiley and Lieberman 2011). The main problem presented by maximum likelihood analysis is that it is computationally very intensive and therefore slower than other phylogenetic analyses (Page and Holmes 1998).

A relatively new phylogenetic approach is Bayesian inference of phylogeny (Archibald et al. 2003, Holder and Lewis 2003). The field of Bayesian statistics is defined by its use of prior knowledge in addition to observed data to calculate the probability of an event occurring (Archibald et al. 2003). A prior probability is the known probability of an event occurring and is usually calculated from previous occurrences of events that are similar to the one being studied (Archibald et al. 2003). In the case of Bayesian inference of phylogeny, little is known about the prior probability of a candidate tree being the ‘true’ tree and flat priors are usually used (i.e. all candidate trees are assigned equal prior probability at the start of an analysis) (Archibald et al. 2003). Bayesian inference uses an evolutionary model to infer the posterior probability of each tree explaining the observed data (i.e. posterior probability is the optimality criterion) (Holder and Lewis 2003, Knowles and Kubatko 2011). Posterior probabilities are calculated from prior probability

and likelihood function (Holder and Lewis 2003, Knowles and Kubatko 2011).

Bayesian inference of phylogeny uses a Markov chain Monte Carlo (MCMC) algorithm to search parameter space and approximate posterior probability distributions (Holder and Lewis 2003, Knowles and Kubatko 2011). The MCMC moves through parameter space by selecting a new point close to its current position by perturbing the parameters of its current position (Holder and Lewis 2003). The posterior probability density of its current position is compared to that of the proposed position and the chain will move to the proposed position if its posterior probability density is higher (Holder and Lewis 2003, Wiley and Lieberman 2011). On a random basis the MCMC will occasionally move to areas of lower probability in order to avoid getting stuck in sub optimal peaks (Regions of parameter space that are more optimal than neighbouring regions but are not the most optimal region) (Holder and Lewis 2003). By design, the the MCMC spends more time in areas with greater posterior probability density than in areas with lower posterior probability density (Holder and Lewis 2003, Wiley and Lieberman 2011). By recording the time spent in each area of parameter space, the MCMC can accurately estimate the posterior probability of each tree (Holder and Lewis 2003, Wiley and Lieberman 2011). The posterior probability can also be estimated for each clade of a tree by determining the proportion of time that the MCMC spent in trees with that clade (Wiley and Lieberman 2011).

1.2.2 Application to Brachyglottidinae

Wagstaff and Breitwieser (2002) conducted a phylogenetic study of New Zealand Asteraceae using ITS sequences from 95 species representing all New Zealand Asteraceae genera. In this study genus *Brachyglottis* was represented by the rosette-forming herb *B. southlandica* and the woody shrub *B. elaeagnifolia* (Hook fil.) B. Nord. In the resulting consensus tree of 660 maximum parsimony trees,

Brachyglottis southlandica and *B. elleagnifolia* formed a unresolved polytomy with four other genera: *Bedfordia*, *Dolichoglottis*, *Haastia* and *Traversia* (Wagstaff and Breitwieser 2002). *Dolichoglottis* and *Traversia* are both New Zealand genera of cacalioid Senecioneae species and have been recognised as closely allied to *Brachyglottis* (Nordenstam 1978). *Bedfordia* is an Australian genus of small to medium sized trees (Nordenstam 1978, Orchard 2004). *Bedfordia* is another genus of Brachyglottidinae and has been recognised as morphologically similar and allied to *Brachyglottis* (Nordenstam 1978, Orchard 2004). However, morphological differences in leaf venation, hair types, and floral characters have been used to justify the recognition of *Bedfordia* as a separate genus from *Brachyglottis* (Nordenstam 1978, Orchard 2004). *Haastia* is a New Zealand genus that was not previously recognised as either an ally of *Brachyglottis*, nor a member of tribe Senecioneae (Breitwieser and Ward 2005, Wagstaff and Breitwieser 2002). *Haastia* has undergone a number of taxonomic revisions including placement in tribes Gnaphalieae and then Astereae, the splitting then reforming of the genus, and more recently the intact genus not assigned to a tribe (Breitwieser and Ward 2005). Breitwieser and Ward (2005) found that, based on morphological characters, *Haastia* could be excluded from all the major tribes of Asteraceae except Senecioneae and therefore should be allocated to tribe Senecioneae. The morphological placement of *Haastia* in Senecioneae is congruent with its phylogenetic nesting in *Brachyglottis* (Wagstaff and Breitwieser 2004, Pelter et al. 2007).

Wagstaff and Breitwieser (2004) specifically investigated phylogenetic relationships among *Brachyglottis* and other allied genera of the *Brachyglottis* clade described in Wagstaff and Breitwieser (2002). In their study *Bedfordia* was represented by two taxa, *Brachyglottis* by 15 species, *Haastia* by three species, and *Dolichoglottis* and *Traversia* were each represented by a single species. This study used ITS and 5' *trnK/matK* regions both separately and in combination to

produce three maximum parsimony consensus trees.

The strict consensus tree derived from 4632 maximum parsimony trees based on the ITS genetic region recovered a similar polytomy to Wagstaff and Breitwieser (2002). Wagstaff and Breitwieser (2004) recovered several clades within the larger polytomy. *Haastia recurva* and *H. sinclairii* formed a well supported clade (bootstrap value of 92) while the third (*H. pulvinaris*) attached directly to the large polytomy. The Australian genus *Bedfordia* was represented by two taxa (*B. linearis* and *B. salicina*) which formed a highly supported clade (bootstrap value of 100). The *Bedfordia* clade formed a larger clade with *Brachyglottis brunonis* (bootstrap value of 91) which nested in the large polytomy (Wagstaff and Breitwieser 2004).

A strict consensus tree derived from 94000 maximum parsimony trees obtained from the 5' *trnK/matK* data set produced a larger polytomy that included the *Brachyglottis* alliance, *Petasites fragrans* and a clade of *Senecio* species (Wagstaff and Breitwieser 2004). Furthermore, fewer clades were recovered within the larger polytomy and those clades had lower bootstrap values when compared with those of the ITS analyses (Wagstaff and Breitwieser 2004). Fewer recovered clades and less support of those clades reflect that the 5' *trnK/matK* consensus tree has less phlogenetic resolution.

Pelser et al. (2007) conducted a global phylogenetic analysis of tribe Senecioneae. This study used a combination of maximum parsimony and Bayesian inference of ITS and plastid DNA sequences. Unlike Wagstaff and Breitwieser (2004) who produced strict consensus trees with large polytomies, Pelser et al. (2007) presented a mostly bifurcating tree describing relationships in the *Brachyglottis* alliance. This was done in order to produce a comprehensive hypothesis of phylogenetic relationships in Senecioneae for the purpose of testing sub-tribal classifications.

As a result of this, most nodes depicting relationships within the *Brachyglottis* alliance in Pelser et al. (2007) have either no support (both bootstrap and Bayesian consensus percentages < 50) or mixed to weak statistical support (low bootstrap with low to high Bayesian consensus percentages). If nodes with no/weak support in Pelser et al. (2007) were collapsed, the ITS consensus tree would show a polytomy similar to the polytomy recovered by Wagstaff and Breitwieser (2004).

Analysis of ITS sequences in Pelser et al. (2007) supported the findings of Wagstaff and Breitwieser (2004) that the genera *Traversia*, *Haastia* and *Bedfordia* nest within *Brachyglottis* (Pelser et al. 2007). *Dolichoglottis* (*D. scorzoneroide*s) was found to form a sister clade to *Brachyglottis* and all other Brachyglottidinae represented in the study. However, statistical support for this node was mixed with a bootstrap value of less than 50 and a Bayesian consensus percentage of 95 (Pelser et al. 2007).

Pelser et al. (2007) also found that two additional non-New Zealand genera, *Acrisione* (Chile), and *Papuacalia* (New Guinea), were nested within *Brachyglottis*. *Acrisione* is a genus of two species of small trees or shrubs that are endemic to central Chile (Nordenstam 1985, Moreira-Muñoz and Muñoz-Schick 2007). Both species (*A. cymosa* (Remy) B. Nord., and *A. denticulata* (Hook. & Arn.) B. Nord.) were formerly included in *Senecio* (*S. cymosa* and *S. denticulata*) (Nordenstam 1985). Based on morphological characters and chromosome counts ($2n = 60$) Nordenstam (1985) recognised *S. cymosa* and *S. denticulata* as cacalioid species and reassigned them to the new genus *Acrisione* on the basis that their cacalioid characters distinguished them from the senecioid-*Senecio*.

Papuacalia consists of seventeen species of small trees and large woody shrubs endemic to high altitude areas of New Guinea (Veldkamp 1991, Hind 2002, Hind and Johns 2003). Some *Papuacalia* species were originally assigned to *Senecio*

and others provisionally identified as *Bedfordia* until they were transferred to the newly created *Papuacalia* (calia as in the “cacalioid” *Senecio*) (Veldkamp 1991). Veldkamp (1991) distinguished *Papuacalia* from *Bedfordia* by the presence of a hypodermis in the leaf blades, vascular bundles with a lignified bundle sheath, different glandular hairs, different capitula, and young leaves that are in-folded and overlapping. This is opposed to revolute young leaves that are found in *Bedfordia* (Veldkamp 1991). Cytological analysis of two *Papuacalia* species (*P. dindondl* (Royen) Veldkamp, and *P. mogrere* (Royen) Veldkamp) revealed that the genus has a chromosome number of $2n = 72$ (Veldkamp 1991). This is unusual when compared to $2n = 60$ in *Bedfordia* and *Brachyglottis* or $2n = 40, 80$, or 100 common in senecioid *Senecio* (Nordenstam 1978, Veldkamp 1991, Beuzenberg 1975).

Pelser et al. (2007) found that Brachyglottidinae was the sister group of a clade of three African succulent species that were initially placed in *Senecio*. However, species in this clade were subsequently found to be only remotely related to *Senecio* and therefore, were later placed in the newly erected genus *Caputia*: *C. medley-woodi*, *C. pyramidatus* and *C. scaposus* based on on ITS data (Nordenstam and Pelser 2012). The Brachyglottidinae and *Caputia* group formed a well supported *Caputia*-Brachyglottidinae clade (bootstrap value = 99 and Bayesian consensus percentage = 100) (Pelser et al. 2007). While ITS data supported the monophyly of the *Caputia*-Brachyglottidinae clade, plastid data did not. Instead, plastid data suggests that the *Caputia* is more closely related to other Senecioneae than Brachyglottidinae (bootstrap value = 80 and Bayesian consensus percentage = 100) (Pelser et al. 2007). Incongruence between ITS and plastid data was highly supported and therefore indicates an underlying divergence between the phylogenetic signals of the ITS and plastid data sets (Pelser et al. 2007).

Highly supported, yet incongruent phylogenetic signals between gene regions regarding the phylogenetic position of a taxon suggests differing evolutionary histories of those gene regions (Pamilo and Nei 1988, Pelsner et al. 2010). Events capable of causing incongruence between the phylogenetic signals of different gene regions include hybridisation and incomplete lineage sorting (Knowles and Kubatko 2011). Both of these events allow an organism or species to inherit gene regions with differing evolutionary histories (Pelsner et al. 2010, Knowles and Kubatko 2011). When plants of two species hybridise the resulting offspring inherits its nuclear genetic material from either parent species as well as the phylogenetic signal associated with that material. The offspring will also inherit the plastid genetic material from a single parent. Assuming the hybrid is fertile, it may found a new species of hybrid origin that has genetic regions with incongruent phylogenetic signals (Knowles and Kubatko 2011). Alternatively the hybrid may backcross with one or both parent species, thereby transferring genetic material and associated phylogenetic signal between the parent species in a process called introgression (Knowles and Kubatko 2011).

Incomplete lineage sorting (ILS) is a process in which the genetic lineages differ from the speciation pattern of a group of species (Knowles and Kubatko 2011). ILS does not involve the introduction of genetic material from one species to another species and is often associated with large populations and rapid speciation events (Pamilo and Nei 1988, Kubatko and Meng 2011). It occurs when polymorphism in a DNA region persists in a population throughout multiple speciation events (Kubatko and Meng 2011). Polymorphisms generally persist for longer when reproductive populations are large (Pamilo and Nei 1988, Takahata 1989). In the example shown in figure 1.1 two consecutive speciation events have given rise to species A, B and C with common ancestry. However the relationships of the gene lineages (Blue, Green and Red) do not match the pattern of speciation due to

the polymorphism that predates the speciation events. Phylogenetic analysis of this genetic region would suggest that species A and B are more closely related to one another than either is to species C. Therefore, the phylogenetic signal of this genetic region would be incongruent with that of genetic regions that have not experienced ILS.

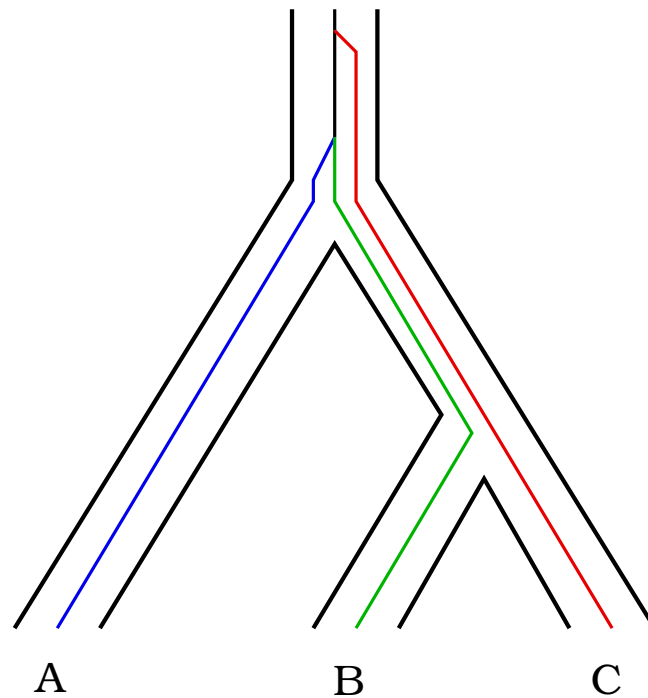


Figure 1.1: A hypothetical example of incomplete lineage sorting. Species B and C share a more recent common ancestor with one another than they do with species A. The lineage of the genetic region shown in blue green and red is incongruent with the pattern of speciation.

Distinction between these events is important for the accurate reconstruction of species evolutionary histories (Knowles and Kubatko 2011). However, it can be difficult to determine whether phylogenetic incongruence between gene trees is caused by hybridisation or ILS because both hybridisation and ILS produce similar symptoms. Many studies have investigated methods of determining whether cases of phylogenetic incongruence among gene regions can be specifically attributed to

either historical hybridisation or ILS but there is no universally accepted method for doing this (Maddison and Knowles 2006, Joly et al. 2009, Pelser et al. 2010, Yu et al. 2013).

Pelser et al. (2010) further investigated the presence of highly supported incongruence between genomic and plastid phylogenies in the Senecioneae. They trialled a method of distinguishing between hybridisation and ILS using molecular dating to estimate the length of potential ILS events. They hypothesised that, if the length of time taken by a potential ILS event requires an “irrationally large” population size, hybridisation was the more likely cause of incongruent phylogenies. However, if an irrationally large population was not required to explain the length of an ILS event, this method can not distinguish between hybridisation and ILS events. Pelser et al. (2010) found that genomic and plastid data produced highly supported, yet incongruent, phylogenetic relationships between the *S. medley-woodii* and *Brachyglottis* clades. Furthermore they found that the length of time required for a ILS explanation of this incongruence would require much larger effective population sizes than observed for these taxa. Therefore, Pelser et al. (2010) concluded that a historic hybridisation event is the most likely cause of the observed incongruence between genomic and plastid phylogenies.

Relationships between species of the *Brachyglottis* alliance are not sufficiently understood. Phylogenetic studies suggest that *Brachyglottis* is a polyphyletic genus with multiple other genera nested within it including *Acrisione*, *Bedfordia*, *Haastia*, *Papuacalia* and *Traversia* (Wagstaff and Breitwieser 2004, Pelser et al. 2007). Geographically these genera span the width of the South Pacific Ocean and include representatives from Australia, Chile, New Guinea, Tasmania and New Zealand. Some of these nested genera may also be polyphyletic, for example *Haastia pulvinaris* appears to nest within the *Brachyglottis* at a separate position than other *Haastia* species (Wagstaff and Breitwieser 2004, Pelser et al. 2007). The

taxonomic inclusion of all the nested genera into an expanded *Brachyglottis* would be an unsatisfactory solution as there are significant morphological and cytological differences between the various groups (Nordenstam 1978, 1985, Veldkamp 1991, Orchard 2004). Therefore, a more satisfactory solution would involve splitting *Brachyglottis* into multiple genera. However, this would require a better understanding of phylogenetic species level relationships within Brachyglottidinae. The relationship between *Papuacalia* and other genera of the alliance is particularly in need of further investigation because of its unusual chromosome count. Genus *Dolichoglottis* seems to be a well supported sister to all other Brachyglottidinae species and because of this it is unlikely to need reclassification (Wagstaff and Breitwieser 2004, Pelser et al. 2007). Although phylogenetic analyses suggest that *Brachyglottis* is a polyphyletic genus, several subgroups of the genus form well support clades. The best supported of these is the clade of rosette-forming *Brachyglottis* which is represented by *B. bellidioides*, *B. haastii* and *B. southlandica* in both Wagstaff and Breitwieser (2004) and Pelser et al. (2007). These three species formed well supported clades based on analysis of both the ITS and the 5' *trnK/matK* genetic regions (Wagstaff and Breitwieser 2004, Pelser et al. 2007).

Chapter 2

Rosette-*Brachyglottis*

2.1 Introduction to Rosette-*Brachyglottis*

Brachyglottis contains a clade of rosette-forming herbs (Wagstaff and Breitwieser 2004, Pelser et al. 2007). The currently recognised species in this clade are *B. bellidioides*, *B. haastii*, *B. lagopus*, *B. southlandica* and *B. traversii* (Allan 1961, Mennes et al. 2013). A sixth species, *B. saxifragoides*, has historically been recognised from populations on the Port Hills of Banks Peninsula, Canterbury, and has recently been synonymised with *B. lagopus* (Hooker 1853, Wall 1918, Mennes et al. 2013). All species of rosette-forming *Brachyglottis* were initially placed in *Senecio* and later transferred to *Brachyglottis* by Nordenstam (1978) along with many other New Zealand species of cacalioid *Senecio*.

Rosette-*Brachyglottis* are readily distinguished from their woody relatives by their sessile growth form composed of a rosette of radical (growing from ground level) leaves growing from a dense fibrous rhizome (Allan 1961). The rhizome is often covered in brown silky hairs and is deeply rooted (Hooker 1853, Allan 1961). Leaves may be appressed (flat against the ground) to erect and spreading (Hooker 1853, Allan 1961). Petioles (stem) and lamina (blade) of the leaf can vary remarkably (Allan 1961). The petiole can be nearing absent in smaller specimens

of *B. bellidioides* or up to 15cm in larger specimens of *B. saxifragoides* and *B. haastii* (Allan 1961). Shape of the leaf lamina may vary within and among species, ranging from orbicular (circular) to elongated-elliptic or ovate (elliptical with one end wider than the other) (Allan 1961). Attachment of the leaf and petiole ranges from cuneate (lamina narrows into petiole) to cordate (lamina is lobed around the petiole) (Allan 1961). The leaf tip is often obtuse (rounded) but may be acute (pointed) (Allan 1961). Length and width of the leaf lamina varies substantially both within and among species from a length of 1cm in smaller *B. bellidioides* up to 20cm in *B. saxifragoides* and with a width of up to 15cm in *B. saxifragoides* (Allan 1961).

When flowering, rosette-*Brachyglottis* species produce one or more scapes (flower stems) that each hold one to many capitula (flower heads) (Allan 1961). In specimens with multiple capitula on a single scape, the scape is branched (Hooker 1853, Allan 1961). A branched scape may have primary branching (all branches coming from the central stem) or secondary branching (branches coming off branches). Scapes tend to be more highly branched in larger specimens or species and are often unbranched in smaller specimens (Hooker 1853, Allan 1961). However this is not always the case and some larger specimens will produce many capitula on separate unbranched scapes. Scapes can vary dramatically in size and may exceed 30cm in height (Allan 1961).

A key component of rosette-*Brachyglottis* morphology is the abundance and placement of various hair types on all surfaces of the plant (Hooker 1853, 1864, Kirk 1899, Cockayne 1914, Allan 1961). These hairs can broadly be divided into glandular and eglandular hairs (Wall 1918). Glandular hairs grow from multiple cells and often have a variegated appearance due to red-black material in some of the hair cells (Wall 1918). Glandular hairs have historically been described as bristles, hispid hairs and setae by flora writers but these vary from one another

only in size and length and presence of coloured material (Hooker 1853, Allan 1961, Wall 1918). Glandular hairs may be found on both the upper and lower surfaces of the lamina and are common around the lamina margin and on the scape (Allan 1961, Wall 1918). They are usually white (lack the red pigmentation) when found on lamina surfaces and are more often red-black when present around the lamina margin and on the petiole and scape (see figure 2.1) (Hooker 1853, Wall 1918). Eglandular hairs are non-glandular and grow from a single cell (Wall 1918). Eglandular hairs can appear as long ‘silky’ hairs scattered amongst glandular hairs in some species (*B. lagopus* and *B. saxifragoides*) (Hooker 1853, Allan 1961, Wall 1918). They may also form a densely matted tomentum in some species which may be thick and woolly as in the lower lamina of *B. lagopus* and *B. saxifragoides*, or thinner and subappressed (very tightly matted) as seen on both upper and lower laminae of *B. haastii* (Hooker 1853, Allan 1961). Both hair types may be found on the bracts (small leaf like structures) and surfaces of the scape (Hooker 1853, Allan 1961).



Figure 2.1: White and red glandular hairs on a young leaf and the scape of *B. lagopus* from the Kaweka Range, North Island.

2.2 Rosette-*Brachyglottis* Species

Rosette-*Brachyglottis* species are found throughout the South Island, Stewart Island and the lower North Island of New Zealand (Allan 1961). In the South Island all five species are represented and their type specimens are from South Island locations with the exception of *Brachyglottis bellidioides* var. *crassa* (Simpson & Thomson) B. Nord., which was described from plants from Stewart Island (Allan 1961, Simpson and Thomson 1943). In the South Island, there is substantial overlap in the distribution ranges of all rosette-*Brachyglottis* species with the exception of *B. saxifragoides* (Allan 1961, Wall 1918, Cockayne 1921).

Brachyglottis lagopus. *Brachyglottis lagopus* was described by Raoul (1844) as *Senecio lagopus* from a population near Akaroa, Banks Peninsula, Canterbury, and was the first species of the rosette-group to be described. Hooker (1853) described *B. lagopus* as having two to four inch (circa five to ten cm) leaves on stout, woolly petioles with silky brown hairs covering the rhizome. The size range of the leaves has been expanded several times (Hooker 1864, Kirk 1899) and has most recently been set at 2.5 to 15cm by Allan (1961). The lamina was described by Hooker (1853) as entire (smooth margins), blunt and round to elliptic, rugose (wrinkled) with stout setae (glandular hairs) on upper lamina and woolly (eglandular) lower lamina. Later authors observed a wider range of leaf shapes including a cordate base and or crenate margins (Hooker 1864, Kirk 1899, Allan 1961). Allan (1961) also describes white silky (eglandular) hairs scattered among the glandular hairs of the upper lamina. Hooker (1853) describes scapes that are six inches high (15cm), branched with multiple capitula, and covered with black glandular hairs. The size and branching of the scape has also expanded in later descriptions and Allan (1961) describes the scape as up to 35cm tall and branched or simple (a single capitula). *Brachyglottis lagopus* is the only member of the rosette-forming group that is found in the North Island with the northern

extent of its distribution at approximately latitude 39° South (Allan 1961). This species is also found in the South Island as far south as latitude 45° South (Allan 1961). The type locality of *B. lagopus* is “Akaroa in montosis”, Banks Peninsula, Canterbury (Raoul 1844, Allan 1961).

Brachyglottis saxifragiodes. Hooker (1853) recognised two additional species of rosette-*Senecio*; *B. bellidioides* and *B. saxifragiodes* (as *S. bellidioides* and *S. saxifragiodes*), stating that they and *B. lagopus* “are closely allied and very singular species”. *Brachyglottis saxifragiodes* was distinguished from *B. lagopus* by its larger leaves with shorter petioles, larger and more branched scapes, and lack of hispid hairs on the upper lamina of leaves (Hooker 1853). Allan (1961) described *B. saxifragiodes* as having thick leaves, seven to 20cm long, that are sub-orbicular or broad-elliptic shaped with an obtuse tip and cordate base. The lower lamina is woolly with dense tomentose (eglandular) hairs and the upper lamina with silky (eglandular) hairs or nearly glabrous (Allan 1961). The scape is up to 45cm, usually branched and covered with eglandular and glandular hairs (Allan 1961). *Brachyglottis saxifragiodes* is limited to the Port Hills of Banks Peninsula, Canterbury and its type locality is “Port Cooper” near Lyttelton, Banks Peninsula (Hooker 1853, Allan 1961).

Brachyglottis bellidioides. *Brachyglottis bellidioides* was distinguished from *B. lagopus* by its smaller size, single headed scapes, and glabrous lower lamina of leaves (Hooker 1853). It was further distinguished from *B. lagopus* by the presence of hispid hairs on the upper lamina (Hooker 1853). The “hispid” hairs of *B. bellidioides* were confirmed to be glandular hairs that are structurally the same as those of *B. lagopus* (Wall 1918).

Tomentose forms of *B. bellidioides* were first recorded by Hooker (1864) who recognised [*B.*] *bellidioides* var β which was characterised by a “densely woolly”

lower lamina. Kirk (1899) viewed tomentum on the lower lamina as a character of ‘typical’ *B. bellidioides* (*B. bellidioides* var. *bellidioides*) and did not recognise a *B. bellidioides* variety that explicitly had a tomentose lower lamina. It is not clear why Kirk (1899) circumscribed forms of *B. bellidioides* with a tomentose lower lamina in a differently manner from Hooker (1864). Allan (1961) circumscribed *B. bellidioides* in similar manner to Kirk (1899), recognising a tomentose lower lamina as a possible feature of *B. bellidioides* var. *bellidioides* and not explicitly stating it as a character of any named varieties.

Leaf size of *B. bellidioides* has varied in descriptions and was recorded as up to four inches (ten cm) by Kirk (1899), however, Allan (1961) placed the upper limit at five cm long which is in agreement with the initial description of Hooker (1853). Descriptions of the scape have also expanded to include branched forms up to 30cm tall (Hooker 1864, Kirk 1899, Allan 1961). The scape is often near glabrous with few woolly or glandular hairs (Hooker 1853, Allan 1961) *Brachyglottis bellidioides* and its varieties are distributed throughout the South Island and Stewart Island, its type locality is “Canterbury” (Hooker 1853, Allan 1961).

Flora writers have recognised many varieties of *B. bellidioides* (see table 2.1) that mostly vary in leaf hairs and shape (Hooker 1864, Kirk 1899, Simpson and Thomson 1942, 1943, Allan 1961). Allan (1961) recognised five varieties of *B. bellidioides*. The geographical ranges of these varieties are generally not well established and the type localities of some are unknown (Allan 1961). *Brachyglottis bellidioides* var. *angustata* (Kirk) B. Nord. has elongated, sub-acute leaves with glandular hairs on the margins and upper lamina (Allan 1961). It is limited to the South island between 41°30' and 43° South, its type locality is also unknown (Kirk 1899, Allan 1961). *Brachyglottis bellidioides* var. *setosa* (Simpson & Thomson) B. Nord. has ovate or obovate leaves and is entirely clad in “setose” glandular hairs (Allan 1961). It is known only from its type locality of “sub-alpine scrub

and grassland on Mount Fyffe, Kaikoura Range” (Simpson and Thomson 1942, Allan 1961). *Brachyglottis bellidioides* var. *orbiculata* (Simpson & Thomson) B. Nord. has rugose (wrinkled), sub-orbicular leaves that are glabrous and up to six cm long (Allan 1961). It is known from its type locality of “grassland and sub-alpine scrub on the Garvie Mountains at 1200-1500m.” and its distribution is not established beyond the Garvie Mountains (Simpson and Thomson 1942, Allan 1961). *Brachyglottis bellidioides* var. *crassa* has thick rugose leaves with glandular hairs on the upper and lower lamina (Allan 1961). It is found on Stewart Island and its outlying islands, its type locality is Table Hill, Stewart Island (Simpson and Thomson 1943, Allan 1961). The variety *Senecio bellidioides* var. *glabratus* Kirk was accepted by Allan (1961) but was not accepted by Nordenstam (1978). It is described as (nearly) glabrous on both the upper and lower lamina (Allan 1961). It is distributed through the South Island above 44° south and its type locality is unknown (Kirk 1899, Allan 1961).

Brachyglottis traversii. *Brachyglottis traversii* was first described by Mueller (1863) as *Senecio traversii*. The species was distinguished from other rosette-*Brachyglottis* by near glabrous, elongated leaves that may be sparingly hispid (few glandular hairs) on the upper lamina and finely downy on the lower, and by prominent hydathoids between slight crenulations of the leaf margin. Allan (1961) described *B. traversii* with acute, elongated leaves up to 15cm long, four cm wide, and with few hispid (glandular) hairs. Allan (1961) describes lamina margins as in Mueller (1863). The scape is up to 30cm long, usually branched, covered in glandular and or eglandular hairs, and has large leafy bracts (Allan 1961). *Brachyglottis traversii* has the ambiguous type locality of “Alps near Nelson”, however, in later treatments of the species it was associated with stream sides of the Buller river catchment (Allan 1961). Despite being described in 1863, *B. traversii* was not included in the taxonomic accounts of Hooker (1864) and Kirk

(1899) (see Table 2.1). However, in 1867 Hooker listed *B. bellidioides* var. γ of Hooker (1864) as a synonym of *B. traversii* (Hooker 1867).

Senecio cochlearis (Simpson & Thomson) B. Nord. was identified from near the Ohikini river in the Buller catchment (Simpson and Thomson 1942). It was characterised by, nearly glabrous, long slender leaves (15 to 30cm long, two to four cm wide) with few bristles and prominent hydathodes; petioles slender, three to 20cm long, often purple, with purple bristles; and 20 to 30cm long branched scape with four to six heads (Simpson and Thomson 1942). *Senecio cochlearis* was synonymised with *B. traversii* by Allan (1961) as *S. traversii*.

Brachyglottis haastii. *Brachyglottis haastii* Hooker (1864) was primarily distinguished from other species by being covered everywhere with white, appressed, cottony wool (tomentum). It also has a tendency towards long slender petioles with elliptic to cordate-elliptic, crenate, leaves. Hooker (1864) does not describe branching of the scape or number of capitula in *B. haastii*. Allan (1961) described *B. haastii* with elliptic to sub-orbicular lamina up to 15cm long with cordate to truncate bases and obtuse tips. The petiole is often slender and long, up to 15cm (Allan 1961). The scape is up to 35cm, often slender and branched (Allan 1961). Both lamina surfaces, the petioles and scape are covered in a soft white, sub-appressed tomentum (Allan 1961). The upper lamina of older leaves is sometimes glabrous Allan 1961. *Brachyglottis haastii* is found throughout the South Island south of latitude 42° South and is generally found at higher elevations (Hooker 1864, Allan 1961). The type locality of *Brachyglottis haastii* is “Mount Cook, alt. 2700-4000 ft.” (Hooker 1864).

Brachyglottis southlandica. *Brachyglottis southlandica* was differentiated from *B. bellidioides* by its larger size, the (near) absence of “bristles” in most specimens, highly branched scape, and often purplish lower lamina (Cockayne

1914). *Brachyglottis southlandica* was distinguished from *B. lagopus* by its thinner leaves with few or no “bristles”, and the lack of glandular hairs on its scape (Cockayne 1914). Allan (1961) described *B. southlandica* with ovate-elliptic to sub-orbicular lamina up to 11cm long and cordate at the base with obtuse tips. Both the upper and lower lamina are often purplish and nearly glabrous with few “delicate” hairs (Allan 1961). The scape is up to 35cm, is usually branched and has both “delicate” and glandular hairs (Allan 1961). Allan (1961) describes the distribution of *Brachyglottis southlandica* as limited to the lower South Island below 45° 30' south, however, it is commonly described further north in central Otago. Its type locality is “neighbourhood of Balclutha, Gore, Tapanui, etc.” (Cockayne 1914). *Brachyglottis southlandica* var. *albidula* (Allan) B. Nord. is limited to the “Blue Mountains, Tapanui and neighbourhood” and its type locality is “Whisky Gully, Blue Mountains” (Allan 1961).

Table 2.1: Historical circumscriptions of rosette-*Brachyglottis*.

Hooker 1855	Hooker 1864	Kirk 1899	Allan 1961	Nordenstam 1978
<i>Senecio lagopus</i>	<i>Senecio lagopus</i>	<i>Senecio lagopus</i>	<i>Senecio lagopus</i>	<i>Brachyglottis lagopus</i>
<i>Senecio bellidioides</i>	<i>Senecio bellidioides</i>	<i>Senecio bellidioides</i>	<i>Senecio bellidioides</i>	<i>Brachyglottis bellidioides</i>
<i>Senecio saxifragoides</i>	–var. α	–var. <i>glabratus</i>	–var. <i>glabratus</i>	–var. <i>bellidioides</i>
	–var. β	–var. <i>angustatus</i>	–var. <i>angustatus</i>	–var. <i>angustata</i>
	–var. γ	<i>Senecio saxifragoides</i>	–var. <i>orbiculatus</i>	–var. <i>orbiculata</i>
<i>Senecio saxifragoides</i>	<i>Senecio saxifragoides</i>	<i>Senecio haastii</i>	–var. <i>setosus</i>	–var. <i>setosa</i>
<i>Senecio haastii</i>	<i>Senecio haastii</i>		–var. <i>crassus</i>	–var. <i>crassa</i>
			<i>Senecio saxifragoides</i>	<i>Brachyglottis saxifragoides</i>
			<i>Senecio haastii</i>	<i>Brachyglottis haastii</i>
			<i>Senecio traversii</i>	<i>Brachyglottis traversii</i>
			<i>Senecio southlandicus</i>	<i>Brachyglottis southlandica</i>
			–var. <i>albidulus</i>	–var. <i>southlandica</i>
				–var. <i>albidula</i>

2.3 Issues in Rosette-*Brachyglottis*

The rosette-*Brachyglottis* form a taxonomically problematic group because taxa overlap in both morphology and geographical distribution (Hooker 1864, Cockayne 1914, Wall 1918, Mennes et al. 2013). Uncertainties regarding the morphological distinction between rosette-species dates back to early in their taxonomic history (Hooker 1864, Cockayne 1914). Cockayne (1914) stated that “the classification of the whole series” (*Brachyglottis lagopus*, *B. bellidioides*, *B. saxifragoides*, *B. haastii* and *B. southlandicus*) “is in a most unsatisfactory position” and that he had seen many specimens that “are impossible to place with any degree of satisfaction”. In particular Cockayne (1914) highlighted the similarity of *B. lagopus* and *B. bellidioides*, and that *B. lagopus* and *B. saxifragoides* are often only distinguishable by the presence or absence of “bristles” on the upper lamina. However, Cockayne (1914) maintained that there are “well marked forms, which demand, at the least, varietal names”. Hooker (1864) also indicated issues in distinguishing between species. Hooker (1864) stated that although *B. lagopus*, *B. bellidioides* and *B. saxifragoides* were “most dissimilar in their usual states” they appear “to be united by intermediate forms”. Furthermore, Hooker (1864) noted that *B. haastii* “shows a tendency to pass into [*B.*] *bellidioides*” in populations near Lake Ohau, Canterbury.

Early descriptions of rosette-*Brachyglottis* species occurred when relatively few collections would have been available. Because of this, these descriptions did not account for the full variation of morphological characters in rosette-*Brachyglottis* species (Wall 1918). This can be seen when comparing the morphological descriptions of *B. lagopus*, *B. bellidioides* and *B. saxifragoides* in Hooker (1853) and Hooker (1864). In Hooker (1853) *B. bellidioides*, was distinguished from *B. lagopus* by its smaller leaves with glabrous lower lamina and smaller, unbranched scape. In Hookers 1864 publication, Hooker allowed for some overlap between *B. bellidioides*

an *B. lagopus* in leaf size and indicated that the scape of *B. bellidioides* could also be branched up to four times and this overlaps with the degree of branching of the scape in *B. lagopus* (two to eight times). Furthermore Hooker (1864) described several varieties of *S. bellidioides* (= *B. bellidioides*), of which var. β has a woolly lower lamina and tomentose scape similar to that described for *B. lagopus*. *Brachyglottis bellidioides* var. γ from Hooker (1864) was later synonymised with *S. traversii* (= *B. traversii*) (Hooker 1867). The morphological description of *B. saxifragoides* was also broadened by Hooker (1864) with a greater range of leaf size, scape size and scape branching (Wall 1918). Some small differences in capitulum morphology from Hooker (1853) were not included in Hooker (1864) with the capitula of *B. bellidioides* and *B. saxifragoides* being described as “as in [*B.*] *lagopus*” in the later. The broadening of morphological descriptions is likely the result of species initially being described when relatively few specimens were available and those descriptions being amended as more specimens became available (Wall 1918).

Hybridisation among rosette-*Brachyglottis* species may also explain the morphological overlap among species. Naturally occurring hybrids have been recorded between members of the monophyletic rosette-group and more distantly related, woody, *Brachyglottis* species (Drury 1973, Nordenstam 1978). The possibility of hybridisation among rosette-species is further supported by observations of intermediate forms between rosette-species, although these intermediates have never been formally recognised as hybrids (Hooker 1864, Wall 1918). The lack of recorded hybridisation within the rosette-*Brachyglottis* clade is possibly due to the difficulty in distinguishing hybrids from already similar species rather than an absence of hybridisation.

In addition to morphological overlap, *B. lagopus* and *B. saxifragoides* have an unusual geographical distribution (Wall 1918). *Brachyglottis saxifragoides* is only

known from the Port Hills of Western Banks Peninsula where as *Brachyglottis lagopus* is found throughout the lower North Island and upper-central South Island including eastern Banks Peninsula, but is absent from the Port Hills of Banks Peninsula (Wall 1918, Cockayne 1921, Allan 1961). This peculiar distribution pattern, coupled with overlapping morphology and habitat preference, was investigated by Wall (1918). Wall (1918) observed that, while some individuals of *B. saxifragoides* grow larger than any *B. lagopus*, there is significant overlap in all aspects of plant size between the two species. Furthermore he suggested that specimens of *B. lagopus* growing near Akaroa are generally smaller than those growing elsewhere on eastern Banks Peninsula, which may explain why *B. saxifragoides* was originally described as much larger than *B. lagopus*. Additionally the supposed differentiating bristles (glandular hairs) were present in both species, only differing in that they are often sparse to absent on the upper lamina of *B. saxifragoides* (Wall 1918). Glandular hairs were always present around the lamina margins and scapes of both species (Wall 1918). Based on these observations it was proposed that *B. lagopus* and *B. saxifragoides* be considered synonyms and any distinction between them be recognised below the level of species. It was further suggested that the remaining species of the rosette-*Brachyglottis* could also be synonymised with *B. lagopus* on the basis of common morphological characters including a woolly root-stock, hydathoides and bristles (glandular hairs) on lamina margin and surfaces (Wall 1918). Despite the conclusions of Wall (1918), more recent treatment of the rosette-group have continued to treat *B. lagopus* and *B. saxifragoides* as separate species (Allan 1961, Nordenstam 1978).

Mennes et al. (2013) further investigated the relationship between *B. lagopus* and *B. saxifragoides* using modern molecular techniques. They calculated genetic distance from AFLP data to study patterns of genetic similarity among *B. lagopus* (eastern Banks Peninsula), *B. saxifragoides* (Port Hills, western Banks Peninsula),

B. lagopus and other rosette species from the central South Island. They found that populations from eastern Banks Peninsula and the Port Hills formed separate clusters (Mennes et al. 2013). However, populations from eastern Banks Peninsula and the Port Hills were genetically more similar to one another than either was to populations from the central South Island (Mennes et al. 2013). This included central South Island plants identified as *B. lagopus* (Mennes et al. 2013). Based on these results Mennes et al. (2013) synonymised *B. saxifragoides* with *B. lagopus*. The results of Mennes et al. (2013) suggest that, throughout the central South Island, genetic distance between populations of rosette-*Brachyglottis* is correlated with geographical distance between populations. This can be seen in the genetic distance between plants from different locations that were all identified as *B. lagopus*. This is problematic as the currently described rosette-*Brachyglottis* species have overlapping ranges which implicitly hypothesises reproductive barriers other than geographic distance (Allan 1961). They suggested that the small but detectable genetic difference between *B. lagopus* from eastern Banks Peninsula and *B. saxifragoides* may be due to spacial separation. The results of Mennes et al. (2013) are congruent with those of Wall (1918) as both studies indicate that *B. saxifragoides* should be synonymised with *B. lagopus* and that the relationships between other rosette-*Brachyglottis* species require further investigation.

2.4 Summary of Rosette-*Brachyglottis*

The current treatments of rosette-*Brachyglottis* taxonomy is problematic for multiple reasons. It is likely that rosette-*Brachyglottis* species were named when relatively few specimens were available (Hooker 1853, 1864, Wall 1918). As new specimens were examined, morphological descriptions of rosette-*Brachyglottis* species have been broadened and new taxa recognised to accommodate the morphological variation (Hooker 1853, 1864, Kirk 1899, Allan 1961). This process has

resulted in a lack of clear morphological distinction between taxa (Cockayne 1914, Wall 1918). Because of broader species descriptions, many individual specimens cannot be reliably assigned to a species (Cockayne 1914). Taxonomic treatment of the rosette-*Brachyglottis* is further complicated by the possibility that species interbreed with one another. This possibility is indicated by observations of several authors despite the absence of formally recognised hybrids (Hooker 1864, Cockayne 1914). Most recently, molecular analysis has indicated that genetic distance among South Island rosette-*Brachyglottis* seems to be related to geographical location rather than taxonomical identification (Mennes et al. 2013). This may be an indication that geographical isolation is the primary form of reproductive barrier among rosette-*Brachyglottis*. Reproductive isolation by geographical causes alone is at odds with the current delimitation of rosette-*Brachyglottis* which includes overlapping species ranges (Allan 1961) and therefore implicitly hypothesises some form of symmetric reproductive barrier between species.

The above issues cast doubt on the accuracy of current species delimitation within the rosette-*Brachyglottis*. The currently recognised species and their geographical ranges may not accurately represent genetic relationships within the clade. The morphological variation used to distinguish between taxonomic groups may not accurately represent the underlying genetic variation of the clade. Further investigation of the genetic relationships within the rosette-*Brachyglottis* clade is required to resolve these issues.

2.5 Aims and Hypotheses

We conducted an investigation into the species delimitation of rosette-*Brachyglottis*. In this study we aimed to answer the question: does the current delimitation of rosette-*Brachyglottis* species accurately reflect its genetic structure? To answer this question, we tested the morphology-based taxonomic delimitation of Allan

(1961) by determining if morphological similarity is a good predictor of genetic similarity. This was tested against a null hypothesis of no relationship between morphological and genetic similarity, and geographical distance as the primary cause of genetic distance within rosette-*Brachyglottis*.

We also aimed to answer the related question: does the genetic structure in rosette-*Brachyglottis* indicate the presence of multiple biological species? The current delimitation of rosette-*Brachyglottis* recognises species with partially overlapping ranges and therefore, implicitly hypothesises a form reproductive isolation other than geographical distance. This was tested against a null hypothesis of rosette-*Brachyglottis* consisting of a single biological species and geographical distance as the primary cause of genetic distance within this species.

We used the biological species concept (BSC) as described by Coyne and Orr (2004) to determine what constitutes a ‘species’. Much has been written about species concepts and even long established and widely used concepts such as the BSC continue to be scrutinised (Mayr 1996, Noor 2002, Hey 2006, González-Forero 2009). The BSC as described by Coyne and Orr (2004) recognises sympatric populations as species if they exhibit “substantial but not necessarily complete reproductive isolation”. This version of the BSC is less strict than other versions that require complete reproductive isolation among sympatric ‘species’ (Coyne and Orr 2004).

We also used the genotypic cluster species concept (GCSC) of Mallet (1995) as a practical test of both morphological and genetic species. The GCSC is a species concept in which a species must be a distinguishable group of individuals with few or no intermediates when in contact with similarly defined clusters (Mallet 1995, Coyne and Orr 2004). The GCSC is similar in theory to the BSC but is defined in a much more practical manner that can be used to test species hypotheses with

morphological or genetic data (Mallet 1995, Coyne and Orr 2004).

Sexual reproduction via outcrossing is presumed to be the dominant type of reproduction in rosette-*Brachyglottis* (Mennes et al. 2013). The allowance of minimal interbreeding among ‘species’ is appropriate for analysis of rosette-*Brachyglottis* because there is established evidence of hybridisation between rosette-*Brachyglottis* and other woody-*Brachyglottis* species. Therefore, establishing that rosette-*Brachyglottis* ‘species’ are reproductively isolated in the strictest sense should involve cross-breeding experiments which are outside the scope of this investigation.

In our study we aimed to include representatives of all rosette-*Brachyglottis* species described by Allan (1961). As a result of this, *B. saxifragoides* was treated as a distinct species and not a synonym of *B. lagopus* (Mennes et al. 2013). Furthermore, [*B.*] *bellidioides* var. *glabratus* was treated as a distinct variety of *B. bellidioides* (Nordenstam 1978). In order to be certain that we accurately represented all species described by Allan (1961), we aimed to sample populations from the type localities of each species. We also aimed to include representatives from a large number of populations spread across the entire geographical range occupied by rosette-*Brachyglottis* and populations that exemplify the morphological diversity of rosette-*Brachyglottis*.

We used a ‘population genetics’ approach to investigate genetic structure in rosette-*Brachyglottis*. This included the use of model-based clustering and estimates of genetic distance as opposed to the heuristic tree searches of phylogenetics. A population genetics approach was favoured over a phylogenetic approach because we expected some level of reticulated gene flow among populations. This is because we aimed to use data from a number of populations that was much larger than the number of recognised rosette-*Brachyglottis* species and, therefore,

we expected gene-flow between some populations if not between species.

Chapter 3

Materials and Methods

3.1 Population Sampling

Sample populations were selected for three main reasons; to include historical type localities of recognised rosette-*Brachyglottis* taxa, to represent the morphological diversity of rosette-*Brachyglottis*, and to represent the geographical range of rosette-*Brachyglottis*. For each selected population we aimed to recover approximately eight samples for genetic analysis and one or more voucher specimens for morphological analysis. Voucher specimens were selected on the basis that they have the most commonly found characters of that population. When several voucher specimens were collected from a single population they were also selected to document the extent of morphological variation in that population. Samples for genetic analysis were also selected to represent the morphological range of the sampled populations.

3.2 Morphological Methods

3.2.1 Specimen Identification

Voucher specimens were identified using the taxonomic key of Allan (1961). Where possible, voucher specimens from the type locality of each species and variety were chosen to represent that taxon. For species with non-specific type localities, one or more vouchers were selected on the basis that they originated from the broadly circumscribed type locality and were identified as that species according to Allan (1961).

3.2.2 Character Morphology

An existing, unpublished, data set of morphological characters for 287 herbarium specimens was obtained from David G. Drury. Measurements for 18 qualitative characters and six quantitative characters were recorded in this data set (see table 3.1). Floral characters were not included in this data set because their observation would have destroyed the single capitulum of many herbarium specimens. Furthermore, the use of vegetative morphological characters is more practical when plants are not in flower.

The morphology of vouchers from sample populations was scored using the same morphological characters of David G. Drury data set as outlined in table 3.1. Care was taken to ensure that the scoring of characters for sample population vouchers matched the scoring of characters in herbarium specimens from the original data set. To this end many herbarium specimens from the original data set were re-scored to test and improve congruence in scoring character states. In an attempt to replicate the scoring of 16 herbarium specimens, there was an incongruence of 8.5% between original scores and repeats. Incongruent scoring was spread across multiple characters and no specific character(s) caused regular

incongruence.

3.2.3 Analysis of Morphological Data

A total of 354 herbarium specimens were represented in our morphological analysis. This included the 287 herbarium specimens scored by David G. Drury and a further 67 herbarium specimens specifically collected for this study from the populations that were sampled for the genetic analyses.

Pairwise morphometric distance between samples was calculated using Gower's dissimilarity coefficient (Gower 1971). Gower's dissimilarity coefficient was calculated for all 354 herbarium specimens. This only included samples with complete data for all morphological characters and excluded specimens that lacked a scape (i.e. were not in flower). Gower's dissimilarity coefficient has been extensively used for the analysis of morphological data (Crisp and Weston 1993, Ward 1993, Binns et al. 2002, Flann et al. 2008, Flann 2010, Rivero-Guerra 2011). The primary benefit of using Gower's dissimilarity coefficient is its ability to combine both qualitative (including asymmetrical and symmetrical binary data), multistate and quantitative data (Gower 1971, Crisp and Weston 1993, Binns et al. 2002). The use of multiple data types allows for a wider range of morphological characters to be incorporated into a single analysis and ordinations based on Gower's distance have been shown to be more accurate than other methods including principal components analysis (Crisp and Weston 1993). Gower's pairwise morphometric distances between individuals were calculated using R package CLUSTER (Maechler et al. 2014).

Characters 6 (branching of the scape) and 7 (secondary branching of the scape) are not independent from one another because primary branching of the scape must be present in order for secondary branching to be present. The use of a character that depends on the presence of another will add additional weight to the

first character (Pelser et al. 2004). This means that a single morphological feature measured by both characters will have more influence than other morphological features in a statistical analysis (Pelser et al. 2004). To resolve this issue characters six and seven were combined into a single multistate character prior to statistical analysis.

Characters 17 (Glandular hair presence on lower lamina) and 18 (Glandular hairs of lower lamina less than 1mm long) were excluded from the calculation of Gower's distances for two reasons. The primary reason for the exclusion of these characters was the difficulty in detecting glandular hairs on the lower lamina when in the presence of a dense tomentum (woolly or matted eglandular hairs). Glandular hairs can be small ($< 1\text{mm}$) and sparsely positioned on the lower lamina. In such cases, detection of glandular hairs typically requires the partial removal of tomentose hairs. Because glandular hairs may be positioned very sparsely, a thorough search for glandular hairs would ideally involve the complete removal of tomentose hairs. Therefore, a thorough search for glandular hairs on a tomentose lower lamina would incur significant damage to the specimen. Additionally character 18 (indication of glandular hair size) is dependent on character 17 (presence of glandular hairs). Thus these characters are not independent of one another and the use of both could unevenly weight a single morphological feature (Pelser et al. 2004). Examining glandular hairs on the upper lamina was not an issue because, if present, tomentum on the upper lamina was relatively thin and glandular hairs were therefore generally more pronounced.

Pairwise morphometric distances were explored using principal coordinates analysis (PCoA) (Gower 1966). PCoA is an ordination method derived from principal components analysis (PCA) (Gower 1966). Ordinations are used to detect and visualise the most informative dimensions of a multidimensional object (i.e. a set of points in multidimensional space) by identifying and displaying

the most informative dimensions (i.e. the dimensions that contain most of the variation among points). PCoA is an ordination method specifically designed for the exploration of pairwise distance matrices (Gower 1966). It is commonly used for the exploration of both morphometric and genetic distances. A PCoA was performed on the Gower's pairwise distance matrix using R package APE (Paradis et al. 2004).

3.3 Molecular Methods

3.3.1 DNA Extraction

DNA extraction mostly followed the method of Smissen et al. (2006) with the exception of the use of Zymo-Spin IIC columns and buffers in place of a Qiagen DNeasy plant mini kit. Tissue samples for DNA extraction were taken from a juvenile or youngest available leaf of each selected plant. DNA was extracted from fresh tissue or from tissue preserved on silica gel when sampling occurred several days before extraction. Approximately 200 μ g of tissue was used per sample extraction. CTAB buffer (Hexadecyltrimethylammonium bromide) (Doyle and Dickson 1987) was made by combining 36units of sterile water, 4units of 0.5M EDTA, 10units 1M TRIS ph 8.0, 20units 10% weight/volume CTAB, and 30units 5M NaCL. Tissue was ground in 1ml of warm CTAB buffer in a 1.5ml Eppendorf tube using a pestle. Ground tissue and CTAB buffer were incubated for 30min at 60°C.

CTAB extractions were followed by a chloroform:isoamyl alcohol extraction then a phenol:chloroform:isoamyl alcohol extraction and then a second chloroform:isoamyl alcohol extraction (Sambrook et al. 1989) as described below. Samples were centrifuged at 13200rpm for 5min and approximately 700 μ l was recovered. An equal volume of 24:1 chloroform:isoamyl alcohol was added and the

Table 3.1: Morphological characters from a data set of 288 herbarium specimens obtained from Dr. David G. Drury. P, A and NA respectively indicate presence, absence and not applicable.

Code	Character	States	Mode
Phyllary indumentum:			
1	Glandular hairs	2	P/A
2	Indumentum white to eye	2	P/A
Scape hairs:			
3	Red glandular	2	P/A
4	White glandular	2	P/A
Scape leaves:			
5	Glandular hairs	2	P/A
Inflorescence:			
6	Branched	2	P/A
7	Secondary branching	2	P/A
Rosette leaf lamina:			
8	Widest in lower half	2	P/A
9	Auricled (Cordate)	2	P/A
Lamina margin:			
10	Margin crenate	2	P/A
Upper lamina:			
11	Green or green showing through indumentum	2	P/A
12	Eglandular hairs less than 1.5mm	2	P/A
13	Glandular hairs scattered over lamina	2	P/A
14	Glandular hairs crowded at margins	2	P/A
Lower lamina:			
15	Green or green showing through indumentum	2	P/A
16	Eglandular hairs less than 1.5mm	2	P/A
17	Glandular hairs present (excluding median vein)	3	P/A/NA
18	Glandular hairs less than 1mm long	3	P/A/NA
Numerical characters:			
19	Petiole length	NA	Numerical
20	Lamina length	NA	Numerical
21	Lamina width	NA	Numerical
22	Ratio: petiole length/lamina length	NA	Numerical
23	Ratio: lamina length/lamina width	NA	Numerical
24	Ratio: petiole length/lamina width	NA	Numerical

samples were then centrifuged at 13200rpm for 5min and approximately 550 μ l was recovered from the upper layer per sample. Subsequently an equal volume of phenol:chloroform:isoamyl alcohol (buffer saturated phenol pH 8.0) was added and the samples were then centrifuged at 13200rpm for 5min and approximately 450 μ l

was recovered from the upper layer per sample. This was followed by adding an equal volume of 24:1 chloroform:isoamyl alcohol the samples were then centrifuged at 13200rpm for 5min and approximately 350 μ l was recovered from the upper layer per sample.

DNA was then recovered using a Zymo-Spin IIC column and buffers. The DNA sample was added to twice its volume of DNA binding buffer and spun through a Zymo column in centrifuge at 8000rpm for 1min. 500 μ l of wash buffer was added to the column and spun down in centrifuge at 8000rpm for 1min. a further 500 μ l of wash buffer was added to the column and spun down in centrifuge at 13000rpm for 2min. DNA was then collected in 50 μ l of 10 μ M Tris 1mM EDTA pH 8.0 which was spun through the Zymo column at 8000rpm for 1min.

3.3.2 Marker Amplification and Genotyping

All markers used in this study were designed by Rob Smissen (Landcare Research) for use in rosette-*Brachyglottis*. Markers Pgic-12 and Smht-04 are low copy number nuclear gene introns, and markers Bra-06 to Bra-26 are microsatellite loci. Because a combination of low copy number nuclear gene introns and microsatellite loci were used they are collectively referred to as ‘markers’. Each pair of primers used to amplify a marker were designed with one of each pair incorporating a 5’ tail matching the M13 Forward primer sequence (TGTAACGACGCGCCAGT) to allow incorporation of the fluorescent labels FAM and HEX (Boutin-Ganache et al. 2001)

For each sample, Markers were amplified using the nine primer combinations shown in table 3.2. 2 μ l of a 10% dilution of sample DNA was used per PCR reaction per sample. Sample DNA was added to a solution of 2 μ l of 10 \times PCR buffer including 15 μ M MgCl₂, 0.3 μ l of dNTP (concentration 10 μ M each

nucleotide), 0.4 μ l of 10 μ M forwards primer, 0.8 μ l of 10 μ M reverse primer, 0.2 μ l of Roche FastStart taq, and 13.5 μ l of water.

Table 3.2: Primer combinations for genetic markers.

Marker	Direction	Primer
Pgic-12	Foward	TGTAAAACGACGGCCAGTGCAATACGGTTTTTTCAGTCG
	Reverse	ATGCTTCTAGCACCTTTTAGG
Shmt-4	Foward	TGTAAAACGACGGCCAGTTATGGTGGAAACGAGTAAGCA
	Reverse	GCACGCTTTTGACACAAGAG
Bra-6	Foward	TGTAAAACGACGGCCAGTAAACACATTGGAAGCCACC
	Reverse	CCCAAAGGCCGTGCAATAC
Bra-9	Foward	TGTAAAACGACGGCCAGTACCACTGCAAGTTGGGAGC
	Reverse	ACAACAATGGGTTACCTCG
Bra-12	Foward	TGTAAAACGACGGCCAGTGCCGATTGCCTCAACAGTC
	Reverse	AGCCCGGTTTCAGGTGTATC
Bra-19	Foward	TGTAAAACGACGGCCAGTTCCTTCAGAATCACCGCCTC
	Reverse	AGGAATGAACGGTCAAAGCC
Bra-21	Foward	TGTAAAACGACGGCCAGTGATTCCTCAAAGTGTGGCCG
	Reverse	CAGACATCACGACGGTTGC
Bra-25	Foward	TGTAAAACGACGGCCAGTCCTCAACTTCATAGTCAAATCCCG
	Reverse	CTCCTGAAATGACTGCCCC
Bra-26	Foward	TGTAAAACGACGGCCAGTTGAACTGGGTGGAACGAGG
	Reverse	ACAATGGAGTGTTCACCCG

Samples were initially denatured at 95°C for 2min. This was followed by fifteen cycles of 94°C for 30s, 60°C for 30 s, and 72°C for 1min. This was followed by twenty cycles of 94°C for 30s, 49°C for 30 s, and 72°C for 1min. Samples went through a final, single cycle of 72°C for 30min and were then cooled to 4°C. PCR product was genotyped at the University of Canterbury using a ABI 3130xl Genetic Analyzer with GeneScan-500 LIZ size standard.

3.3.3 Scoring Genetic Markers

Single copy Microsatellite regions are usually selected for genetic analyses because they can be scored as co-dominant markers (i.e frequency of all alleles can be recorded) (Kalia et al. 2011). However, few co-dominant markers could be found for the rosette-*Brachyglottis*. Polyploidy is common in the Asteraceae and it is

possible that the chromosome number of *Brachyglottis* ($2n = 60$) is an indication of ancestral polyploidy (Beuzenberg 1975, Barker et al. 2008, Smissen et al. 2011, Soto-Trejo et al. 2013). It is possible that the relatively low number of single copy loci found in rosette-*Brachyglottis* is a result of duplicated loci due to ancestral polyploidy. Analysis of ancestral polyploidy in Asteraceae has shown that the loss or retention of duplicated genes is not random (Barker et al. 2008). Therefore, the selection of only co-dominant markers would have been impractical and could potentially introduce bias to the data by favouring genetic regions in a non-random fashion.

Instead, markers that produced six or less alleles per individual were selected and they were scored as dominant markers (i.e. allele presence can be recorded, but not allele frequency) and the data was treated in a manner similar to AFLP data (Bonin et al. 2007). Markers that produced greater numbers of alleles were excluded to avoid weighting the data set towards some loci over others and to minimise error in scoring the alleles. The possibility of scoring a subset of the markers as co-dominant was revisited once the results of marker amplification were known, but only three of nine markers produced alleles in a manner that was consistent with single copy markers and this was considered too few to pursue. Alleles of all nine markers were scored manually in Gene Marker version 2.4.2.

3.4 Analysis of Genetic Markers

3.4.1 Bayesian Inference of Genetic Groups

A model-based Bayesian inference cluster analysis was performed on the genetic data using STRUCTURE, version 2.3.3 (Falush et al. 2007). Using genetic data, STRUCTURE aims to cluster individuals into a number (K) of hypothetical populations (referred to as genetic clusters for clarity) (Pritchard et al. 2000,

Falush et al. 2003). To this end STRUCTURE needs to estimate the probability of membership of each individual to each genetic cluster (Pritchard et al. 2000, Falush et al. 2003). This is achieved with a Markov chain Monte Carlo (MCMC) algorithm which searches parameter space and proposes K genetic clusters, each with candidate allele frequencies constructed from the genetic data (Pritchard et al. 2000, Falush et al. 2003). The probability of membership of each individual to each genetic cluster is calculated based on the proposed allele frequencies of genetic clusters and the ‘real’ allele frequencies of each individual (Pritchard et al. 2000, Falush et al. 2003). This process is repeated many times by the MCMC which selectively moves through parameter space towards areas of higher probability. Because the MCMC starts at an arbitrary point in parameter space, the earlier steps of the MCMC are discarded (the ‘burn in’ period) and the posterior probability of K is estimated from the later steps (Pritchard et al. 2000, 2010). This process must be repeated for each value of K in order to estimate the ‘best’ value of K (Pritchard et al. 2000, 2010).

One of the more difficult aspects of an analysis in STRUCTURE is determining the ‘best’ value for K (Pritchard et al. 2000, Evanno et al. 2005). The highest estimated posterior probability of K , $L(K)$, will not always indicate the best value of K in part because it is an estimate of the true $L(K)$ (Pritchard et al. 2000, Evanno et al. 2005). Furthermore, $L(K)$ will often continue to gradually increase or plateau with increased K at the same time as the variation of $L(K)$ increases between runs of the same K (i.e. the estimated of $L(K)$ becomes less reliable) (Evanno et al. 2005). One approach to choosing the best value of K is to pick the value at which $L(K)$ begins to plateau and variance between runs increases. Evanno et al. (2005) proposed a second method in which the best value of K is chosen by the highest value of ΔK . ΔK is a measure of the rate in change of the likely hood function for K and the highest value has been used to indicate high

level genetic clustering (Evanno et al. 2005).

Analysis of the rosette-*Brachyglottis* in STRUCTURE used genetic data for all 273 individuals including 253 individuals that each had complete data for all nine markers and 20 individuals with missing data for one to two loci. This analysis used the admixture model with recessive alleles and correlated allele frequencies among populations. The admixture model allows (admixed) individuals to be partially assigned to multiple genetic clusters (Pritchard et al. 2000). Admixed individuals may be interpreted as evidence of mixed ancestry due to immigration and or interbreeding (Pritchard et al. 2000). The analysis was run multiple times with $K = 1:15$ to determine the ‘best’ value of K . The analysis was run ten times for each of the fifteen K values and each run had a burn in period of 20,000 replicates followed by another 80,000 replicates. STRUCTURE HARVESTER (Earl and vonHoldt 2012) was used to summarise the output from STRUCTURE and aid the selection of an optimal K value. Selection of the optimal value of K was based on two criteria. The first of these criteria was the method of Evanno et al. (2005) which chooses the optimal K value by the highest value of ΔK . The second criteria was the $L(K)$ method of Pritchard et al. (2000). Replicate runs for the optimal value of K were merged in CLUMPP (Jakobsson and Rosenberg 2007) using the Greedy algorithm with 1000 replicates. The merged results were visualised in R (R Development Core Team 2008).

3.4.2 Pairwise Genetic Distance

Jaccard distances were calculated among the 253 individuals with complete data sets across all nine markers using the R package ADE4 (Dray and Dufour 2007). The Jaccard dissimilarity coefficient is commonly used for the analysis of dominant markers (Bonin et al. 2007, Meudt and Clarke 2007, Burnier et al. 2009, Galbany-Casals et al. 2012, Medrano et al. 2014). Amplification of a single genetic

marker over multiple individuals can produce alleles of many sizes (Meudt and Clarke 2007). However, relatively few alleles are produced for each individual (see tabel 3.2). Therefore, individuals will often have relatively few ‘present’ alleles and many ‘absent’ alleles (Bonin et al. 2007, Meudt and Clarke 2007). Furthermore, the shared absence of a allele between two individuals is less informative than shared presence because shared absence may be the result of unrelated events (e.g the evolution of a novel allele in a third individual will result in an additional shared absence between the first two individuals). This results in a high frequency of relatively uninformative shared absent alleles among individuals. A key characteristic of the Jaccard dissimilarity coefficient is that it emphasises shared presence of characters between individuals and not shared absence (Bonin et al. 2007). By emphasising shared present of alleles the Jaccard dissimilarity coefficient maximises the informative signal in the data (Bonin et al. 2007, Meudt and Clarke 2007). The Jaccard distance matrix of 253 individuals was visualised using PCoA in R package APE (Paradis et al. 2004).

Genetic distance between populations was quantified using Nei’s corrected genetic distance (Nei 1972, 1978). Nei’s genetic distance also emphasises shared presence of characters over shared absence of characters (Bonin et al. 2007, Meudt and Clarke 2007). As with Jaccard distance, this is beneficial in the analysis of dominant markers because it minimises the less informative signal of shared absence of an allele (Bonin et al. 2007, Meudt and Clarke 2007). Nei’s genetic distance assumes that differences in allele frequencies among populations arise from a combination of mutation and genetic drift (Deza and Deza 2009). Genetic distance among populations is calculated by the pairwise degree of overlap in population allele frequencies (Nei 1972, 1978, Goldstein and Pollock 1997).

Large numbers of loci have have been shown to increase the reliability of distances derived from Nei’s coefficient (Bonin et al. 2007, Simmons et al. 2007) and few

representatives per population are required for accurate estimation of distance when large numbers of characters are used (Nei 1978). Nei's genetic distance is effective at reconstructing genetic relationships between recently diverged or diverging groups and is less accurate when the time since divergence is large (Goldstein and Pollock 1997). Accuracy is lost after long divergence times due to minimal overlap in allele frequencies between populations which leads to 'flat' distances (Goldstein and Pollock 1997). Nei's genetic distance was calculated for all 46 populations in R package GSTUDIO (Dyer 2014) using the 253 individuals with complete data for all nine markers.

A Neighbor-Net analysis was performed on the matrix of pairwise genetic distances among populations calculated using Nei's coefficient. Neighbor-Net analysis (Bryant and Moulton 2004) is a form of distance based, network analyses that is based the Neighbor-Joining algorithm of Saitou and Nei (1987). The use of a network to display distance data has several advantages over tree based methods. Primarily network analyses like Neighbor-Net allow for conflicting genetic signals to be displayed and interpreted as complex relationships rather than forcing the data to conform to a bifurcating pattern of genetic similarities (Bryant and Moulton 2004). In Neighbor-Net analyses, reticulated evolutionary relationships are represented by multiple internal paths between terminal nodes (Bryant and Moulton 2004, Huson and Bryant 2006). This flexibility means that the assumption of a single evolutionary history for all genetic characters is not required in a Neighbor-Net analysis (Bryant and Moulton 2004, Huson and Bryant 2006, Huson and Scornavacca 2011). The interpretation of reticulated evolution is of particular relevance when investigating systems of taxa that may not have fully diverged from one another (Bryant and Moulton 2004). For example, Neighbor-Net analyses have been used to successfully identify hybrids in several systems (Smitsen and Heenan 2007, Galbany-Casals et al. 2012). The ability

to identify hybrids is relevant to our study because little is known about the potential for hybridisation among rosette-*Brachyglottis* species and hybridisation has been identified between rosette-*Brachyglottis* and woody-*Brachyglottis* species (Drury 1973). Neighbor-Net is less conservative than other network methods and does not lose resolution with large data sets (Huson and Bryant 2006, Huson and Scornavacca 2011). Neighbor-Net analysis was performed using the Splits Tree software package, version 4.13.1 (Huson and Bryant 2006).

3.4.3 Isolation by Distance Analysis

Analyses to quantify the relationship between geographical and genetic distance were performed at the population level using Nei's genetic distance and euclidean geographical distance between populations. Euclidean distance was calculated between the geographical locations of the 46 populations in the R package STATS (R Development Core Team 2008). Correlation between geographical and genetic distance was assessed with a Mantel test using R the package ADE4 (Dray and Dufour 2007). Simple linear regression of pairwise geographical and genetic distances was performed in R to visualise the geographic-genetic relationship.

Chapter 4

Results

4.1 Population Sampling

4.1.1 Population Sampling

A total of 46 populations were represented in this study. This included five North Island, 39 South Island and two Stewart Island populations shown in table 4.1 and mapped in figure 4.1. The most Northerly population was population 1 from the Kaweka Range, Hawke's Bay at latitude -39.347312. The most Southerly population was population 46 from Table Hill, Stewart Island at latitude -47.039826. The altitude of sampled populations varied from approximately 10m at coastal Charleston (population 17) to approximately 1700m at the Remarkables Ski Field (population 35). Sample populations occupied a wide variety of habitats including coastal scrub and grasslands, alpine scrub and grasslands, sub-alpine forest floors, river banks and exposed rocky bluffs.

Table 4.1: Summary sampled populations.

Pop	Location	Latitude	Longitude	Elevation
1	Smith Russell Track, Kaweka Range	-39.347312	176.326896	1300m
2	Rangiwahia Track, Whanahiu Range	-39.897108	176.013692	1000m
3	Herepai Ridge, Tararua Range	-40.689034	175.519681	1000m
4	Rimutaka Hill, Rimutaka Range	-41.118353	175.229702	650m
5	Red Rocks Track, Wellington	-41.349782	174.725086	220m
6	Mt Fyffe, Seaward Kaikoura Range	-42.322656	173.595392	1150m
7	Balaclava Ridge, Crimea Range	-42.168333	172.821667	1200m
8	Mt Riley, Richmond Range	-41.410549	173.696743	1220m
9	Red Hills Plateau, Richmond Range	-41.726327	172.998927	1020m
10	Salisbury Bridge, Aorere River	-40.802292	172.534129	60m
11	Mt Arthur, Arthur Range	-41.21929	172.690159	1650m
12	Ghost Valley, Marino Mountains	-41.525	172.558333	1350m
13	Blackwater River, Buller River	-41.854753	171.795483	50m
14	St James Walkway, Lewis Pass	-42.381667	172.401667	860m
15	Denniston Plateau, West Coast	-41.76037	171.784212	680m
16	Coastal Charleston, West Coast	-41.906078	171.428304	10m
17	Sewell Peak, West Coast	-42.409	171.341667	800m
18	Otira Valley, Arthurs Pass	-42.898599	171.555943	900m
19	Bealey Valley, Arthurs Pass	-42.916338	171.555504	850m
20	Mt Bealey, Arthurs Pass	-42.961416	171.55808	1550m
21	Cass Field Station	-43.034832	171.751044	580m
22	Cave Stream, Castle Hill	-43.200358	171.74592	720m
23	Porters Pass, Torlesse Range	-43.296564	171.74007	950m
24	Mt Somers, Canterbury	-43.618262	171.305647	750m
25	Pareora River Reserve, Hunters Hills	-44.351854	170.834354	360m
26	North Branch, Godley River	-43.59686	170.509214	1020m
27	Sugar Loaf, Banks Peninsula	-43.605	172.65	400m
28	Walls Track, Banks Peninsula	-43.67644	172.625144	300m
29	Otepatotu Reserve, Banks Peninsula	-43.746545	173.016558	750m
30	Lighthouse Road, Banks Peninsula	-43.85	172.966667	600m
31	Maitland Stream, Lake Ohau	-44.234143	169.725431	1100m
32	Red Tarns, Mount Cook	-43.749302	170.101461	1150m
33	Lindis Pass	-44.579623	169.634408	850
34	Ben Lomond, Queenstown	-45.020273	168.637515	850m
35	Remarkables Ski Field	-45.050152	168.819871	1700m
36	Wye Creek, The Remarkables	-45.137733	168.770167	700m
37	Key Summit, Livingstone Mountains	-44.831039	168.126097	1050m
38	Mount Burns, Fiordland	-45.749463	167.402193	1550m
39	West Dome, Mossburn	-45.605031	168.241453	650m
40	Whiskey Gully, Blue Mountains	-45.94797	169.316313	600m
41	Black Gully, Blue Mountains	-45.90103	169.370386	750m
42	Maungatua Route, Maungatua	-45.857649	170.14565	650m
43	Flagstaff, Dunedin	-45.834482	170.458948	600m
44	Dunstan Road, Rock & Pillar Range	-45.528495	169.978274	1000m
45	Mount Rakeahua, Stewart Island	-46.944842	167.883968	620m
46	Table Hill, Stewart Island	-47.039826	167.841884	710m

‘Populations’ of rosette-*Brachyglottis* can vary in size dramatically. Large populations include countless individuals and have a continuous distribution that can expand across large areas of suitable habitat. This is commonly the case in Alpine areas of the South Island where rosette-*Brachyglottis* often grow continuously throughout the sub-alpine zone, extending along mountain ranges. When sampling very large populations the sampling area was restricted to an area of approximately 3km². The smallest sample *Brachyglottis* populations have a few dozen to a few hundred individuals. Examples of small populations include stream side locations like Aorere river, Golden Bay (Population 10) and isolated mountains with minimal alpine zone like Mount Riley, Marlborough (population 9 in figure 4.1).

The most dense sampling occurred in Arthur’s Pass, Canterbury (populations 18, 19 and 20 in figure 4.1). Sampling was particularly thorough in this area because preliminary data indicated pronounced variation in genetic structure between populations west and east of Arthur’s Pass. Pronounced variation was also indicated in this area by the results of Mennes et al. (2013).

4.1.2 Representation of Species

Populations of all six rosette-*Brachyglottis* species recognised by Allan (1961) were represented by populations in our analysis. Specific attention was paid to the populations that most accurately represent the type locality of each species and populations that are otherwise significant in representing the historically recognised range of each species. Some samples of *Brachyglottis lagopus*, *B. saxifragoides* and *B. haastii* were collected from populations from at or very near to their type localities. *Brachyglottis bellidioides*, *B. traversii* and *B. southlandica* have type localities that were less precisely defined (Allan 1961) and the samples taken from these species included those taken from several populations in the

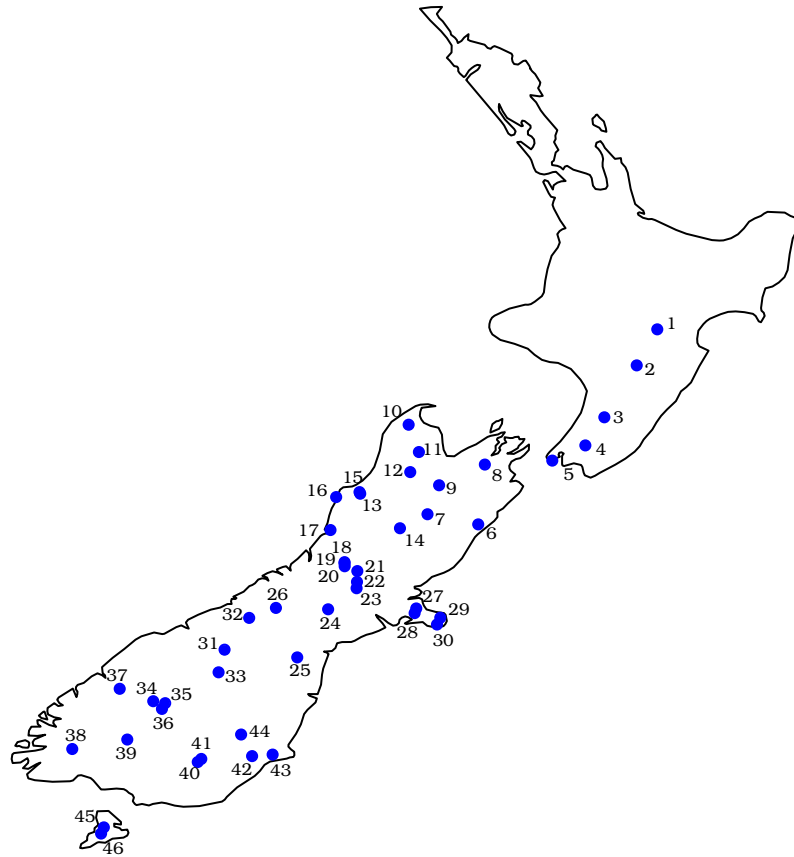


Figure 4.1: Locations of the 46 *Brachyglottis* populations represented in the genetic analyses. Numbers identify individual populations as indicated in Table 4.1.

general area that was specified as the type locality. The sampled plants were identified using the key of Allan (1961) (see table 4.2). Plants from populations in the vicinity of the type locality of a rosette-*Brachyglottis* species or otherwise of significant relevance to a species are summarised below.

Table 4.2: Summary of voucher identification by population. Uncertainty is indicated by ‘?’ or multiple species names.

Pop.	Location	Taxa ID of vouchers
1	Smith Russell Track, Kaweka Range	<i>B. lagopus</i> ×3
2	Rangiwahia Track, Whanahiua Range	<i>B. lagopus</i>
3	Herepai Ridge, Tararua Range	<i>B. lagopus</i>
4	Rimutaka Hill, Rimutaka Range	<i>B. lagopus</i>
5	Red Rocks Track, Wellington	<i>B. lagopus</i> ×2
6	Mt Fyffe, Seaward Kaikoura Range	<i>B. saxifragoides</i> > <i>lagopus</i>
7	Balaclava Ridge, Crimea Range	<i>B. bellidioides</i> ×6
8	Mt Riley, Richmond Range	<i>B. southlandica</i> ?
9	Red Hills Plateau, Richmond Range	<i>B. lagopus</i>
10	Salisbury Bridge, Aorere River	<i>B. traversii</i>
11	Mt Arthur, Arthur Range	<i>B. lagopus</i> ?
12	Ghost Valley, Marino Mountains	<i>B. bellidioides</i> ?×2, unknown×2
13	Blackwater River, Buller River	<i>B. traversii</i>
14	St James Walkway, Lewis Pass	<i>B. bellidioides</i> var. <i>angustata</i>
15	Denniston Plateau, West Coast	NA
16	Coastal Charleston, West Coast	<i>B. bellidioides</i> ×2
17	Sewell Peak, West Coast	NA
18	Otira Valley, Arthurs Pass	<i>B. bellidioides</i> var. <i>glabrata</i> ×3
19	Bealey Valley, Arthurs Pass	<i>B. bellidioides</i> var. <i>angustata</i> ×5
20	Mt Bealey, Arthurs Pass	<i>B. bellidioides</i> ×4
21	Cass Field Station	<i>B. bellidioides</i>
22	Cave Stream, Castle Hill	
23	Porters Pass, Torlesse Range	<i>B. bellidioides</i> > <i>lagopus</i>
24	Mt Somers, Canterbury	
25	Pareora River Reserve, Hunters Hills	<i>B. southlandica</i>
26	North Branch, Godley River	<i>B. bellidioides</i> ?×4
27	Sugar Loaf, Banks Peninsula	<i>B. saxifragoides</i>
28	Walls Track, Banks Peninsula	<i>B. lagopus</i>
29	Otepatotu Reserve, Banks Peninsula	<i>B. lagopus</i> ×2
30	Lighthouse Road, Banks Peninsula	<i>B. lagopus</i>
31	Maitland Stream, Lake Ohau	NA
32	Red Tarns, Mount Cook	<i>B. haastii</i> ×3
33	Lindis Pass	NA
34	Ben Lomond, Queenstown	<i>B. southlandica</i>
35	Remarkables Ski Field	<i>B. bellidioides</i> > <i>southlandica</i>
36	Wye Creek, The Remarkables	NA
37	Key Summit, Livingstone Mountains	<i>B. bellidioides</i>
38	Mount Burns, Fiordland	<i>B. bellidioides</i> ×2
39	West Dome, Mossburn	<i>B. bellidioides</i>
40	Whiskey Gully, Blue Mountains	<i>B. southlandica</i> var. <i>albidula</i>
41	Black Gully, Blue Mountains	<i>B. southlandica</i> var. <i>albidula</i>
42	Maungatua Route, Maungatua	<i>B. bellidioides</i> - <i>southlandica</i>
43	Flagstaff, Dunedin	<i>B. southlandica</i> - <i>bellidioides</i>
44	Dunstan Road, Rock & Pillar Range	<i>B. bellidioides</i> > <i>southlandica</i>
45	Mount Rakeahua, Stewart Island	<i>B. bellidioides</i> var. <i>crassa</i>
46	Table Hill, Stewart Island	<i>B. bellidioides</i> var. <i>crassa</i>

Brachyglottis lagopus. *Brachyglottis lagopus* was represented by population 29 from near Otepatotu Reserve, Banks Peninsula. This population is located less than ten kilometres north-east of Akaroa at an elevation of approximately 750m which matches the type locality of “Akaroa in montosis” described by Raoul (1844). The two vouchers taken from this population were both identified as *B. lagopus* using the key of Allan (1961). Both had branching scapes with red glandular hairs, spreading broad-elliptic leaves ranging from cordate to truncate at the base with entire to crenate margins, lower lamina were woolly with tomentum and upper lamina covered with glandular hairs. The largest leaves were 11.7cm long (a 4cm petiole and 7.7cm lamina) by 4cm wide and 13.7cm (a 4cm petiole and 9.7cm lamina) by 7cm wide. These sizes slightly exceed the upper limit of leaf size described by Hooker (1853) (four inches or slightly over ten centimetres) but is well within the upper limit of Allan (1961) (10cm petiole and 15cm lamina).

Population 30 from near Lighthouse Road, Banks peninsular was also considered to be an accurate representation of *B. lagopus*. This population is located approximately five kilometres south of Akaroa at an elevation of approximately 600m. A single voucher was taken from this location. It had broad-elliptic leaves with crenate margins and cordate bases, the lower lamina was tomentose and upper lamina covered in glandular hairs. The largest leaf was 22cm long (10.3cm petiole and 11.7cm lamina) and 7.4cm wide. The scape had secondary branching and was covered in a mixture of silky and red glandular hairs.

All voucher specimens from populations 1-5 in the North Island were identified as *B. lagopus*. Vouchers from the North Island usually had large, cordate, broad-elliptic leaves with a densely tomentose lower lamina and long glandular hairs covering the upper lamina. The ‘silky’ eglandular hairs of the upper lamina described by Allan (1961) were only found in younger leaves. Scapes were highly branched and covered in a combination of red glandular and eglandular hairs.

Despite their geographical distance from the type locality of *B. lagopus*, the identification of these populations is significant because *B. lagopus* is the only rosette-*Brachyglottis* species recognised from the North Island (Allan 1961).

Brachyglottis saxifragoides. *Brachyglottis saxifragoides* was represented by population 27 from the Sugar Loaf, Port Hills, Banks Peninsula. This population is approximately five kilometres west of the type locality of “Port Cooper” (Lyttelton) given by Hooker (1853), and at an elevation of approximately 400m. The single voucher taken from this location was identified as *B. saxifragoides* according to Allan (1961). It had broad-elliptic leaves with crenate margins and cordate bases, the lower lamina was covered in a woolly tomentum and upper lamina was almost glabrous with some glandular hairs around the margin. The silky hairs described by Allan (1961) were not present on the upper lamina of the largest leaf. The leaf lamina was 12.8cm long by 9cm wide and on a 8.1cm petiole. The scape was branched and covered in a mixture of red glandular hairs and silky, almost tomentose, eglandular hairs.

Brachyglottis haastii. *Brachyglottis haastii* was represented by population 32 from Red Tarns, Mount Cook, Canterbury at approximately 1150m elevation. The type locality of *B. haastii* is the somewhat ambiguous “Mount Cook, alt. 2700-4000ft.” (820-1220m) (Hooker 1853). It is unclear as to whether Hooker (1853) was referring to Aoraki/Mount Cook itself or the general area known as ‘Mount Cook’. The Hooker Valley extends from north to south between Aoraki/Mount Cook and Mount Cook village. No rosette-*Brachyglottis* plants could be found during an extensive search of the slopes east of the Hooker Lake, Hooker Valley, at the elevations specified by Hooker (1853). The Red Tarns area is one kilometre south of Mount Cook village and 18km south of the summit of Aoraki/Mount Cook. All three vouchers recovered from the the Red Tarns population were identified as *B. haastii* using the key of Allan (1961). Each of these vouchers were clad everywhere

in a white, sub-appressed tomentum with the exception of some older leaves where the tomentum had peeled away from the upper lamina leaving a glabrous surface (see figure 4.2). Scapes were branched (secondary branching in two vouchers) and covered with white glandular hairs in addition to eglandular hairs. Leaf shape was sub-orbicular to elliptic with obtuse tips, cordate to truncate bases, and entire to crenate lamina margins. The largest leaf of each voucher had a petiole length of 5.5-8.5cm and a lamina length of 4.2-10cm. All of these measurements fall within the ranges given by Allan (1961) (petiole and lamina 3-15cm each).



Figure 4.2: Upper lamina of *B. haastii* from Red Tarns, Mount Cook.

The single voucher specimen of population 22 from Cave Stream near Castle Hill, central Canterbury was also identified as *B. haastii*. The leaves of this voucher specimen were small (largest leaf 2.7cm by 1.9cm lamina on 1.7cm petiole), crenate, nearly appressed with a sub-appressed white tomentum covering all surfaces. The scape was unbranched and covered in white tomentose hairs. *B. haastii* has

been identified in the area around Castle Hill from an early period (Kirk 1899). Because other rosette-*Brachyglottis* populations from central Canterbury have been identified as other species, this population represents a potential overlap in species ranges.

Brachyglottis bellidioides. *Brachyglottis bellidioides* is the most morphologically diverse rosette-species described in Allan (1961) and multiple varieties are recognised. Several populations were selected as potential representatives of *B. bellidioides* due to its broad type locality of “Canterbury” (Hooker 1853). These included populations 18-20 from Arthurs Pass, population 21 from Cass and population 23 from Porters Pass. There was morphological variation within and among these populations, however all were identified as *B. bellidioides*.

***Brachyglottis bellidioides* var. *bellidioides*.** All four vouchers from population 20 on Mt Bealey, Arthurs Pass were identified as *B. bellidioides* var. *bellidioides* (‘typical’ *B. bellidioides* in Allan (1961)). These vouchers had small appressed to spreading, sub-orbicular to broad-elliptic leaves (see figure 4.3). The upper lamina was usually covered in glandular hairs, but ranged to (near) glabrous. Lower lamina was glabrous to thinly tomentose. Scapes were usually unbranched.

The single voucher taken from Porters Pass had small appressed truncate leaves growing from a root stock with silky brown hairs. The lower lamina was tomentose and the upper lamina was covered with glandular hairs and some silky eglandular hairs. This voucher was identified as *B. bellidioides* however it was approaching a small form of *B. lagopus*.

Vouchers from population 26 near the Godley River, Canterbury were tentatively identified as *B. bellidioides*. These vouchers did not key well due to the lack of glandular hairs on either lamina and the common presence of eglandular hairs on



Figure 4.3: Rosette-leaves of *B. bellidioides* from Mount Bealey, Arthurs Pass.

both lamina surfaces that approached a tomentum. Plants from this population commonly exceeded the size restrictions placed on *B. bellidioides* by Allan (1961). For these reasons, population 26 was not considered to be typical *B. bellidioides*.

[*Brachyglottis*] *bellidioides* var. *glabratus*. Vouchers from population 18 in Otira Valley, Arthurs pass had small appressed-spreading, truncate leaves that were often glabrous on both surfaces (see figure 4.4). These vouchers were identified as *B. bellidioides* in its glabrous form (*S. bellidioides* var. *glabratus* in Allan (1961)). Other individuals in the area were approaching *B. bellidioides* var. *angustata*.

The single voucher taken from Cass, central Canterbury also had small crenate, truncate leaves with glabrous lower lamina and near glabrous upper lamina with few glandular hairs. CHR herbarium specimens from this location often have more glandular and eglandular hairs. Therefore, this population was not assigned



Figure 4.4: Lamina of *B. bellidioides* from Otira Valley, Arthurs Pass.

to a specific variety of *B. bellidioides* in this study.

***Brachyglottis bellidioides* var. *angustata*.** Five vouchers from population 19 in the Bealey Valley, Arthurs Pass were primarily identified as *B. bellidioides* var. *angustata*. These vouchers had small, sub-acute, elongated leaves with few to many glandular hairs on the upper lamina and glabrous lower lamina. This population is growing in and around a swamp. Individuals growing in the waterlogged soil of the swamp often had extremely elongated leaves (see figure 4.5). *B. bellidioides* var. *angustata* has no known type locality, however, the Bealey Valley falls inside the southern limit of its range as described by Allan (1961).

***Brachyglottis bellidioides* var. *crassa*.** Population 46 was sampled from Table Hill, Stewart Island which is the type locality of *B. bellidioides* var. *crassa*. The single voucher taken from Table Hill and a second voucher taken from near



Figure 4.5: *B. bellidioides* var. *angustata* from Bealey Valley, Arthurs Pass.

by Mount Rakeahua (population 45) matched the morphological description of *B. bellidioides* var. *crassa* in Allan (1961). Vouchers of both populations had small, appressed, broad-elliptic to sub-orbicular, highly rugose, dark green leaves (see figure 4.6). The upper lamina was covered with dark glandular hairs and the lower lamina had few glandular hairs mixed with long coarse eglandular hairs.

***Brachyglottis bellidioides* var. *setosa*.** Population 6 was sampled from Mount Fyffe near Kaikoura which is the type locality of *Brachyglottis bellidioides* var. *setosa*. However, typical rosette-*Brachyglottis* at this location closely match the morphological descriptions of *B. lagopus* and/or *B. saxifragoides* (see figure 4.7). They commonly have large, broad-elliptic, cordate, leaves with crenate margins. The lower lamina is usually densely tomentose and the upper lamina is covered in glandular hairs or nearly glabrous in some older leaves. Scapes are often highly branched and covered with red glandular and eglandular hairs. The single voucher taken from this location was identified as an intermediate of *B.*



Figure 4.6: *B. bellidioides* var. *crassa* from Table Hill, Stewart Isl.

lagopus and *B. saxifragoides* due to the near absence of glandular hairs on most of the upper lamina of its largest leaf.



Figure 4.7: A large *B. cf. lagopus* from Mt Fyffe, Kaikoura.

Brachyglottis traversii. *Brachyglottis traversii* was primarily represented by population 13 from the Blackwater River, West Coast. The voucher from the Blackwater River had highly elongated, sub-acute leaves, the largest of which had a lamina 6.8cm long by 3.2cm wide lamina and a 6.4cm petiole. The leaves were nearly glabrous with few glandular hairs on either lamina surface and prominent, erect hydathodes. Scapes were long and branched with large leafy bracts with similar morphology to rosette leaves. This voucher matches the description of *B. traversii* given by Allan (1961) and the description of *S. cochlearis* given by Simpson and Thomson (1942).

Population 10 from the Aorere River, Tasman was also a potential candidate for *B. traversii*. The voucher from this location had elongated elliptic leaves, the largest of which had a lamina 5.4cm long by 2.0cm wide lamina and a 2.2cm petiole. The leaves had few, small glandular hairs on both lamina surfaces and many glandular hairs interspersed with prominent hydathodes around the margin. Individuals from this population had many unbranched scapes with few to many glandular hairs. The Aorere River does not match the type locality or distribution of *B. traversii* described by Allan (1961) because it is unlikely to be described as “Alps near Nelson” and is not a tributary of the Buller River.

Brachyglottis southlandica. Two populations were chosen to represent *B. southlandica*. Population 41 from Black Gully, Blue Mountains, Otago fits the type locality of “neighbourhood of Balclutha, Gore, Tapanui, etc.”. The voucher from this population had broad-elliptic to sub-orbicular, cordate base, crenate margins and leaves with a very fine sub-appressed tomentum on the lower lamina and very fine downy eglandular hairs on the upper lamina. The scape was branched and covered with fine eglandular hairs. This population had a mix of plants that were identified as *B. southlandica* var. *southlandica* and *B. southlandica* var. *albidula* (including the voucher).

Population 34 from Ben Lomond, Queenstown was also identified as *B. southlandica*. This population contained a mixture of plants with morphologies that passed from *B. southlandica* into *B. bellidioides*. The single voucher resembled *B. southlandica*, however, plants of the full morphological range were represented in genetic analyses. The voucher had broad-elliptic to sub-orbicular leaves with a cordate base and entire margins. The lower lamina was deep purple with few glandular hairs and the upper lamina green with few glandular hairs. The scape was branched with few glandular hairs. Ben Lomond, Queenstown is north of the geographical range of *B. southlandica* given by (Allan 1961), however, many herbarium specimens in CHR have been identified as *B. southlandica* from this area.

Populations 35, 42, 43 and 44 from Otago all had individuals that could be identified as *B. southlandica* or *B. bellidioides*. In these populations few specimens could be conclusively assigned to a species. This was because many specimens had few glandular hairs on the upper lamina, sporadic purple colouration on the lower lamina and leaves that were not clearly spreading or appressed and between five and eight centimetres in length (Allan 1961).

***Brachyglottis southlandica* var. *albidula*.** *Brachyglottis southlandica* var. *albidula* was represented by population 40 from its type locality of Whiskey Gully, Blue Mountains, Otago (Allan 1961). The voucher from this population had large, broad-elliptic, cordate leaves on long thin petioles. The lamina margin was crenate and approaching serrate in some individuals including the voucher. The lower lamina was covered in a fine sub-appressed tomentum and the upper lamina was near glabrous but with very fine downy eglandular hairs. The scape was highly branched with leaf-like bracts and covered with fine downy hairs.

4.2 Morphological Analyses

A PCoA of morphometric distances among 354 herbarium specimens, including those collected as part of this research project, revealed some taxonomic structure as shown in figures 4.8 and 4.9. The 354 specimens formed a large continuous cloud which contained several clusters of individuals. Each of these clusters mostly consisted of individuals that were assigned to the same species.

Specimens identified as *B. lagopus* and *B. saxifragoides* formed a cluster between *B. bellidioides* and *B. haastii*. The type specimen of *B. saxifragoides* was somewhat removed from this cluster and the type specimen of *B. lagopus* was not included in the analysis. Population vouchers identified as *B. lagopus* and *B. saxifragoides* were mostly included in this cluster. Specimens identified as *B. bellidioides* and its varieties formed a dense cluster around the four varietal type specimens of *B. bellidioides* included in the analysis. The majority of population vouchers identified as *B. bellidioides* or its varieties were included in this cluster. Specimens identified as *B. haastii* mostly clustered around the type specimen of *B. haastii*. Two of the three vouchers from population 32 grouped closely with this voucher. The voucher of population 40 identified as *B. southlandica* var. *albidula* also grouped with the type specimen on *B. haastii*. Specimens identified as *B. traversii* mostly grouped with *B. bellidioides*. However, the type specimen of *S. cochlearis* was distanced from this cluster along axis two and was positioned close to specimens identified as *B. southlandica*. Specimens identified as *B. southlandica* mostly fell between *B. lagopus* and *B. bellidioides*. Those identified as *B. southlandica* var. *albidula* mostly fell between *B. lagopus* and *B. haastii*. The distinction between *B. southlandica* and *B. southlandica* var. *albidula* was along axes one and three.

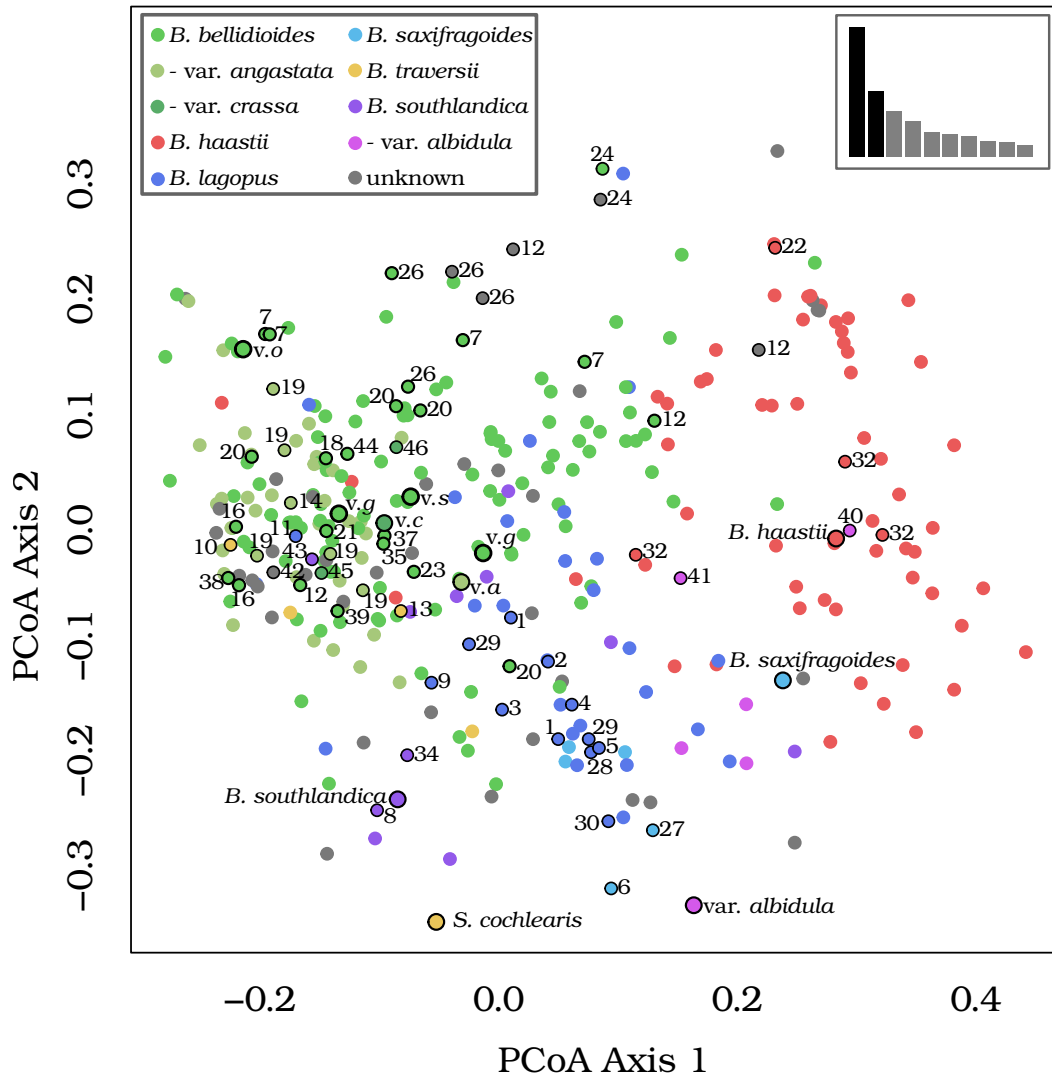


Figure 4.8: Bidimensional plot of the first and second axes from a principal coordinates analysis of 354 rosette-*Brachyglottis* specimens based on Gower's dissimilarity coefficient. Numbers indicate vouchers with populations as numbered in table 4.1. Type specimens included in the analysis are labelled with their respective species names. Type specimens of varieties of *B. bellidioides* are labelled as follows: 'v.a' = var. *angustata*, 'v.c' = var. *crassa*, 'v.g' = var. *glabratus*, 'v.o' = var. *orbiculata*, 'v.s' = var. *setosa*. The relative variation explained by each of the first five PCoA axes is shown in a bar plot. The first and second axes explain 44.4% and 22.4% of the variation respectively.

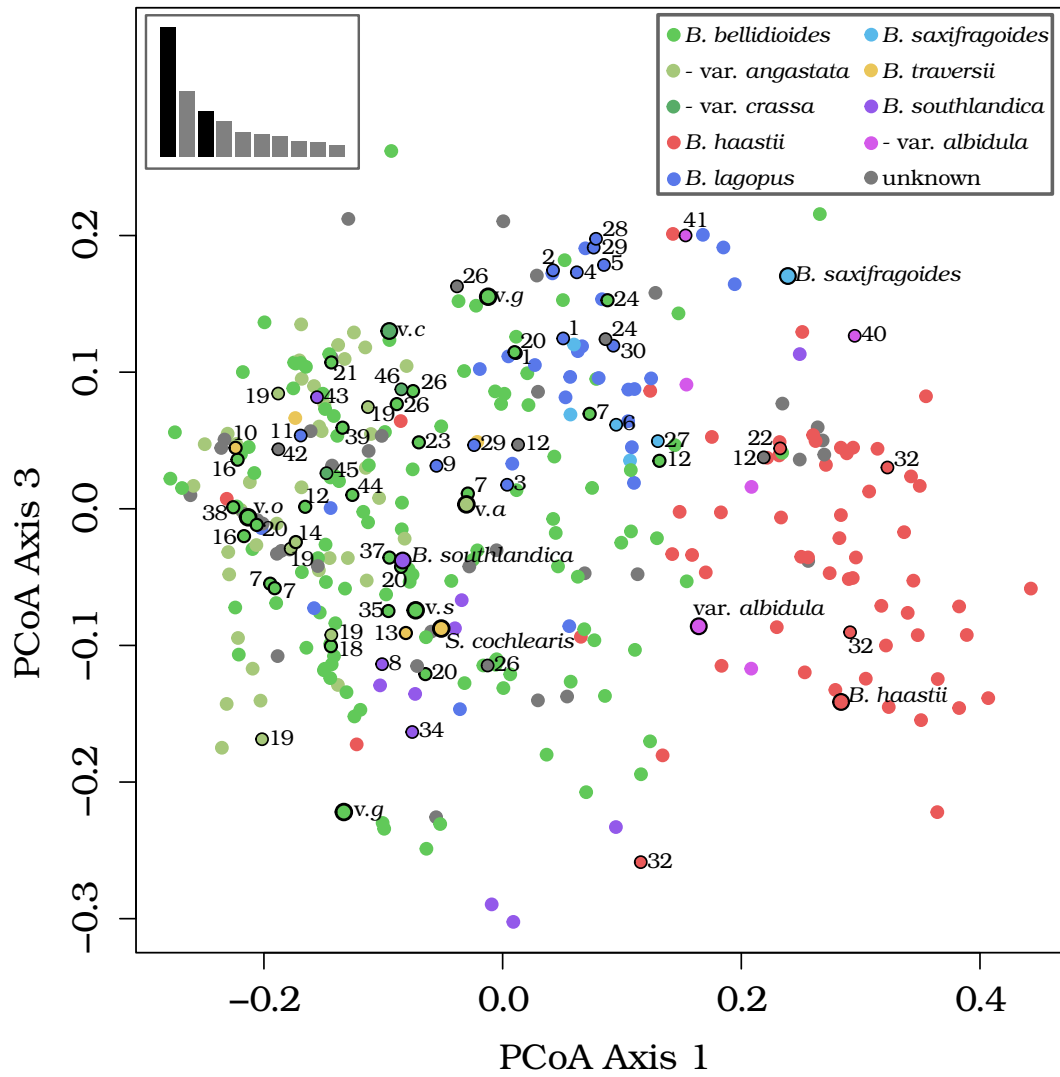


Figure 4.9: Bidimensional plot of the first and third axes from a principal coordinates analysis of 354 rosette-*Brachyglottis* specimens based on Gower's dissimilarity coefficient. Numbers indicate vouchers with populations as numbered in table 4.1. Type specimens included in the analysis are labelled with their respective species names. Type specimens of varieties of *B. bellidioides* are labelled as follows: 'v.a' = var. *angustata*, 'v.c' = var. *crassa*, 'v.g' = var. *glabratus*, 'v.o' = var. *orbiculata*, 'v.s' = var. *setosa*. The relative variation explained by each of the first five PCoA axes is shown in a bar plot. The first and third axes explain 44.4% and 15.7% of the variation respectively.

4.3 Analyses of Genetic Markers

4.3.1 Molecular Marker Diversity

The nine markers yielded a total of 177 alleles from 273 individual plants across 46 sample locations. Twenty individuals did not amplify for one to two markers leaving a total of 253 complete samples. Each marker amplified five to 41 alleles from 184 to 475 base pairs in length as shown in Table 4.3. Table 4.3 also shows the maximum and mean numbers of alleles produced per plant sample by each marker. The mean number of alleles produced per sample gives an indication of the weight given to each marker in the genetic analyses described below.

Table 4.3: Summary of genetic alleles produced by nine markers.

Marker	Total Alleles	Size Range	Sample Maximum	Sample Mean
Pgic-12	5	184-189	2	1.1
Smht-04	18	202-259	5	3.3
Bra-06	33	374-475	4	2.2
Bra-09	15	295-340	5	2.4
Bra-12	7	247-268	4	1.5
Bra-19	41	203-377	3	1.5
Bra-21	25	199-307	2	1.5
Bra-25	23	342-409	4	1.8
Bra-26	10	256-282	2	1.5

4.3.2 Delimitation of Genetic Groups

Bayesian clustering analysis in Structure of 177 allele characters for 273 individuals indicated different hierarchical levels of clustering. In situations of complex hierarchical clustering the first optimal value of K will only show a single level of clustering (Evanno et al. 2005). This has lead to some studies performing additional separate STRUCTURE analyses on each genetic grouping that was determined by the first optimal value of K (Medrano et al. 2014). In our genetic

analysis, criteria used to choose the best value of K were incongruent with one another. The ΔK method of Evanno et al. (2005) used for choosing K (shown in figure 4.10A) indicated a maximum value of $K = 2$. This method has been shown to accurately recover a K value that indicates the highest level of hierarchical clustering in the data (Evanno et al. 2005). The $L(K)$ method of choosing K indicated a K value of eight shown by the plateauing of likelihood at $K = 8$ in figure 4.10B. Beyond $K = 8$ there was a large increase in the variation of $L(K)$ between runs of the same value. The incongruence between methods of choosing K was interpreted as evidence of hierarchical levels of clustering. Structure results for both $K = 2$ and $K = 8$ were explored to investigate multiple levels of genetic clustering in the rosette-*Brachyglottis*.

4.3.3 High Level Genetic Clustering

The higher level of genetic structure in the data found when $K = 2$ separated the plant populations on an approximately north-south gradient (see figures 4.11 and 4.12). The northern genetic cluster (cluster one in figure 4.11) was primarily composed of the North Island populations and some populations from the northern South Island. Population 9 from the Red Hills Plateau, Marlborough was the only South Island population included in the northern genetic cluster with near 100% membership across all individuals. Four more South Island populations had greater than 50% membership to the northern genetic cluster. These included Mount Fyffe, Mount Riley, Blackwater River and Lewis Pass (populations 6, 8, 13 and 14 respectively). Plants From Mount Arthur and Ghost valley (populations 11 and 12 respectively) also had admixed individuals with majority membership to the northern genetic cluster. Populations from the central South Island and further south were predominantly assigned to the southern genetic cluster.

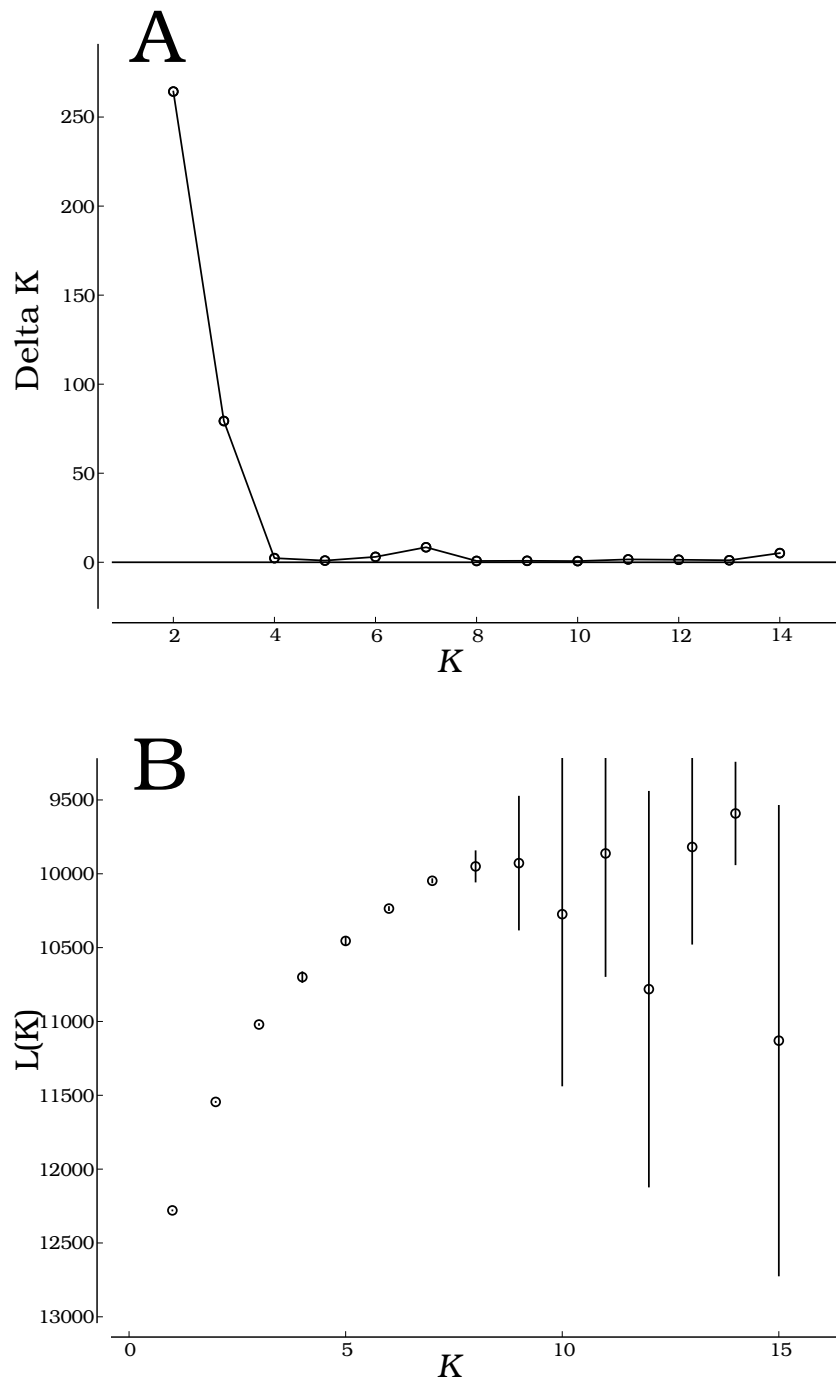


Figure 4.10: Choosing the optimal value of K for Bayesian clustering analysis in STRUCTURE. Figure 4.10A shows the rate of change in estimated posterior probability of K (ΔK) at each value of K . Figure 4.10B shows the estimated posterior probability of K ($L(K)$) and its variation among runs for each value of K .

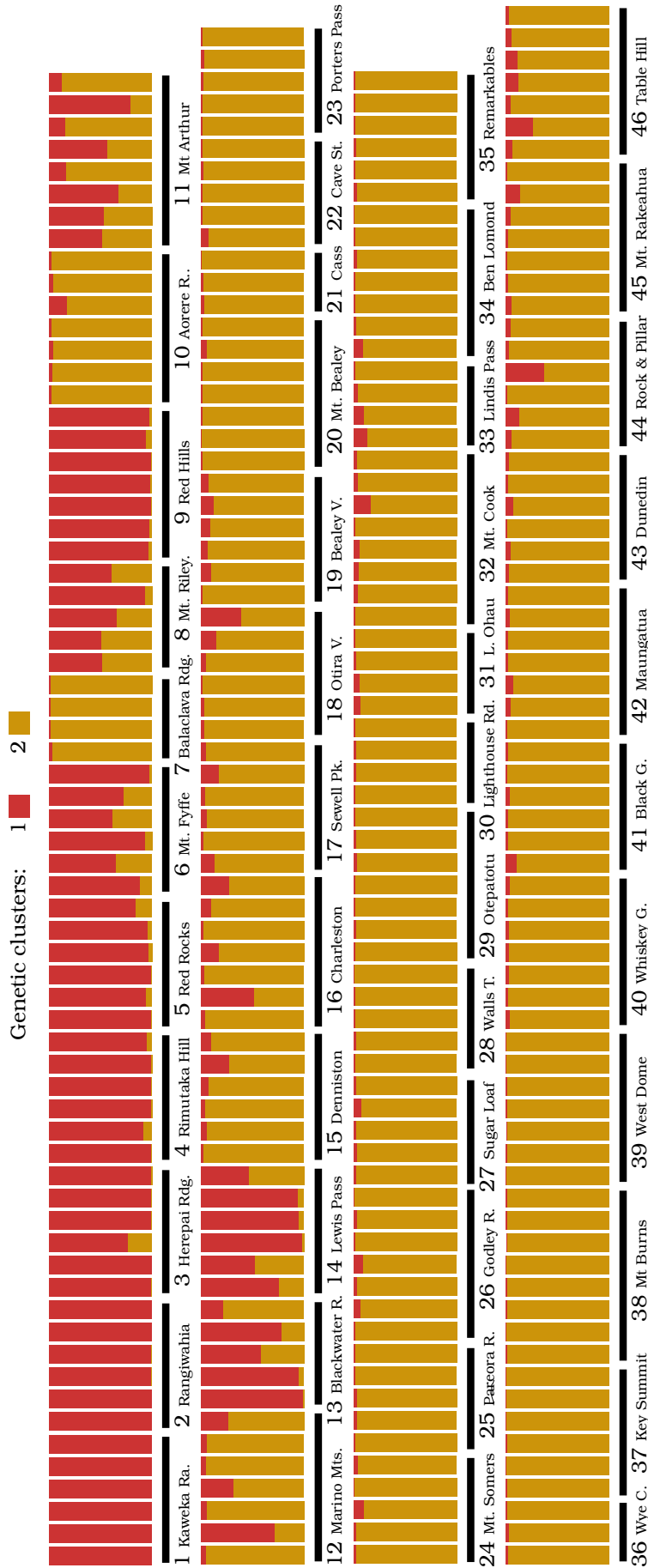


Figure 4.11: Membership of 273 rosette-*Brachyglottis* plants to genetic clusters as assigned by Bayesian cluster analysis in Structure when $K = 2$. Each bar represents an individual plant and bar colours are proportional to that individual's membership to genetic clusters. Individuals are grouped by population, and population numbers correspond to those listed in table 4.1. Individuals were assigned to clusters in ten separate runs of the analysis and results of all runs were merged with Clumpp.

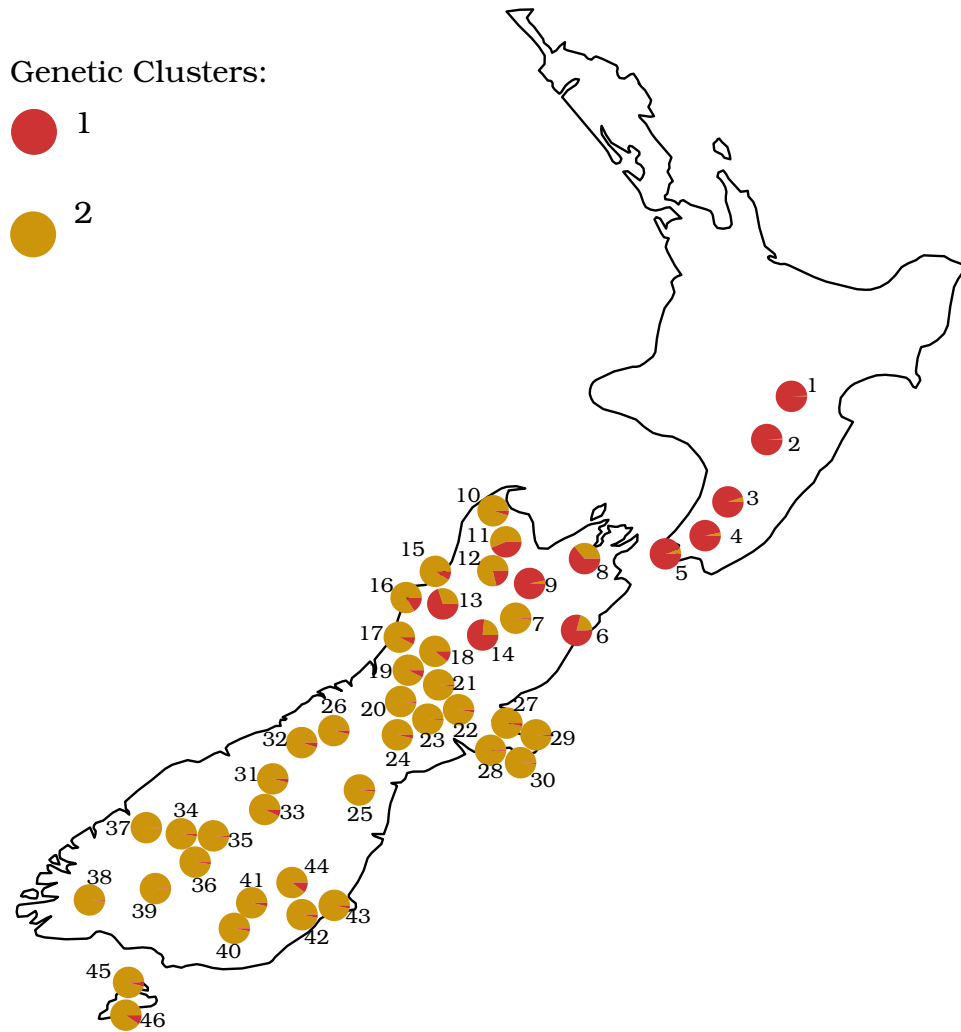


Figure 4.12: Map of sample populations showing membership to genetic clusters as assigned by STRUCTURE when $K = 2$. Proportion of membership to each genetic cluster has been averaged across individuals for each sample population. Population locations have been adjusted where necessary to ensure visibility of all populations. Population numbers correspond to those listed in table 4.1.

4.3.4 Low Level Genetic Clustering

Lower level genetic structure was explored by using a K value of eight and is shown in figure 4.13. Lower level divisions of the genetic data generally clustered plants and populations from similar geographical regions together as shown in figure 4.14. It also revealed that individuals within each population were often genetically similar to one another (see figure 4.13).

The north-south division found for $K = 2$ was partially preserved for $K = 8$. However, when $K = 8$ the northern genetic cluster was almost restricted to the North Island (populations 1-5). The increased value of K highlighted lower level genetic structure in the South Island. This genetic structure grouped the populations into broad geographical regions of Marlborough-North Canterbury (populations 6, 8 and 9), Central Canterbury (populations 31-33), Banks Peninsular (populations 27-30), South Canterbury (populations 31-33), western Southland (populations 37-39) and central to eastern Otago-Stewart Island (populations 34-36 and 41-46). Population 7 from Balaclava Ridge, Marlborough grouped with central Canterbury populations rather than with the Marlborough-North Canterbury group. Population 10 from the Aorere River, north-west Tasman, was highly homogeneous and its members had almost 100% membership to genetic cluster three. Other populations throughout Tasman, West Coast and Arthurs Pass (populations 11-20) were highly mixed with many admixed individuals. The mixed genetic groupings of these populations formed a gradient between the Aorere River, Marlborough and central Canterbury Populations. There were also many admixed individuals between the western Southland and Otago groupings, and in population 33 from Lindis Pass, South Canterbury which represents the confluence the Canterbury and Southland-Otago genetic groups. Figure 4.14 shows an abrupt genetic difference between the Banks peninsula (populations 27-30) and central Canterbury groups (populations 31-33).

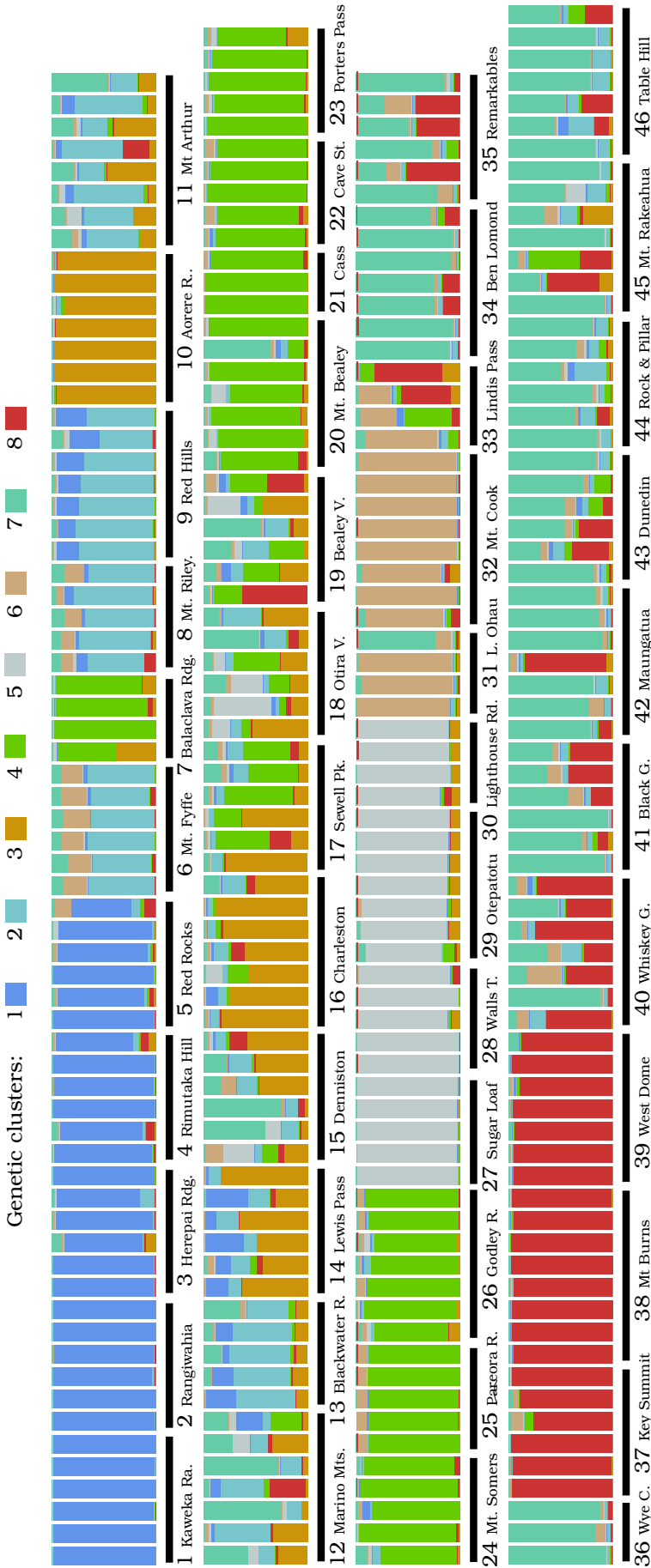


Figure 4.13: Membership of 273 rosette-*Brachyglottis* plants to genetic clusters as assigned by Bayesian cluster analysis in Structure when $K = 8$. Each bar represents an individual plant and bar colours are proportional to that individual's membership to genetic clusters. Individuals are grouped by population, and population numbers correspond to those listed in table 4.1. Individuals were assigned to clusters in ten separate runs of the analysis and results of all runs were merged with Clumpp.

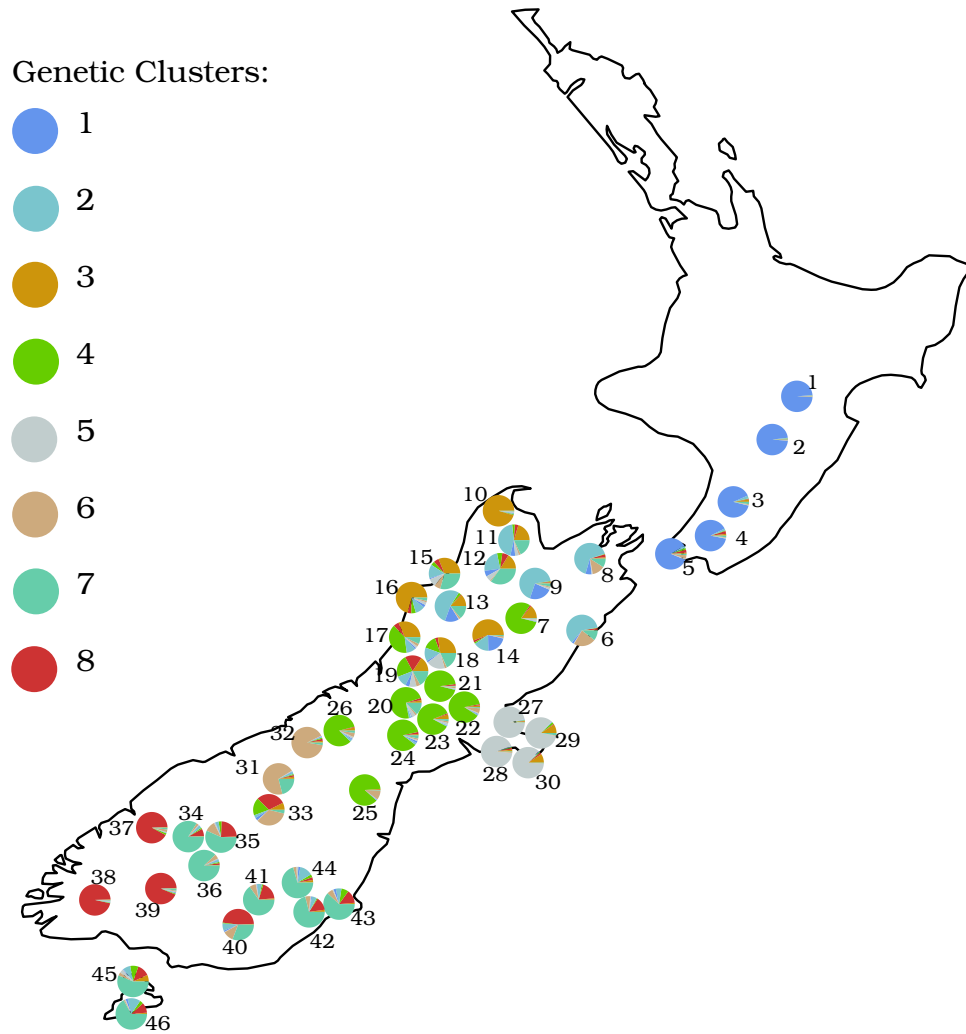


Figure 4.14: Map of sample populations showing membership to genetic clusters as assigned by STRUCTURE when $K = 8$. Proportion of membership to each genetic cluster has been averaged across individuals for each sample population. Population locations have been adjusted where necessary to ensure visibility of all populations. Population numbers correspond to those listed in table 4.1.

4.3.5 Pairwise Genetic Distance

A PCoA of pairwise, Jaccard genetic distances between individuals showed a high level of congruence with the results of the Bayesian cluster analysis performed in STRUCTURE (see figures 4.15 and 4.16). Individual plants generally clustered closely to others from the same populations (plant populations indicated by numbers in figures 4.15 and 4.16). The first three principal coordinate axes respectively explained 6.1%, 3.8% and 3.5% of the variation in the data.

Visualisation of the first three coordinate axes showed that the first axis primarily differentiated North Island plants (populations one to five) from southern populations (see figures 4.15 and 4.16). Most individuals from the two most northern populations (populations 1-2) formed a tight cluster that was slightly removed from other North Island populations. Axis two highlighted relatively large genetic distances between populations 27-30 from Banks Peninsula and populations 31-40 from central and eastern southern South Island (see figure 4.15). Axis three primarily distinguished populations 21-26 from central Canterbury and population 7 from Balaclava Ridge, Marlborough from other south Island Populations.

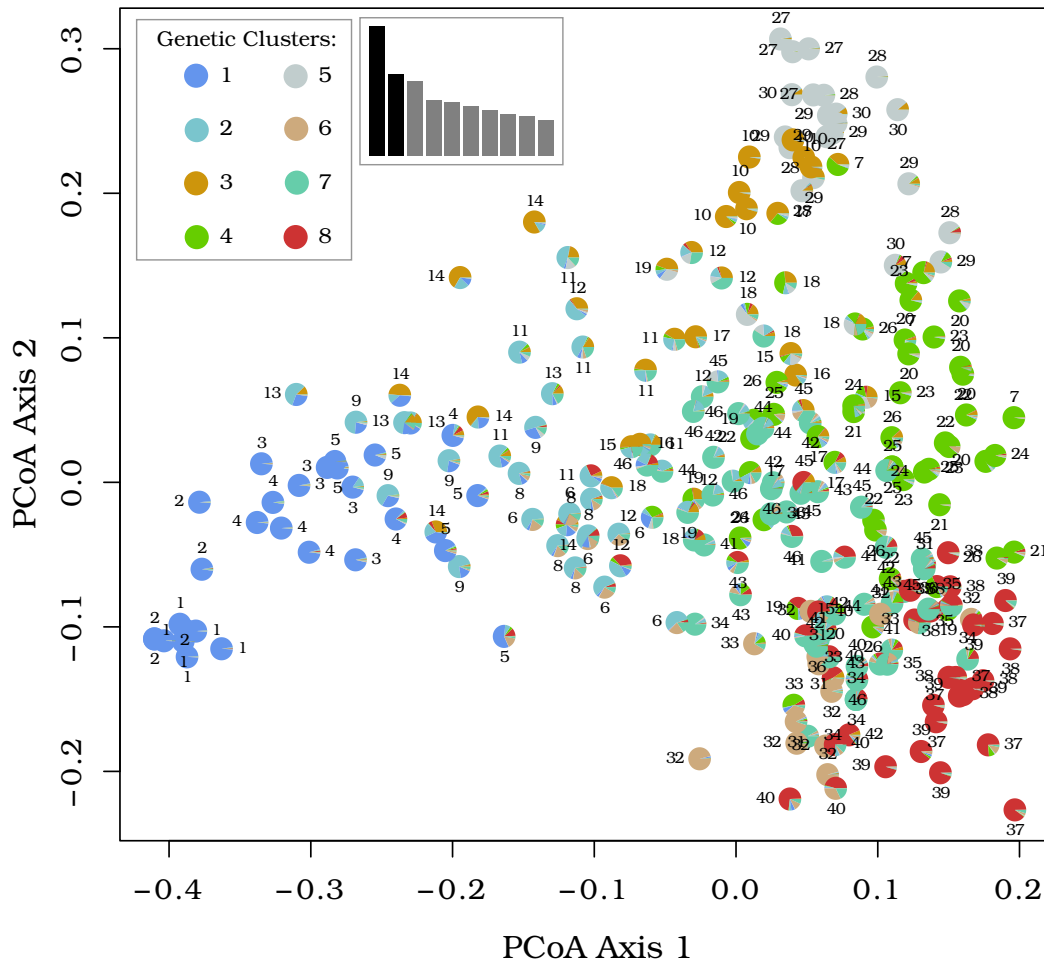


Figure 4.15: Bidimensional plot of the first two axes from a principal coordinates analysis of 253 rosette-*Brachyglottis* specimens based on Jaccard distance. Numbers match individuals with populations as numbered in table 4.1. Colours show the proportional membership of each individual to genetic clusters as assigned by analysis in STRUCTURE when $K = 8$. The relative variation explained by each of the first five PCoA axes is shown in a bar plot. The first and second axes explain 6.1% and 3.8% of the variation respectively. The Jaccard distance matrix was calculated from 177 alleles generated from nine markers.

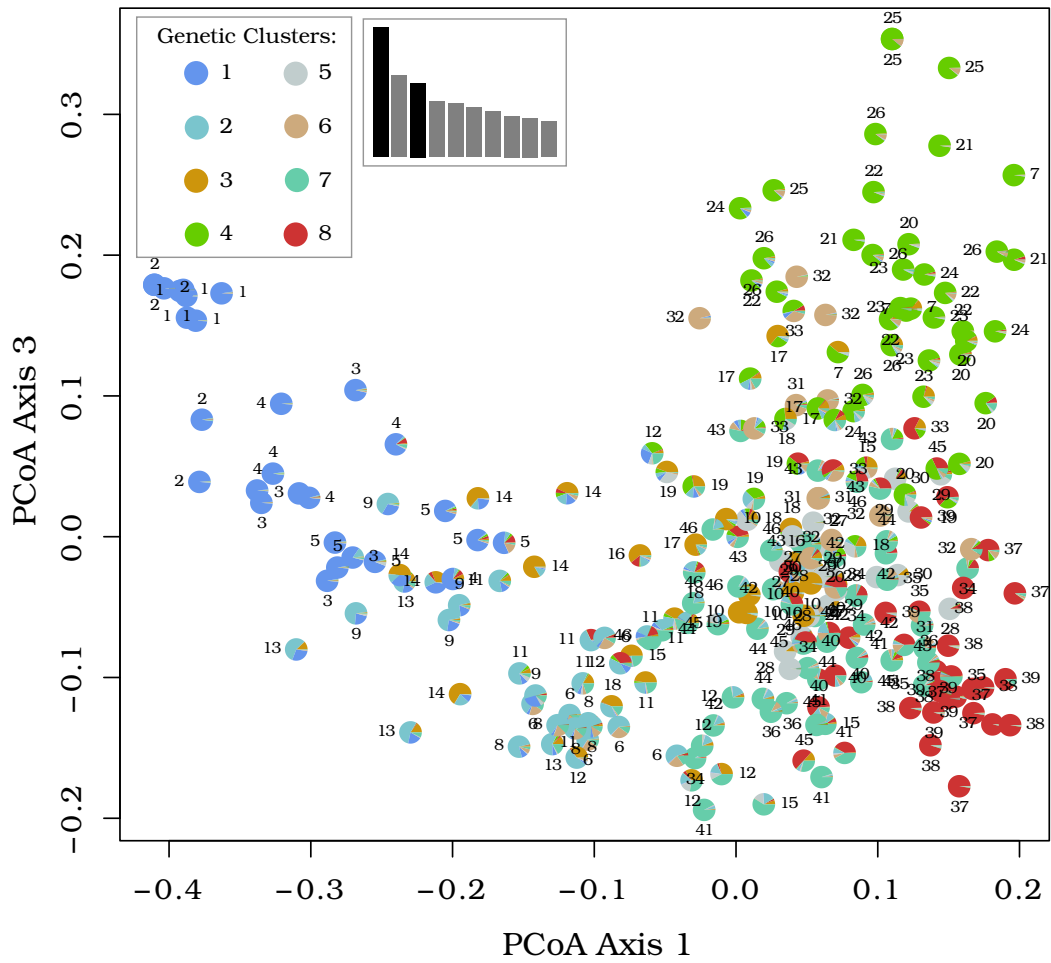


Figure 4.16: Bidimensional plot of the first and third axes from a principal coordinates analysis of 253 rosette-*Brachyglottis* specimens based on Jaccard distance. Numbers match individuals with populations as numbered in table 4.1. Colours show the proportional membership of each individual to genetic clusters as assigned by analysis in STRUCTURE when $K = 8$. The relative variation explained by each of the first five PCoA axes is shown in a bar plot. The first and third axes explain 6.1% and 3.5% of the variation respectively. The Jaccard distance matrix was calculated from 177 alleles generated from nine markers.

A Neighbour-Net analysis of Nei's pairwise population genetic distances was also congruent with the genetic clusters found using STRUCTURE (see figure 4.17). This analysis grouped most populations together with their geographical neighbours. The most prominent exception was population 7 from Balaclava Ridge, Marlborough which grouped with populations 21-26 from central Canterbury. Populations 10-17 from Tasman and the West Coast were inter-dispersed with one another and formed a gradient between North Island and central to southern South Island populations. The three populations (18-20) from Arthurs pass grouped separately from one another although distances between them were not especially large. Population 18 from Otira Valley was the most northern of these (closest to the West Coast) and grouped with populations from Tasman and the West Coast. Population 20 from Mt Bealey was the most southern of these populations (closest to the Central Canterbury) and grouped with populations from Central Canterbury. Population 19 from the Bealey Valley is geographically located between populations 18 and 20 and did not group strongly with other populations.

Identification of population vouchers did not reflect the broader patterns in the genetic data (see figure 4.17). Vouchers from genetically similar populations were often assigned to the same species. However, vouchers from genetically distant populations were also commonly assigned to the same species. A prominent example of this is the two groups of populations 1-5 from the North Island and 27-30 from Banks Peninsula. The North Island vouchers and most Banks Peninsula vouchers were identified as *B. lagopus*, yet they form two very distinct and distant genetic groups that match their geographical locations (see figure 4.17). It was also common for vouchers specimens from genetically similar populations to be assigned to different species.

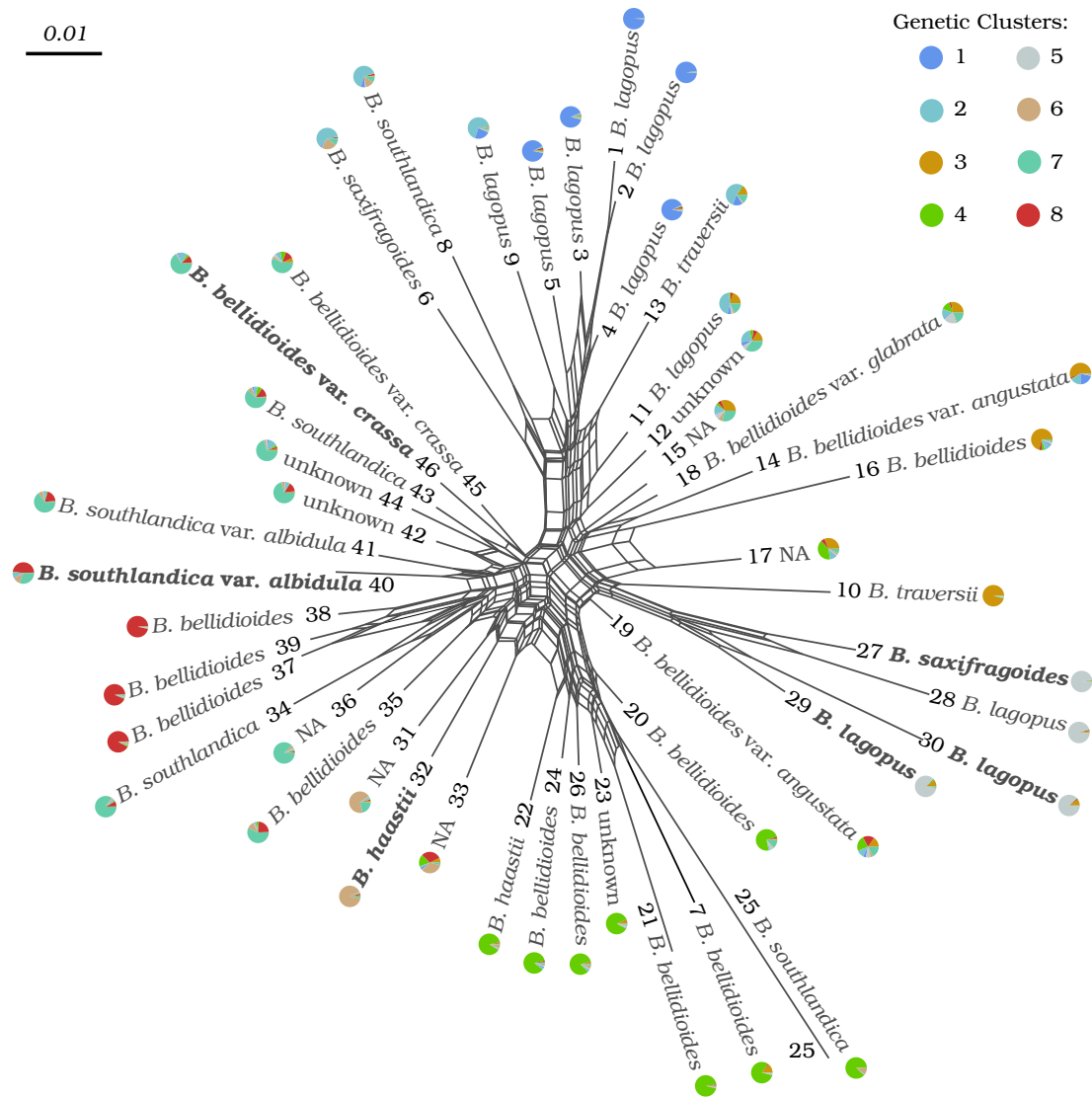


Figure 4.17: Neighbour-Net analysis of pairwise population genetic distances calculated using Nei's dissimilarity coefficient. Population numbers correspond to those listed in table 4.1. The identification of population vouchers are shown for populations (NA = no voucher). Populations with voucher species names shown in bold are from the type locality of that species. Population 6 is from the type locality of *B. bellidioides* var. *crassa* however, it was identified as a different taxon. Vouchers were identified using Allan (1961). A pie chart show each population's proportional membership to genetic clusters as assigned by STRUCTURE when $K = 8$. Proportion of membership to each genetic cluster has been averaged across individuals for each sample population.

4.3.6 Isolation by Distance

The results of both Bayesian clustering and distance based analyses indicated that genetic distance between populations seemed to be positively correlated to their geographical distance (see figures 4.12, 4.14 and 4.17). This relationship was further investigated using linear regression and a Mantel test. These analyses indicated a statistically significant, positive correlation between pairwise population genetic and geographical distances as shown in figure 4.18.

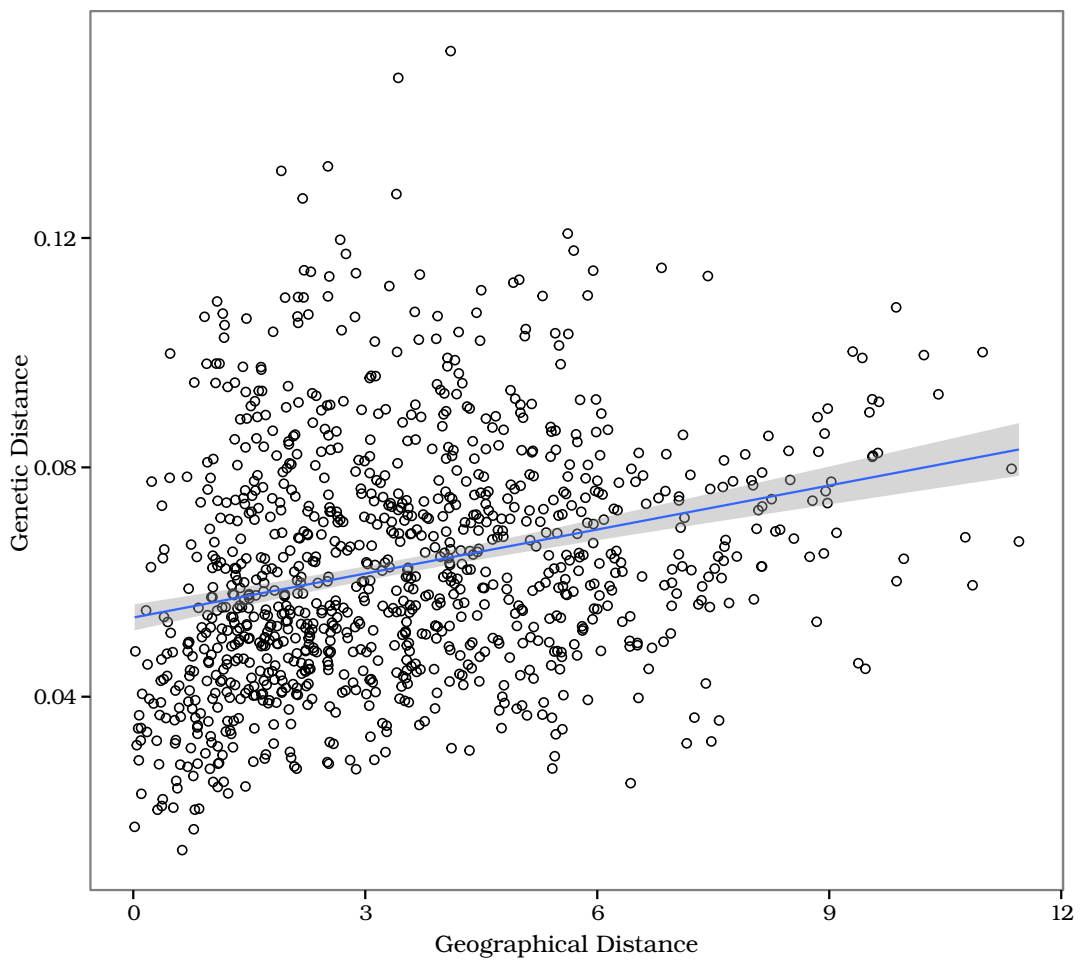


Figure 4.18: Linear regression of pairwise euclidean geographical distance and pairwise Nei's genetic distance between populations. A fitted line is shown with confidence interval. Results of a Mantel test using the same data indicate statistically significant ($P = 0.0005$) positive correlation (observed $r = 0.271$).

Chapter 5

Discussion

5.1 Morphology and Taxonomy

5.1.1 Species Identification

Identification of voucher specimens was often problematic. Many populations showed high morphological diversity among rosette-*Brachyglottis* plants. While this variability could, at times, allow for the identification of multiple species from the same population, intermediate morphological forms were always present. This often resulted in populations that seemed to pass from one species into another. Similar observations have been made by Hooker (1864), Cockayne (1914), and Wall (1918). The presence of intermediate forms meant that the identification of multiple species from a single population was an unsatisfactory explanation of the morphological variation.

Morphological variation was most often a problem in large populations from the sub-alpine locations in the central South Island. When using the descriptions and key of Allan (1961), problematic populations from the central South Island usually appeared to pass from *B. bellidioides* into one of the other rosette-*Brachyglottis* species. This is in part due to the broad morphological description of *B. bellidioides*

and its varieties. Allan (1961) describes *B. bellidioides* as plants with an upper lamina that may be glabrous to covered in glandular hairs and a lower lamina that is glabrous to glandular hairy or tomentose. The scape may be unbranched to branched and can be glabrous to covered with tomentose and or glandular hairs. *Brachyglottis bellidioides* and its varieties also show large variation in leaf shape (Allan 1961). The leaf shape of this group ranges from sub-orbicular to narrow-elliptic with a sub-acute to obtuse apex and cuneate to near cordate base (Allan 1961). This variability means that the hair characters and leaf shape of *B. bellidioides*, as described by Allan (1961), often overlap with the hair characters and leaf shape of all other rosette-*Brachyglottis* species. This is highly problematic because hair characters and leaf shape are the primary morphological features used by Allan (1961) and other authors to distinguish between rosette-*Brachyglottis* species (Hooker 1853, 1864, Kirk 1899).

In practice, the distinction between *B. bellidioides* and other rosette-*Brachyglottis* species often comes down to leaf size and whether the leaves are appressed or erect to spreading (Allan 1961). Neither of these characters are ideal for identification as they vary greatly within populations of otherwise similar individuals. This is reflected across multiple circumscriptions of rosette-*Brachyglottis* by the changing leaf sizes of *B. bellidioides* and other species (Hooker 1853, 1864, Kirk 1899, Allan 1961).

Variation in leaf size within populations often seems to relate to the micro-habitat of individuals. Leaves are often smaller and more appressed when plants are growing in exposed locations and larger with longer, spreading petioles when growing in sheltered locations or amongst dense foliage. This was commonly the case in populations from Otago where large specimens were keyed to *B. southlandica* and smaller specimens to *B. bellidioides*. However, a large number of specimens (often the majority) would fall between the lower leaf size of *B.*

southlandica (eight centimetres) and the upper leaf size of *B. bellidioides* (five centimetres) given by Allan (1961). An example of this is population 34 from Ben Lomond in Otago shown in figure 5.1. Morphological variation in this population and others like it could be interpreted as as evidence of hybridisation, although hybridisation on this scale would indicate little to no reproductive barriers between species. Furthermore, genetic analysis of population 34 indicates that it is, genetically, a relatively homogeneous population (see figure 4.13).



Figure 5.1: Variation in rosette-*Brachyglottis* from Ben Lomond, Otago.

5.1.2 Morphological Analysis

Morphological analysis of herbarium specimens indicate that the rosette-*Brachyglottis* species as delimited by (Allan 1961) are morphologically overlapping. The overall pattern is of continuous morphological variation without clearly separated clusters

(see figures 4.8 and 4.9). Within the general ‘cloud’ of data, specimens were loosely grouped according to the species they were assigned to and these groups sometimes formed denser clusters. Identification among these specimens will be somewhat inconsistent due to their separate determination by multiple botanists however, inconsistent identification will not affect the pattern of continuous morphological variation. This is highly indicative of a single morphological species when applying the genotypic cluster species concept (Mallet 1995, Coyne and Orr 2004).

Interestingly, several type specimens were located around the perimeter of the data cloud indicating that they represent morphological extremes of the rosette-*Brachyglottis*. This may indicate that they were recognised as separate species because they are morphological extremes and would partially explain why many individuals seem to fall ‘between’ species.

Specimens that were identified as *B. lagopus* and *B. saxifragoides* grouped closely to one another indicating their morphological similarity. This supports the conclusions of Wall (1918) who highlighted morphological similarity between *B. lagopus* and *B. saxifragoides* and suggested that they are “microspecies” and should be grouped together as an “aggregate”. Unlike Wall (1918) our study also included specimens identified as *B. lagopus* from outside of Banks Peninsula. This included specimens from throughout the North Island which were also grouped with the Banks Peninsula specimens by morphological similarities (see figures 4.8 and 4.9).

Many specimens identified as *B. bellidioides* and *B. bellidioides* var. *angustata* were included in the morphological analysis and they formed a dense group with the other *B. bellidioides* varietal type specimens. There was relatively little distinction among specimens identified as varieties of *B. bellidioides* which is indicative of similar morphological forms. With the exception of an unnamed

variety, all *B. bellidioides* varieties described by Allan (1961) are characterised by the presence of glandular hairs and or glabrous surfaces and no mention is made of silky or tomentose eglandular hairs. It is likely that this has resulted in most or all tomentose or eglandular hairy *B. bellidioides* specimens being assigned to *B. bellidioides* var. *bellidioides*. This would partially explain why specimens identified as *B. bellidioides* var. *bellidioides* were more likely to overlap with specimens identified as other species and the apparent morphological similarity among other varieties of *B. bellidioides*.

Specimens identified as *Brachyglottis southlandica* var. *southlandica* and *B. southlandica* var. *albidula* were relatively poorly represented in our morphological analysis, although specimens identified as these taxa did form loose groups. The distinction between *B. southlandica* var. *southlandica* and *B. southlandica* var. *albidula* is likely the result of increased downy-tomentose eglandular hairs on the upper and lower lamina of *B. southlandica* var. *albidula* making it appear morphologically similar with *B. haastii*. Likewise the often glabrous lamina surfaces of *B. southlandica* var. *southlandica* could account for its morphological proximity to *B. bellidioides*.

5.2 Genetic Analysis

5.2.1 Molecular Markers and Congruence of Analyses

The nine molecular markers provided adequate resolution for the purpose of this study which was to investigate broad patterns of genetic structure in rosette-*Brachyglottis*. Treating the molecular markers as dominant as opposed to co-dominant markers allowed us to include a larger total number of genetic markers and markers with a larger number of alleles.

Results among Bayesian clustering analysis, PCoA of Jaccard individual genetic distance and Neighbour-Net analysis of Nei's population genetic distances were highly congruent. Each of the analyses made separate assumptions about the genetic data and the congruence of their results is indicative of robust signal in the genetic data.

5.2.2 Incongruence of Genetic Analyses and Taxonomy

The current delimitation of New Zealand's rosette-*Brachyglottis* does not reflect the underlying patterns of their genetic structure. The results of our study clearly show that taxonomic identification of herbarium specimens is not a reliable indication of their genetic (dis)similarity. Furthermore, the historically recognised geographical ranges of several rosette-*Brachyglottis* species was not reflected by the genetic data.

Brachyglottis lagopus is the most prominent example of incongruence between morphological identification and genetic structure among rosette-*Brachyglottis*. Voucher samples from throughout the North Island and eastern Banks Peninsula were identified as *B. lagopus*. Both of these areas reflect the recognised species range of *B. lagopus* which was originally identified from eastern Banks Peninsula and is the only rosette-*Brachyglottis* species recognised in the North Island (Hooker 1853, Allan 1961). The morphological similarity among these populations was highlighted by a morphological analysis in which they grouped closely to one another (see figures 4.8 and 4.9). However populations from these two regions formed two of the most genetically distinct and distant groups among all sampled populations (see figures 4.15 and 4.17). Vouchers from genetically intermediate populations between these two groups were identified as belonging to multiple different species (see figure 4.17).

Genetic results also indicated that populations historically identified as *B. saxifragoides* from western Banks Peninsula are genetically very similar to *B. lagopus* from eastern Banks Peninsula. Morphological analysis also indicated similarity between vouchers of these populations (see figures 4.8 and 4.9). These results confirm the findings of Mennes et al. (2013) who first described genetic similarity between *B. lagopus* and *B. saxafragoides* on Bank Peninsula and the relative genetic dissimilarity between *B. lagopus* from Bank Peninsula and *B. lagopus* from elsewhere in the South Island using AFLP data.

The example of *B. lagopus* was paralleled by similar patterns of incongruence between morphological and genetic marker data for *B. haastii* from Red Tarns, Mount Cook and Cave Stream, Castle Hill Basin. *Brachyglottis haastii* was originally described from Mount Cook and has historically been identified from Castle Hill Basin (Hooker 1853, Kirk 1899). Population vouchers from both of these locations were identified as *B. haastii* without uncertainty and their morphological similarity was reflected by their relatively close proximity in our morphological analysis (see figures 4.8 and 4.9). Despite this apparent morphological similarity, the Cave Stream population was genetically most similar to its geographical neighbours that were generally identified as *B. bellidioides* (see figure 4.17).

The morphological identification of other species also did not always reflect patterns of genetic similarity. However, because identification of vouchers was often problematic it is difficult to give conclusive examples of incongruence between species delimitation and genetic dissimilarity for all rosette-*Brachyglottis* species. For example *B. bellidioides* var. *bellidioides* was identified, with varying degrees of confidence from populations throughout the South Island but these populations were often genetically dissimilar from one another. Populations identified as *B. bellidioides* var. *bellidioides* were only genetically similar to one another when they were geographically close to one another.

5.2.3 Reproductive Isolation by Genetic Distance

Genetic analysis of rosette-*Brachyglottis* indicated a statistically significant, strong correlation between genetic and geographic distances among populations (see figure 4.18). This relationship was evident in the results of Bayesian clustering and distance based analyses (see figures 4.14, 4.17). Two of the most distinct genetic groups of populations were from the North Island and Banks Peninsula (see figures 4.12 and 4.17). Both of these groups are separated from other rosette-*Brachyglottis* populations by large geographical distances (Cooks Strait and the Canterbury Plains respectively). This fits a hypothesis of (partial) reproductive isolation by geographical distance alone.

Reproductive isolation by distance is also indicated within geographic regions. Among North Island populations, genetic distance from South Island rosette-*Brachyglottis* increased the further north populations were located (see figure 4.17). This is also indicated by the relative genetic similarity among populations from the North Island and north eastern South Island.

Geographical distances were also reflected in the genetic distances among populations from Banks Peninsula. Populations 27 and 28 from eastern Banks Peninsula were genetically more similar to one another than either were to Populations 29 and 30 from the Port Hills, western Banks Peninsula (see figure 4.17). The Port hills population were likewise genetically more similar to one another than either were to populations from eastern Banks Peninsula. This pattern of genetic distances among rosette-*Brachyglottis* on Banks Peninsula matches the patterns found by Mennes et al. (2013). Mennes et al. (2013) also found that rosette-*Brachyglottis* from Banks Peninsula were genetically distant from other South Island populations which is congruent with our results.

Populations from Stewart Island were not particularly genetically distinct despite the potential barrier to reproduction represented by Foveaux Strait. A Bayesian clustering analysis indicated that individuals from Stewart Island had very mixed genetic backgrounds with a strong genetic influence from eastern Otago populations and to a lesser extent from western Southland populations (see figure 4.13). Distance based genetic analysis indicated that Stewart island individuals were roughly equally distant from most other populations and were most similar to populations from eastern Otago (see figure 4.17). The genetic similarity of Stewart Island populations to those from the South Island and high number of admixed individuals that were partially assigned to genetic clusters found in the southern south Island are indicative of recent migration of southern South Island genotypes to Stewart Island.

Throughout the South Island, population genetic structure was generally congruent with a hypothesis of reproductive isolation by geographical distance alone. Analysis of pairwise genetic distance among individuals revealed a continuum of South Island genotypes between the more genetically distinct populations of the North Island, Banks Peninsula and south western South Island (see figure 4.15).

A fourth somewhat genetically distinct group was identified from central Canterbury (see figure 4.16). This grouping was predominantly assigned to genetic cluster 4 found in the second iteration of Bayesian clustering analyses (4.13) and also included population 7 from southern Marlborough and population 25 from south eastern Canterbury (see figure 4.14).

Population 26 from the ‘central’ Canterbury group and 32 from the ‘southern’ Canterbury group are geographically close to one another, yet were assigned to separate genetic groups in the second Bayesian analysis with minimal evidence of admixture (see figure 4.14). Analysis of genetic distance indicated that these

populations and their genetic groups are in fact relatively similar to one another, but that there is still a significant ‘gap’ between them given their geographical proximity (see figure 4.17).

A similar genetic ‘gap’ was detected between the central Canterbury and West Coast populations. In this case, dense placement of sample populations 18-20 throughout Arthurs Pass allowed for a more detailed interpretation (see figure 4.1). Bayesian genetic analyses indicated that individuals of these populations had highly mixed genetic backgrounds with many admixed individuals and that they represented a sharp genetic gradient between the West Coast and Canterbury (see figure 4.13). This was also reflected in a Neighbour-Net analyses of Nei’s genetic distance among populations (see figure 4.17).

The cause of these genetic ‘gaps’ or sharp ‘gradients’ cannot be determined from our analyses alone. However, potential causes include physical barriers to gene flow and abrupt environmental differences between populations. For example the sharp genetic gradient found in rosette-*Brachyglottis* across Arthurs Pass may in part be caused by the difference in precipitation between the wet West Coast and dry central Canterbury.

5.2.4 Evidence for Biological Species

Genetic analysis did not indicate the presence of distinct biological species of rosette-*Brachyglottis*. This was indicated by Bayesian clustering analysis and two forms of dissimilarity analysis of the genetic data.

High level structure found with Bayesian clustering analysis indicated interbreeding between two genetic clusters highlighted by many admixed individuals in the northern South Island. The ΔK method of Evanno et al. (2005) was used to indicate high level genetic structure found by Bayesian clustering analysis. This

method identifies the greatest rate of change in estimated posterior probability for differing values of K (the number of hypothetical genetic groups) (Evanno et al. 2005). However, this method requires an estimated posterior probability for $K + 1$ and $K - 1$ in order to determine the rate of change (ΔK) at any given value of K . Therefore, this method cannot select $K = 1$ because that would require an estimated posterior probability for $K = 0$. Due to this limitation we cannot rule out the possibility that high level genetic structure in the data is most appropriately described by a single genetic group containing all individuals.

Lower level genetic structure found with Bayesian clustering analysis when $K = 8$ was characterised by genetically mixed populations with many (admixed) individuals that were partially assigned to multiple genetic clusters. This is indicative of (indirect) gene flow among populations across the entire geographical range of rosette-*Brachyglottis*.

Analysis of pairwise genetic distance among individuals produced a pattern that was congruent with genetic isolation by distance. This pattern was continuous in nature and did not reveal multiple discrete clusters of individuals that could indicate reproductively isolated species. This highly indicative of a single genetic species when applying the genotypic cluster species concept (Mallet 1995, Coyne and Orr 2004). These results are incongruent with a hypothesis of reproductively isolated species with overlapping ranges and congruent with our null hypothesis of reproductive isolation by geographical distance alone.

5.3 Summary and Conclusions

In this study we have tested the current morphology-based species delimitation of rosette-*Brachyglottis* with a combination of morphological and genetic analyses. The current delimitation of rosette-*Brachyglottis* recognises species with partially

overlapping ranges and, therefore, implicitly hypothesises a form of reproductive isolation other than geographical distance. This hypothesis was tested against a null hypothesis of geographical distance as the primary form of reproductive isolation in rosette-*Brachyglottis*.

Morphological analysis of 354 specimens produced a pattern of morphological similarities among specimens that was generally congruent with the currently delimited species in rosette-*Brachyglottis*. However, this analyses also revealed many morphologically intermediate forms between the currently delimited species. This complicates species identification of morphologically intermediate specimens.

Despite general congruence between the morphological data and the current species delimitation, our results showed that the current species delimitation does not accurately reflect the genetic structure of New Zealands rosette-*Brachyglottis*. Analysis of nine genetic markers indicated that rosette-*Brachyglottis* form a single biological species. Genetic structure within this species appears to be primarily driven by geographical isolation although there was no evidence of complete reproductive isolation among sample populations. This is indicative of a single species under both the biological species concept of Coyne and Orr (2004) and the genetic cluster species concept of Mallet (1995).

In conclusion the current delimitation of New Zealands rosette-*Brachyglottis* does not reflect genetic patterns among populations of rosette-*Brachyglottis* and is rejected as a hypothesis. The null hypothesis of geographical distance as the only form of reproductive isolation in rosette-*Brachyglottis* is congruent with our results and cannot be rejected. Furthermore, our results indicate that New Zealands rosette-*Brachyglottis* are best described as a single biological species. Our results also show that geographical location is a relatively accurate proxy for estimating genetic similarity among rosette-*Brachyglottis*.

5.4 Future Research

A single species of rosette-*Brachyglottis* would incorporate a remarkable range of morphological forms. Future research in rosette-*Brachyglottis* should focus on determining taxonomic units below the species level that accurately categorise its genetic and morphological diversity. These taxonomic units may, to some extent, reflect the currently recognised species of rosette-*Brachyglottis*. However, new taxa would need to be recognised if this approach would be taken. For example, North Island rosette-*Brachyglottis* form a relatively distinct genetic group but have never been independently recognised as a taxonomic unit.

A second avenue of research is to determine the extent of environmental influences on rosette-*Brachyglottis* morphology and genetic structure. Rosette-*Brachyglottis* morphology clearly varies within and among population but it is unclear what proportion of this variation is plastic response to environmental factors or genetically determined phenotypes. It is also possible that sharp genetic gradients between populations of rosette-*Brachyglottis* are a response to environmental factors.

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Appendices

Appendix A

Morphological Data

Morphological data for 354 herbarium specimens. Character numbers correspond with those described in table 3.1. Herbarium or collector ID is shown for each specimen. Presence or absence of a character is indicated by a ‘1’ or ‘0’ respectively for binary characters.

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
AK-100651	1	0	0	0	1	0	0	0	0	0	1	1	1	0	1	1	2.452	3.448	1.31	0.711136891	2.6320610687	1.8717557252
AK-104643 α	1	0	0	1	1	0	0	0	0	0	1	1	1	0	0	1	1.255	2.44	2.432	0.5143442623	1.0032894737	0.5160361842
AK-104643 β	1	0	0	1	1	0	0	0	0	0	1	1	1	0	1	1	1.255	2.584	1.514	0.4856811146	1.7067371202	0.8289299868
AK-10538	1	0	0	1	1	1	1	1	1	1	0	0	0	1	0	0	10.489	7.624	5.696	1.3757869885	1.3384831461	1.8414676966
AK-10551	1	0	0	0	1	0	0	0	0	0	1	1	1	0	1	1	1.426	1.72	1.004	0.8290697674	1.7131474104	1.4203187251
AK-10555	1	0	1	0	1	0	0	1	0	1	1	1	1	1	1	1	3.307	3.16	1.412	1.0465189873	2.2379603399	2.3420679887
AK-35398	1	1	0	1	1	0	0	0	0	1	0	0	0	0	0	0	1.426	2.008	1.412	0.7101593625	1.4220963173	1.0099150142
AK-35405	1	0	0	1	1	1	0	0	0	0	1	1	1	1	1	0	7.582	6.04	3.962	1.2552980132	1.5244825846	1.9136799596
AK-35407	1	0	1	0	1	0	0	0	0	0	1	0	1	1	0	0	2.794	2.872	2.024	0.9728412256	1.418972332	1.3804347826
AK-35409	1	1	0	1	1	1	1	0	1	1	1	0	1	1	0	0	7.582	8.056	4.574	0.9411618669	1.7612592916	1.6576300831
AK-35410	1	0	0	1	1	1	0	1	1	0	1	0	1	1	1	0	6.385	4.6	3.044	1.3880434783	1.5111695138	2.0975689882
AK-44802	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	1	2.452	2.872	1.922	0.8537604457	1.494276795	1.2757644225
AK-71363	1	0	0	0	1	0	0	1	0	1	1	1	1	0	1	1	2.281	2.584	1.208	0.8827399381	2.1390728477	1.8882450331
CANTY-HAGLEY	1	0	1	0	1	1	0	0	0	1	1	0	1	1	0	0	1.597	3.592	3.044	0.4445991091	1.1800262812	0.5246386334

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
CANTY-M1	1	1	0	1	1	1	0	1	1	1	0	0	0	1	0	0	9.292	5.752	3.86	1.6154381085	1.4901554404	2.407253886
CANTY-M10	1	1	0	1	1	1	1	1	0	1	0	0	0	0	0	0	9.463	6.904	4.064	1.3706546929	1.6988188976	2.3284940945
CANTY-M17	1	0	0	1	1	0	0	0	0	1	1	0	1	0	0	0	1.768	1.864	1.718	0.9484978541	1.0849825378	1.0291036088
CANTY-M19	1	0	1	0	1	0	0	0	0	0	1	1	0	1	1	1	3.82	3.16	2.33	1.2088607595	1.356223176	1.6394849785
CANTY-M2	1	0	0	1	1	1	1	1	1	1	0	0	1	1	0	0	9.976	6.76	4.37	1.475739645	1.5469107551	2.2828375286
CANTY-M22	1	0	0	1	1	0	0	0	0	0	1	0	1	0	0	0	2.452	3.448	2.126	0.711136891	1.6218250235	1.1533396049
CANTY-M24	1	0	1	0	1	0	0	0	0	0	1	0	1	0	0	0	1.597	3.88	2.33	0.4115979381	1.6652360515	0.6854077253
CANTY-M3	1	0	0	1	1	1	1	1	1	1	1	0	1	1	0	0	13.567	9.064	6.002	1.4968005296	1.5101632789	2.2604131956
CANTY-M4	1	1	1	0	1	1	1	1	1	1	1	0	1	0	0	0	7.582	8.056	6.614	0.9411618669	1.2180223768	1.1463562141
CANTY-M5	1	1	0	1	1	1	1	0	0	1	1	0	1	0	0	0	1.939	3.448	2.636	0.5623549884	1.3080424886	0.7355842185
CANTY-YALD	1	1	0	1	1	1	0	0	0	0	1	0	1	0	0	0	1.255	2.728	2.024	0.4600439883	1.347826087	0.6200592885
CANU-10692	1	0	1	1	1	0	0	0	0	1	1	1	0	1	1	1	2.623	3.448	1.922	0.7607308585	1.7939646202	1.3647242456
CANU-1107 α	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	0	2.794	4.744	3.248	0.5889544688	1.460591133	0.8602216749
CANU-1107 β	1	0	0	1	1	1	0	1	1	0	1	1	1	1	1	1	6.385	7.768	6.002	0.821961895	1.2942352549	1.0638120626
CANU-11159	1	0	0	1	1	1	0	0	0	0	1	1	0	1	1	1	2.281	2.728	1.616	0.836143695	1.6881188119	1.411509901
CANU-11162	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	1	1.255	2.584	1.208	0.4856811146	2.1390728477	1.0389072848
CANU-11168	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	1.939	2.872	1.718	0.6751392758	1.6717112922	1.1286379511
CANU-11171	1	0	0	1	1	1	1	0	1	1	0	0	0	0	0	0	7.582	6.04	4.574	1.2552980132	1.3205072147	1.6576300831
CANU-11172	1	1	0	1	1	1	0	0	0	1	0	0	0	0	0	0	3.307	3.448	3.86	0.9591067285	0.8932642487	0.8567357513
CANU-11173	1	0	1	0	1	1	1	0	0	1	0	0	1	1	0	0	1.768	1.72	1.514	1.0279069767	1.1360634082	1.1677675033
CANU-11187	1	0	0	1	1	0	0	0	0	1	0	0	1	1	0	0	0.742	3.016	2.024	0.2460212202	1.4901185771	0.3666007905
CANU-1169 α	1	0	0	1	1	1	0	0	1	1	1	0	1	1	0	0	12.883	7.768	6.002	1.6584706488	1.2942352549	2.1464511829
CANU-12477	1	0	0	1	1	0	0	1	0	0	1	0	0	0	0	0	2.281	3.736	2.534	0.6105460385	1.4743488556	0.9001578532
CANU-12851	1	0	0	1	1	0	0	0	0	1	1	0	1	1	1	0	1.768	5.752	3.35	0.3073713491	1.7170149254	0.527761194
CANU-384	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	1	1.939	2.872	1.922	0.6751392758	1.494276795	1.0088449532
CANU-401	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	1	6.214	4.312	2.84	1.4410946197	1.5183098592	2.188028169
CANU-4514	1	0	0	1	1	0	0	0	0	0	1	0	1	1	1	0	8.437	5.608	8.654	1.5044579173	0.6480240351	0.9749248902
CANU-4523	1	1	1	1	1	1	0	0	0	1	0	0	0	0	0	0	2.281	4.312	3.35	0.5289888683	1.2871641791	0.6808955224
CANU-4524	1	0	1	0	1	1	0	0	0	1	1	1	1	0	0	0	5.701	9.352	6.308	0.6096022241	1.4825618263	0.9037729867
CANU-5895	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	0	1.255	2.872	1.412	0.4369777159	2.0339943343	0.8888101983
CANU-5896	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	0	2.281	3.16	1.514	0.721835443	2.0871862616	1.5066050198
CANU-6262	1	0	1	0	1	1	1	0	1	0	1	0	1	1	0	0	3.991	7.48	5.288	0.5355561497	1.4145234493	0.7547276853
CANU-8838	1	1	0	1	1	1	0	0	0	1	0	0	0	0	0	0	2.281	3.592	2.636	0.6350222717	1.3626707132	0.8653262519
CANU-9140 α	1	0	1	0	1	1	1	0	1	1	1	1	0	1	1	0	5.017	5.464	3.86	0.9181918009	1.4155440415	1.2997409326
CANU-9185	1	0	1	1	1	1	0	1	1	0	1	0	1	1	1	0	1.939	3.448	3.146	0.5623549884	1.0959949142	0.6163382072
CHR-10147	1	1	0	1	1	1	0	0	0	1	0	0	0	0	0	0	2.965	2.872	1.514	1.0323816156	1.8969616909	1.9583883752
CHR-10181	1	1	0	1	0	1	0	1	0	1	0	0	0	0	0	0	7.24	3.88	3.554	1.8659793814	1.0917276308	2.0371412493

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
CHR-10307	1	0	0	1	1	1	0	0	1	1	1	0	0	1	0	0	7.582	6.472	4.064	1.1715080346	1.592519685	1.8656496063
CHR-10406	1	0	0	0	1	1	0	0	0	1	1	1	1	0	1	1	3.991	8.056	3.758	0.49540715	2.143693454	1.0620010644
CHR-10407	1	0	1	0	1	1	0	1	0	0	1	1	0	0	1	1	3.478	7.768	2.636	0.4477342945	2.9468892261	1.3194233687
CHR-10416	1	0	0	1	1	0	0	0	0	0	1	1	1	0	0	1	1.255	3.592	1.922	0.3493875278	1.8688865765	0.6529656608
CHR-10417	1	0	0	1	1	0	0	0	0	1	1	1	1	1	0	1	1.597	3.448	2.534	0.4631670534	1.3606945541	0.6302288871
CHR-1045.a?	0	1	0	1	0	1	0	1	0	1	1	0	0	0	1	0	3.478	4.456	2.534	0.7805206463	1.7584846093	1.3725335438
CHR-1045.B?	1	0	0	1	1	0	0	0	0	0	1	0	1	1	1	0	1.426	2.584	1.82	0.5518575851	1.4197802198	0.7835164835
CHR-10464	1	0	0	0	1	1	1	0	1	1	1	1	0	0	1	1	15.448	12.952	7.838	1.1927115503	1.6524623628	1.9709109467
CHR-108696a	1	0	1	0	1	1	0	0	0	1	1	1	1	1	0	0	0.742	3.736	2.432	0.198608137	1.5361842105	0.3050986842
CHR-108696b	1	0	0	1	1	0	0	0	0	0	1	1	1	0	1	1	0.742	2.296	1.412	0.3231707317	1.6260623229	0.5254957507
CHR-108698a	1	0	1	0	1	1	0	0	0	1	1	0	1	1	0	0	8.95	8.776	4.37	1.0198268004	2.0082379863	2.0480549199
CHR-108698y	1	0	1	0	1	1	0	0	0	1	1	0	1	1	0	0	1.768	4.168	3.248	0.424184261	1.2832512315	0.5443349754
CHR-108703	1	0	1	0	1	1	1	0	1	1	1	1	1	0	0	0	1.255	5.032	2.636	0.2494038156	1.908952959	0.4761001517
CHR-108704a	1	0	1	1	1	1	1	1	1	1	1	1	1	0	1	1	5.188	8.344	4.676	0.6217641419	1.7844311377	1.1094952951
CHR-108704b	1	0	1	0	1	1	1	0	1	1	1	1	1	0	1	0	1.426	4.456	3.35	0.3200179533	1.3301492537	0.4256716418
CHR-108705	1	0	0	0	0	0	0	0	0	1	1	1	1	0	1	0	5.701	6.776	3.452	0.8433431953	1.9582850521	1.6515063731
CHR-108706	1	0	0	0	0	0	0	1	0	0	1	1	1	1	1	1	2.281	1.864	0.902	1.2237124464	2.066518847	2.5288248337
CHR-108707a	1	0	1	0	1	1	1	1	1	1	1	1	1	0	0	0	8.437	7.912	6.614	1.0663549039	1.196250378	1.2756274569
CHR-108707b	1	0	1	0	1	1	0	1	1	1	1	1	1	0	0	0	2.623	3.448	2.126	0.7607308585	1.6218250235	1.2337723424
CHR-108711	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	0	0.742	2.728	2.024	0.2719941349	1.347826087	0.3666007905
CHR-108713	1	0	0	1	1	0	0	0	0	1	1	1	1	0	0	0	2.452	3.448	2.126	0.711136891	1.6218250235	1.1533396049
CHR-108714	1	0	1	0	1	0	0	1	0	1	1	1	0	1	1	1	3.136	4.312	2.024	0.7272727273	2.1304347826	1.5494071146
CHR-108715	1	0	1	0	1	0	0	1	0	0	1	1	1	1	1	1	0.913	1.72	0.8	0.5308139535	2.15	1.14125
CHR-108716	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	1	1.939	2.296	1.106	0.8445121951	2.0759493671	1.753164557
CHR-115377	1	0	1	0	1	1	0	0	0	0	1	1	1	0	0	0	2.452	5.752	3.554	0.426286509	1.6184580754	0.689926843
CHR-11555	1	0	1	0	1	1	1	0	1	0	1	1	1	0	1	1	16.474	13.672	8.654	1.2049444119	1.5798474694	1.9036283799
CHR-117530	1	0	1	0	1	0	0	0	1	1	1	1	1	0	1	1	1.255	4.456	4.064	0.2816427289	1.0964566929	0.3088090551
CHR-117694	1	0	0	0	0	1	0	1	1	1	1	1	1	0	1	1	2.623	7.336	3.86	0.3575517993	1.9005181347	0.6795336788
CHR-118963	1	0	0	1	1	1	1	0	1	1	0	0	0	0	0	0	9.121	5.896	3.86	1.5469810041	1.5274611399	2.3629533679
CHR-119007	1	0	0	1	1	1	1	0	0	1	0	0	0	0	0	0	11.686	6.04	3.554	1.9347682119	1.6994935284	3.2881260551
CHR-119643	1	0	0	1	1	1	0	1	1	1	0	0	0	0	0	0	8.779	5.752	4.268	1.5262517385	1.3477038425	2.0569353327
CHR-122007	1	0	0	1	1	1	0	0	0	1	1	0	1	1	1	0	1.768	3.016	1.922	0.5862068966	1.5691987513	0.9198751301
CHR-122040	1	0	0	1	1	1	0	1	1	1	0	0	0	0	0	0	4.846	6.472	3.452	0.7487639061	1.8748551564	1.4038238702
CHR-122504	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	0	1.426	2.44	1.82	0.5844262295	1.3406593407	0.7835164835
CHR-125667a	1	0	0	1	1	1	0	0	0	1	1	0	1	1	0	0	5.872	5.464	3.758	1.074670571	1.4539648749	1.5625332624
CHR-125667b	1	1	0	1	1	0	0	1	0	1	1	0	1	0	0	0	2.281	3.16	2.84	0.7218354443	1.1126760563	0.8031690141
CHR-125681	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	1.768	2.872	2.432	0.6155988858	1.1809210526	0.7269736842

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
CHR-125774 α	1	1	0	1	0	1	0	1	0	1	1	0	0	0	0	0	4.504	3.448	2.432	1.3062645012	1.4177631579	1.8519736842
CHR-125774 β	1	0	0	1	1	0	0	0	0	1	1	0	0	1	0	0	2.11	2.728	1.514	0.7734604106	1.8018494055	1.393659181
CHR-127547	1	0	0	0	0	1	0	0	0	1	1	1	1	0	1	1	1.084	2.296	1.412	0.4721254355	1.6260623229	0.7677053824
CHR-127652	1	0	0	0	0	1	0	0	0	0	1	1	1	0	1	1	1.426	3.736	2.024	0.3816916488	1.8458498024	0.7045454545
CHR-140181	1	0	1	0	1	1	0	0	0	0	1	1	1	0	0	1	4.333	4.168	2.636	1.0395873321	1.5811836115	1.6437784522
CHR-140182	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	1	3.82	4.024	2.024	0.949304175	1.9881422925	1.8873517787
CHR-140204 α	1	0	0	1	1	1	0	0	1	0	1	1	1	1	1	1	3.649	3.448	2.84	1.0582946636	1.214084507	1.2848591549
CHR-140206	1	0	0	1	1	1	1	1	1	1	1	0	0	1	0	0	8.266	7.48	5.39	1.1050802139	1.387755102	1.533580705
CHR-140617 α	1	1	1	0	1	1	0	0	0	1	1	0	0	1	0	0	0.571	2.152	1.718	0.2653345725	1.2526193248	0.332363213
CHR-140617 β	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	1	0.742	1.864	1.616	0.3980686695	1.1534653465	0.4591584158
CHR-140826	1	0	0	0	1	0	0	0	0	1	1	1	1	1	1	0	1.255	2.44	1.616	0.51434442623	1.5099009901	0.7766089109
CHR-141055 α	0	0	0	0	0	1	0	0	0	0	1	0	1	0	1	0	1.255	6.616	4.676	0.1896916566	1.4148845167	0.2683917879
CHR-141066	1	0	1	0	1	1	0	0	1	1	1	1	1	1	1	0	10.318	8.776	6.614	1.1757064722	1.3268823707	1.5600241911
CHR-141415	1	0	1	0	1	1	0	0	1	1	1	1	1	0	0	0	1.426	5.176	3.248	0.2755023184	1.5935960591	0.4390394089
CHR-141693	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	1	1.255	2.872	1.616	0.4369777159	1.7772277228	0.7766089109
CHR-142520	1	0	1	0	1	1	1	0	0	1	1	1	1	1	0	0	13.054	10.936	7.838	1.1936722751	1.3952538913	1.6654758867
CHR-142803	1	0	0	0	1	0	0	1	0	1	1	1	1	0	1	1	1.768	3.304	2.024	0.5351089588	1.6324110672	0.8735177866
CHR-144532	1	0	0	0	1	1	0	0	0	1	1	1	1	0	1	1	5.53	4.888	2.024	1.1313420622	2.4150197628	2.7322134387
CHR-144897 α	1	0	1	0	0	0	0	0	0	1	1	1	0	0	1	1	3.136	3.16	1.922	0.9924050633	1.6441207076	1.6316337149
CHR-146792	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	1	1.255	3.448	1.514	0.3639791183	2.2774108322	0.8289299868
CHR-148771	1	0	0	0	1	1	0	1	1	1	1	1	1	0	1	1	2.11	7.336	3.554	0.2876226827	2.064153067	0.5936972425
CHR-148842	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	1	1.255	3.16	1.718	0.3971518987	1.8393480792	0.7305005821
CHR-149278 α	1	0	0	1	1	0	0	0	0	0	1	1	1	0	1	0	1.255	1.576	1.31	0.796319797	1.2030534351	0.9580152672
CHR-149278 β	1	0	0	0	1	0	0	1	0	0	1	1	1	0	1	1	0.4	1.432	0.8	0.2793296089	1.79	0.5
CHR-150017	1	0	0	0	1	0	0	0	0	1	1	1	1	0	1	1	1.255	3.736	2.228	0.3359207709	1.6768402154	0.5632854578
CHR-150259	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	1	1.255	6.472	2.33	0.1939122373	2.7776824034	0.5386266094
CHR-151345	1	0	1	0	1	1	0	0	1	1	1	0	1	1	0	0	3.136	5.176	3.86	0.6058732612	1.3409326425	0.8124352332
CHR-158245	1	0	0	0	1	0	0	1	0	1	1	1	1	0	1	1	0.742	1.432	1.004	0.5181564246	1.4262948207	0.7390438247
CHR-158248	1	0	1	0	1	1	0	1	1	0	1	0	0	1	1	1	3.991	5.608	3.35	0.71166619116	1.6740298507	1.1913432836
CHR-158250	1	1	0	1	0	1	0	0	1	1	0	0	0	0	0	0	5.017	4.6	2.942	1.0906251739	1.5635622026	1.7053025153
CHR-158459	1	0	0	0	1	1	0	1	0	1	1	1	1	0	1	1	3.82	4.312	1.718	0.8858998145	2.509895227	2.2235157159
CHR-166081	1	0	0	1	1	1	1	1	1	1	0	0	0	0	0	0	10.66	6.472	4.778	1.6470951792	1.3545416492	2.2310590205
CHR-166426 α	1	0	0	1	1	0	0	0	0	1	1	1	1	0	1	1	0.742	2.152	1.004	0.344795539	2.1434262948	0.7390438247
CHR-166427	1	1	0	1	1	0	0	1	0	0	1	0	1	1	0	0	1.939	4.312	2.738	0.4496753247	1.5748721695	0.7081811541
CHR-166428	1	0	0	1	1	0	0	1	0	0	1	0	0	1	0	0	1.939	3.304	2.024	0.5808644068	1.6324110672	0.9580039526
CHR-166699	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	1	1.255	2.44	1.514	0.5143442623	1.6116248349	0.8289299868
CHR-167002 α	1	1	0	1	1	1	0	0	1	1	0	0	0	1	0	0	4.162	4.456	2.942	0.934021544	1.5146159075	1.4146838885

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
CHR-167002 β	1	0	0	1	1	0	0	1	0	0	0	0	1	0	0	0	3.136	2.872	1.718	1.0919220056	1.6717112922	1.8253783469
CHR-167008 β	1	0	0	1	1	1	0	1	0	0	1	1	1	0	1	1	1.084	1.72	1.106	0.6302325581	1.5551537071	0.9801084991
CHR-168185	1	0	1	0	1	0	0	0	0	1	1	1	1	1	1	1	2.281	8.344	2.126	0.2733700863	3.9247412982	1.0729068674
CHR-168206	1	1	1	0	1	1	0	0	0	1	1	0	0	0	0	0	1.255	2.296	1.412	0.5466027875	1.6260623229	0.8888101983
CHR-168216	1	0	1	1	1	0	0	0	0	1	1	1	1	1	1	1	1.084	2.44	1.31	0.4442622951	1.8625954198	0.827480916
CHR-169524	1	0	0	1	1	0	0	0	0	0	1	1	1	0	1	0	2.452	4.744	3.044	0.5168634064	1.5584756899	0.8055190539
CHR-170595	1	0	0	0	1	0	0	1	0	0	1	1	1	0	1	1	2.794	1.432	2.33	1.9511173184	0.6145922747	1.1991416309
CHR-171448	1	0	0	0	0	0	0	1	0	0	1	1	1	1	1	1	1.939	3.448	1.718	0.5623549884	2.0069848661	1.1286379511
CHR-17221	1	1	0	1	0	1	1	0	1	1	1	0	0	0	1	0	5.017	4.024	2.636	1.2467693837	1.5265553869	1.903262519
CHR-173482	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	0	1.084	2.44	1.514	0.4442622951	1.6116248349	0.715984148
CHR-173500	1	0	0	1	1	0	0	0	0	0	1	0	1	1	1	0	3.478	4.456	2.738	0.7805206463	1.6274653031	1.2702702703
CHR-174942	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	1	0.742	2.152	1.82	0.344795539	1.1824175824	0.4076923077
CHR-175130	1	0	1	0	1	1	1	0	1	1	1	0	1	0	0	0	13.738	8.488	7.124	1.6185202639	1.1914654688	1.9284110051
CHR-175204	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	1	1.084	3.736	2.432	0.2901498929	1.5361842105	0.4457236842
CHR-17537	1	1	0	1	1	1	0	0	0	1	0	0	0	0	0	0	1.255	3.448	2.228	0.3639791183	1.5475763016	0.5632854578
CHR-176057	1	0	1	0	1	1	1	0	1	0	1	0	1	1	0	0	2.623	8.344	7.022	0.3143576222	1.1882654514	0.3735403019
CHR-17737	1	0	0	1	1	1	0	1	1	1	1	1	0	1	0	0	12.541	9.496	4.676	1.3206613311	2.0307955518	2.6819931565
CHR-179339	1	1	0	1	1	0	0	0	0	1	1	0	0	1	0	0	1.255	3.16	2.228	0.3971518987	1.4183123878	0.5632854578
CHR-182046	1	1	0	1	1	1	0	0	0	1	0	0	0	1	0	0	3.82	1.72	3.044	2.2209302326	0.5650459921	1.2549277267
CHR-182095	1	0	1	0	1	0	0	1	0	0	1	1	1	0	1	1	3.478	1.288	3.35	2.700310559	0.3844776119	1.0382089552
CHR-185926 α	1	0	1	0	1	0	0	0	0	0	1	0	1	0	0	0	0.913	2.584	1.412	0.3533281734	1.8300283286	0.6466005666
CHR-185926 β	1	0	0	1	1	0	0	0	0	0	1	0	1	1	1	0	1.255	2.872	2.024	0.4369777159	1.418972332	0.6200592885
CHR-18700 α	1	1	0	1	0	0	0	0	0	1	0	0	0	0	0	0	1.597	2.008	1.718	0.7953187251	1.1688009313	0.9295692666
CHR-188176 α	1	0	1	0	0	0	0	0	0	0	1	1	0	0	1	1	4.162	3.16	1.208	1.3170886076	2.6158940397	3.4453642384
CHR-188176 β	1	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	3.82	2.872	1.208	1.3300835655	2.3774834437	3.1622516556
CHR-189047	1	0	1	0	1	0	0	1	0	0	1	1	1	1	1	0	1.426	3.16	2.33	0.4512658228	1.356223176	0.6120171674
CHR-189360	1	0	0	1	1	0	0	0	0	0	1	0	0	1	0	0	2.965	3.736	2.942	0.7936295503	1.2698844324	1.007817811
CHR-190342	0	0	0	0	0	0	0	0	1	1	1	1	0	0	1	1	5.872	9.064	7.328	0.6478375993	1.2368995633	0.8013100437
CHR-19039	1	0	0	0	1	1	0	1	0	0	1	1	1	0	1	1	1.597	5.032	1.514	0.3173688894	3.3236459709	1.0548216645
CHR-194962	1	0	1	0	1	1	0	0	0	1	1	0	1	0	1	0	2.11	6.04	3.962	0.3493377483	1.5244825846	0.5325593135
CHR-195784 α , β	1	0	0	1	1	1	0	0	0	0	1	1	1	1	0	0	11.857	9.208	5.186	1.2876846221	1.7755495565	2.2863478596
CHR-197046	1	0	0	0	1	0	0	0	0	0	1	1	1	0	1	1	2.965	6.472	4.166	0.4581273177	1.5535285646	0.7117138742
CHR-199665	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	1	2.623	3.448	1.82	0.7607308585	1.8945054945	1.4412087912
CHR-203092	1	0	0	1	1	0	0	0	0	1	1	1	1	1	0	0	1.426	2.152	1.718	0.6626394052	1.2526193248	0.8300349243
CHR-203094	1	0	0	1	1	0	0	1	0	0	1	0	1	0	0	0	2.281	3.16	2.738	0.7218354443	1.1541271001	0.83030898466
CHR-20921 α	0	1	0	0	0	0	0	0	1	1	0	0	0	0	0	0	1.768	2.296	1.31	0.7700348432	1.7526717557	1.3496183206
CHR-21018	1	1	0	1	1	1	0	1	0	1	0	0	0	0	0	0	1.768	3.304	2.534	0.5351089588	1.3038674033	0.6977111287

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
CHR-21020	1	0	0	1	1	1	0	1	0	0	1	0	1	1	1	1	10.831	6.904	3.902	1.5688006952	1.7425542655	2.7337203433
CHR-21041	1	0	0	1	1	1	0	1	0	0	1	0	0	1	1	1	5.701	3.304	2.534	1.7254842615	1.3038674033	2.2498026835
CHR-21046 α	1	1	0	1	1	1	0	0	0	1	0	0	0	0	0	0	2.794	2.152	4.676	1.2983271375	0.4602224123	0.5975192472
CHR-21116 α	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1.939	3.16	2.126	0.6136075949	1.4863593603	0.9120413923
CHR-21116 β	0	0	0	0	0	0	0	0	0	0	1	1	1	0	1	1	1.597	2.872	1.514	0.5560584958	1.8969616909	1.0548216645
CHR-21123	1	1	0	1	1	0	0	0	0	1	1	1	1	1	0	0	1.939	3.88	2.33	0.499742268	1.6652360515	0.8321888412
CHR-2116y	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	1	1.084	2.152	1.208	0.5037174721	1.7814569536	0.8973509934
CHR-215907	1	0	1	0	1	1	1	0	1	1	1	1	1	0	0	0	2.281	8.632	5.798	0.2642493049	1.4887892377	0.3934115212
CHR-215943 α	1	0	1	0	1	1	1	0	0	0	1	1	1	0	0	0	2.794	5.464	3.452	0.5113469985	1.5828505214	0.8093858633
CHR-215943 β	1	0	1	1	1	1	0	0	0	0	1	1	1	0	0	0	1.939	3.016	2.636	0.6429045093	1.1441578149	0.7355842185
CHR-24173 α	1	0	1	0	1	0	0	0	0	1	1	1	1	1	1	1	1.768	2.44	1.718	0.7245901639	1.4202561118	1.0291036088
CHR-24176	1	0	0	1	1	1	1	1	1	1	0	0	1	1	0	0	14.935	7.192	5.39	2.0766129032	1.33432282	2.7708719852
CHR-24336	1	0	1	0	1	1	0	0	1	1	1	1	1	0	1	1	1.426	4.456	2.33	0.3200179533	1.9124463519	0.6120171674
CHR-28029	1	0	1	0	1	1	0	1	1	0	1	1	1	1	1	0	12.028	8.056	6.308	1.4930486594	1.2771084337	1.9067850349
CHR-28030	1	0	1	1	1	1	1	1	1	1	1	1	1	0	0	0	12.199	8.776	6.41	1.390041021	1.3691107644	1.9031201248
CHR-34905	1	0	0	0	1	0	0	0	0	1	1	1	1	0	1	1	1.255	5.032	2.432	0.2494038156	2.0690789474	0.5160361842
CHR-4457	1	0	1	0	1	0	0	0	0	1	1	0	1	0	0	0	1.939	4.168	2.84	0.4652111324	1.4676056338	0.6827464789
CHR-51275	1	1	0	1	1	0	0	0	0	1	1	0	1	0	0	0	2.281	3.736	2.126	0.6105460385	1.7572906867	1.0729068674
CHR-54360	1	0	1	1	1	1	0	0	0	1	1	1	1	0	0	0	1.255	2.296	2.228	0.5466027875	1.0305206463	0.5632854578
CHR-57504	1	0	1	0	1	1	1	1	1	1	1	0	0	1	0	0	7.753	11.08	10.388	0.6997292419	1.0666153254	0.746341933
CHR-58620	1	0	1	0	1	1	0	0	0	1	1	1	1	0	1	1	2.623	6.04	2.942	0.4342715232	2.053025153	0.8915703603
CHR-60315	1	0	1	0	1	0	0	0	1	0	1	1	1	0	1	1	1.255	1	1.718	1.255	0.5820721769	0.7305005821
CHR-68575	1	1	0	1	1	1	0	0	1	1	1	0	1	1	0	0	3.991	5.032	3.248	0.7931240064	1.5492610837	1.2287561576
CHR-69239	1	0	0	1	1	1	0	0	1	1	0	0	0	1	0	0	2.965	5.752	3.758	0.515472879	1.5306013837	0.7889835019
CHR-71231	1	1	0	1	1	1	0	0	0	1	0	0	0	0	0	0	1.426	2.296	1.718	0.6210801394	1.3364377183	0.8300349243
CHR-72560	1	0	1	0	1	1	1	0	1	0	1	1	0	1	0	0	2.281	9.064	5.288	0.2516548985	1.7140695915	0.4313540091
CHR-75778	1	0	1	0	1	0	0	0	0	1	1	1	1	1	1	1	1.255	2.296	1.922	0.5466027875	1.1945889698	0.6529656608
CHR-77140	1	0	1	0	1	1	0	1	1	0	1	1	0	1	1	1	10.318	8.056	4.166	1.2807845084	1.9337493999	2.4767162746
CHR-77684	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	0	1.084	2.44	1.82	0.4442622951	1.3406593407	0.5956043956
CHR-78817	1	0	0	1	1	0	0	1	1	0	1	1	1	1	1	1	2.965	2.872	2.636	1.0323816156	1.0895295903	1.1248103187
CHR-79648	1	0	0	1	1	1	0	1	1	0	1	1	1	1	1	1	5.53	3.448	2.432	1.6038283063	1.4177631579	2.2738486842
CHR-79667	1	0	0	1	1	0	0	0	0	1	1	0	1	0	1	0	3.136	3.16	2.228	0.9924050633	1.4183123878	1.407540395
CHR-79669	1	1	0	1	1	1	0	1	1	1	0	0	0	1	0	0	1.939	1.576	2.738	1.2303299492	0.5756026297	0.7081811541
CHR-80877 α	1	0	0	0	1	0	0	1	0	0	1	1	1	0	1	1	1.255	1.864	1.004	0.6732832618	1.8565737052	1.25
CHR-80877 β	1	0	1	0	1	0	0	1	0	0	1	1	1	0	1	1	1.084	1.864	0.902	0.5815450644	2.066518847	1.2017738359
CHR-81808 α	1	0	0	1	1	0	0	0	0	1	1	1	1	1	1	0	1.708	2.152	1.106	0.8215613383	1.9457504521	1.5985533454
CHR-82507	1	0	1	0	1	1	0	0	1	1	1	1	1	1	0	0	6.556	10.936	7.022	0.5994879298	1.5573910567	0.936371404

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
CHR-85617	1	1	0	1	1	1	0	1	0	1	0	0	0	0	0	0	1.255	2.44	1.922	0.5143442623	1.2695109261	0.6529656608
CHR-90101	1	0	0	1	1	1	0	1	1	1	1	0	0	1	0	0	4.504	4.456	3.35	1.0107719928	1.3301492537	1.3444776119
CHR-94341	1	0	1	0	1	1	0	0	1	1	1	1	1	1	0	0	3.82	8.632	6.512	0.4425393883	1.3255528256	0.5866093366
CHR-97829	1	0	1	0	1	1	0	1	1	0	1	0	1	1	0	0	4.675	9.352	7.43	0.499893071	1.2586810229	0.6292059219
CHR-9924	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	1	3.136	2.872	2.636	1.0919220056	1.0895295903	1.1896813354
DeptAgr-1154	1	0	1	0	1	0	0	0	0	1	1	1	1	1	1	0	4.846	4.168	1.616	1.1626679463	2.5792079208	2.9987623762
OTA-002597	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	1	1.939	3.592	1.922	0.5398106904	1.8688865765	1.0088449532
OTA-002599	1	0	1	0	1	1	0	0	1	1	1	0	1	1	0	0	9.463	8.488	5.492	1.114868049	1.5455207575	1.7230517116
OTA-002608	1	0	0	1	1	1	1	0	0	0	0	0	0	0	0	0	6.385	4.312	2.738	1.4807513915	1.5748721695	2.3319941563
OTA-002614	1	0	0	1	1	1	1	1	1	1	1	0	1	0	1	0	10.318	6.184	6.308	1.6684993532	0.9803424223	1.6357006975
OTA-003270	1	0	1	0	1	1	0	1	1	0	1	1	1	1	1	1	3.649	2.728	2.33	1.3376099707	1.1708154506	1.5660944206
OTA-003351	1	0	1	0	1	0	0	1	1	0	1	1	0	1	1	1	1.426	2.296	1.514	0.6210801394	1.5165125495	0.9418758256
OTA-003516 α	0	0	0	0	0	0	0	0	0	1	1	1	0	0	1	1	4.162	4.312	2.534	0.9652133581	1.7016574586	1.6424625099
OTA-003535 α	0	1	1	0	0	0	0	0	0	1	1	1	0	0	1	0	1.255	2.008	1.718	0.625	1.1688009313	0.7305005821
OTA-004203	1	1	0	1	1	1	0	0	0	1	0	0	0	0	0	0	1.426	2.872	1.82	0.4965181058	1.578021978	0.7835164835
OTA-004246	1	1	0	1	1	1	0	0	0	1	0	0	0	0	0	0	5.017	5.752	2.534	0.8722183588	2.2699289661	1.9798737174
OTA-006305	1	0	0	0	0	0	0	0	0	0	1	1	1	0	1	1	2.281	3.448	2.636	0.6615429234	1.3080424886	0.8653262519
OTA-006362	1	0	0	1	1	1	1	0	1	1	0	0	0	0	0	0	3.649	6.616	3.656	0.551541717	1.8096280088	0.9980853392
OTA-007200	1	0	0	1	1	0	0	0	0	0	1	0	1	1	1	0	3.136	3.304	1.718	0.9491525424	1.9231664726	1.8253783469
OTA-007959	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	0	1.939	3.448	2.024	0.5623549884	1.7035573123	0.9580039526
OTA-008534	1	0	1	0	1	0	0	1	0	1	1	1	1	1	1	1	2.281	3.592	2.534	0.6350222717	1.4175217048	0.9001578532
OTA-009188	1	0	0	1	1	1	1	0	1	1	1	1	0	0	0	0	1.768	3.448	1.82	0.5127610209	1.8945054945	0.9714285714
OTA-009839	1	1	1	0	1	1	0	1	1	0	1	0	1	0	0	0	5.188	3.736	2.228	1.3886509636	1.6768402154	2.328545781
OTA-010368	1	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	1.939	3.448	1.82	0.5623549884	1.8945054945	1.0653846154
OTA-015584	1	0	0	1	1	1	0	0	1	1	1	0	0	1	0	0	12.541	5.752	3.248	2.1802851182	1.7709359606	3.8611453202
OTA-017356	1	1	0	1	1	1	1	1	1	0	0	0	0	1	0	0	5.53	5.752	4.472	0.9614047288	1.2862254025	1.2365831843
OTA-017828	1	1	0	1	1	1	1	1	1	1	0	0	1	0	0	0	16.132	11.512	8.756	1.4013203614	1.3147555962	1.8423937871
OTA-018264	1	0	0	0	1	0	0	0	0	0	1	1	1	1	1	1	2.281	3.016	1.922	0.7562997347	1.5691987513	1.1867845994
OTA-020589	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	0	2.452	3.448	2.432	0.711136891	1.4177631579	1.0082236842
OTA-020760	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	1	5.701	4.024	2.534	1.416749503	1.5880031571	2.2498026835
OTA-020768	1	0	0	1	1	1	0	1	1	0	1	1	0	1	1	1	1.939	3.448	2.228	0.5623549884	1.5475763016	0.8702872531
OTA-020943	1	0	0	1	1	0	0	0	0	0	1	1	1	0	1	1	2.965	2.872	1.514	1.0323816156	1.8969616909	1.9583883752
OTA-020958	1	0	0	1	1	1	1	0	0	0	1	0	1	1	1	0	4.846	5.176	3.758	0.930244204	1.3773283662	1.2895156998
OTA-021820	1	0	0	1	1	0	0	1	1	0	1	1	1	1	1	1	3.307	3.016	2.228	1.0964854111	1.3536804309	1.4842908438
OTA-022262	1	0	0	1	1	1	1	1	1	1	1	0	0	0	0	0	13.567	9.496	6.614	1.4287068239	1.4357423647	2.0512549138
OTA-022944	1	0	0	1	1	1	1	1	1	0	1	0	0	0	0	0	5.017	1.432	4.37	3.5034916201	0.3276887872	1.1480549199
OTA-025666	1	0	0	1	1	1	0	1	1	0	1	0	1	1	1	0	1.768	2.584	1.922	0.6842105263	1.3444328824	0.9198751301

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
OTA-026027	1	0	0	1	0	1	0	0	0	1	0	0	0	0	0	0	9.292	4.744	2.84	1.9586846543	1.6704225352	3.2718309859
OTA-026393	1	0	0	1	0	1	0	0	0	1	0	0	0	0	0	0	4.162	4.168	2.33	0.9985604607	1.7888412017	1.7862660944
OTA-026394	1	0	0	1	1	0	0	0	0	1	1	1	1	1	1	1	1.597	2.728	1.412	0.5854105572	1.9320113314	1.13101983
OTA-027970	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	1	1.426	2.152	1.412	0.6626394052	1.5240793201	1.0099150142
RDS-470	1	0	1	0	1	1	0	0	0	0	1	1	1	1	0	0	4.04	7.73	4	0.5226390686	1.9325	1.01
RDS-473	1	0	1	0	1	1	1	0	1	1	1	1	1	0	0	0	4.01	9.67	7.01	0.4146845915	1.3794579173	0.5720399429
RDS-478	1	0	1	0	1	1	1	0	1	1	1	1	1	1	0	0	10.27	11.66	7.41	0.8807890223	1.5735492578	1.3859649123
RDS-488	1	0	1	0	1	1	1	0	1	1	1	1	1	0	0	0	5.07	12.44	7.35	0.4075562701	1.6925170068	0.6897959184
RDS-493	1	0	1	0	1	1	0	1	1	1	1	1	0	1	0	0	8.14	12.8	8.98	0.6359375	1.425389755	0.9064587973
RDS-498	1	0	1	0	0	1	0	0	0	1	1	1	0	1	1	1	1.01	4.34	2.4	0.232718894	1.8083333333	0.4208333333
RDS-510	1	1	0	1	1	0	0	0	0	1	0	0	0	0	0	0	1.56	2.58	1.92	0.6046511628	1.34375	0.8125
RDS-515	1	0	1	0	1	0	0	0	0	0	1	1	1	1	0	0	1.04	2.57	1.56	0.4046692607	1.6474358974	0.6666666667
RDS-522	0	1	0	1	0	0	0	0	0	1	1	1	0	0	0	0	0.41	1.84	1.74	0.222826087	1.0574712644	0.2356321839
RDS-524	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1.88	6.11	3	0.3076923077	2.0366666667	0.6266666667
RDS-682	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	1	1.97	2.33	0.97	0.8454935622	2.4020618557	2.0309278351
RDS-683	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	1	1.07	1.31	0.94	0.8167938931	1.3936170213	1.1382978723
RDS-689	1	0	1	1	1	1	1	0	0	0	1	1	0	1	1	1	6.43	6.75	3.22	0.9525925926	2.0962732919	1.9968944099
RDS-698	1	0	0	0	1	1	0	0	0	0	1	1	1	0	1	1	2.89	4.11	1.74	0.703163017	2.3620689655	1.6609195402
RDS-704	0	1	0	0	0	0	0	0	0	0	1	0	0	0	1	0	6.78	8.08	4.92	0.8391089109	1.6422764228	1.3780487805
RDS-707c2	1	0	0	1	1	1	0	0	0	1	1	0	1	1	0	0	1.2	2.23	1.32	0.5381165919	1.6893939394	0.9090909091
RDS-707c3	1	0	0	1	1	1	0	0	0	1	1	0	1	0	0	0	0.79	2.58	1.79	0.3062015504	1.4413407821	0.4413407821
RDS-708	1	0	0	1	1	1	0	0	0	1	0	0	0	0	0	0	0.4	3.12	1.98	0.1282051282	1.5757575758	0.202020202
RDS-710	1	0	1	0	1	1	0	0	0	0	1	1	0	1	1	1	1.25	4.25	2.46	0.2941176471	1.7276422764	0.5081300813
RDS-740	1	0	0	1	0	0	0	0	0	0	1	1	1	0	1	0	0.72	1.73	1.1	0.4161849711	1.5727272727	0.6545454545
RDS-741	1	0	0	1	0	0	0	0	0	0	1	1	1	0	1	1	0.47	1.78	1.89	0.2640449438	0.9417989418	0.2486772487
RDS-742	1	0	0	1	0	0	0	0	0	0	1	1	1	0	1	1	0.8	1.51	1.53	0.5298013245	0.9869281046	0.522875817
RDS-744	1	1	0	1	1	0	0	0	0	1	1	1	1	0	0	0	1.55	2.78	2.54	0.5575539568	1.094488189	0.6102362205
TM-Art	1	0	1	0	1	1	0	0	0	0	1	1	0	0	1	1	1.52	4.46	2.79	0.3408071749	1.5985663082	0.5448028674
TM-BG	1	1	1	1	0	1	1	0	1	1	1	1	1	0	0	0	3.11	7.45	5.02	0.4174496644	1.484063745	0.6195219124
TM-BL	1	0	1	1	1	1	0	1	1	0	1	1	1	1	1	1	5.95	5.6	4	1.0625	1.4	1.4875
TM-BVTD1	1	0	0	0	0	0	0	0	0	1	1	1	1	0	1	1	1.55	3.11	1.53	0.498392283	2.0326797386	1.0130718954
TM-BVTD2	1	0	0	0	1	0	0	1	0	1	1	1	1	1	1	1	1.44	2.128	1.16	0.6766917293	1.8344827586	1.2413793103
TM-BVTD3	1	0	0	0	1	0	0	1	0	0	1	1	1	1	1	1	1.42	1.92	0.91	0.7395833333	2.1098901099	1.5604395604
TM-BVTW1	1	0	0	0	0	0	0	1	0	1	1	1	1	0	1	1	3.18	3.77	1.52	0.8435013263	2.4802631579	2.0921052632
TM-BVTW2	1	0	1	0	1	0	0	0	0	1	1	1	1	1	1	0	2.56	2.47	0.58	1.036437247	4.2586206897	4.4137931034
TM-CP	1	0	0	1	1	1	0	0	1	1	1	1	1	1	1	1	2.44	2.92	2.09	0.8356164384	1.3971291866	1.1674641148
TM-Fs	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	1	6.15	4.02	2.14	1.5298507463	1.8785046729	2.8738317757

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
TM-Gdy-1	0	1	0	0	0	0	0	0	0	1	1	1	0	0	1	0	5.88	5.86	2.82	1.0034129693	2.0780141844	2.085106383
TM-Gdy-2	1	0	1	0	0	0	0	0	0	0	1	0	0	0	1	0	1.71	3.12	1.63	0.5480769231	1.9141104294	1.0490797546
TM-Gdy-3	1	1	0	1	0	1	0	1	0	0	1	1	0	0	1	0	0.32	2.63	1.66	0.1216730038	1.5843373494	0.1927710843
TM-Gdy-4	0	1	1	1	0	0	0	0	0	0	1	1	0	0	1	0	1.94	3.52	2.28	0.5511363636	1.5438596491	0.850877193
TM-HR	1	0	1	0	1	1	0	1	1	0	1	1	1	0	0	0	1.98	3.59	2.55	0.5515320334	1.4078431373	0.7764705882
TM-HT	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	1	2.22	4.76	2.57	0.4663865546	1.8521400778	0.8638132296
TM-KS	1	0	1	1	1	0	0	0	0	0	1	1	1	1	0	1	0.63	1.74	1.15	0.3620689655	1.5130434783	0.547826087
TM-KWK1	1	0	1	0	1	1	0	0	0	1	1	1	1	1	0	0	0.64	4.69	3.49	0.1364605544	1.3438395415	0.1833810888
TM-KWK3	1	0	1	0	1	1	0	0	1	1	1	1	1	1	0	0	2.48	9.46	4.77	0.2621564482	1.9832285115	0.5199161426
TM-LS	1	0	1	1	0	1	0	0	0	1	1	1	1	1	1	1	1.11	3.92	2.33	0.2831632653	1.6824034335	0.4763948498
TM-Lw	0	0	0	0	0	0	0	0	0	1	1	1	1	0	1	1	1.89	2.66	1.61	0.7105263158	1.652173913	1.1739130435
TM-Mau	1	0	1	0	1	1	0	0	0	0	1	1	1	0	1	1	2.352	4.55	2.16	0.5169230769	2.1064814815	1.0888888889
TM-Mbn	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	1	0.26	1.82	1.24	0.1428571429	1.4677419355	0.2096774194
TM-MBT1	1	0	0	1	1	0	0	0	0	0	1	1	1	0	1	0	0.42	2	1.47	0.21	1.3605442177	0.2857142857
TM-MBT2	1	0	0	1	1	0	0	1	0	1	1	1	0	0	1	1	2.54	3.26	2.15	0.7791411043	1.5162790698	1.1813953488
TM-MBT3	1	0	1	0	1	0	0	0	1	1	1	1	1	1	0	0	0.833	2.95	2.35	0.2823728814	1.2553191489	0.3544680851
TM-MBT4	1	0	0	0	1	0	0	0	0	0	1	1	1	0	1	1	1.47	3.36	1.84	0.4375	1.8260869565	0.7989130435
TM-MCV1	1	0	0	1	1	1	0	0	1	1	0	0	0	0	0	0	8.52	9.95	4.65	0.856281407	2.1397849462	1.8322580645
TM-MCV2	1	0	0	1	1	1	1	1	0	1	0	0	0	0	0	0	5.53	4.26	2.65	1.2981220657	1.6075471698	2.0867924528
TM-MCV3	1	0	0	1	1	1	1	1	0	0	1	0	0	1	0	1	6.63	6.13	3.4	1.0815660685	1.8029411765	1.95
TM-Mfy	1	0	1	0	1	1	1	1	1	1	1	1	1	1	0	0	1.95	14.35	11.75	0.1358885017	1.2212765957	0.1659574468
TM-OTV1	1	0	0	0	0	0	0	1	0	1	1	1	0	1	1	1	2.18	1.82	1.11	1.1978021978	1.6396396396	1.963963964
TM-Ra	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	0	1.05	3.3	1.45	0.3181818182	2.275862069	0.724137931
TM-RaG	1	0	1	0	1	1	0	0	1	1	1	1	1	0	0	0	0.88	4.74	3.44	0.1856540084	1.3779069767	0.2558139535
TM-Rd-lower	0	0	1	0	0	1	0	0	1	1	1	1	0	0	1	1	2.47	6.97	3.65	0.3543758967	1.9095890411	0.6767123288
TM-Rd-upper	1	0	1	0	1	1	0	0	1	0	1	1	1	1	1	1	3.04	6.31	4.42	0.4817749604	1.42760181	0.6877828054
TM-RH	1	0	1	0	1	1	0	0	1	0	1	1	0	1	1	0	0.9	3.91	2.81	0.2301790281	1.3914590747	0.3202846975
TM-Rly	1	0	1	0	1	1	0	1	1	0	1	1	0	1	1	1	7.276	7.17	4.54	1.0147838215	1.5792951542	1.6026431718
TM-RP	1	0	0	0	1	0	0	0	0	0	1	1	1	0	1	0	1.81	2.69	1.72	0.6728624535	1.5639534884	1.0523255814
TM-RR-inf	1	0	1	0	1	1	1	0	1	1	1	1	1	0	0	0	7.07	8.95	6.92	0.7899441341	1.2933526012	1.0216763006
TM-Rs	1	0	0	1	1	1	0	0	1	0	1	1	1	0	1	1	1.06	2.43	1.97	0.4362139918	1.2335025381	0.538071066
TM-Ruk	1	0	1	0	1	1	0	0	1	1	1	1	1	0	0	0	5.47	9.12	6.06	0.5997807018	1.504950495	0.902640264
TM-SB	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	1	2.24	5.37	1.96	0.417132216	2.7397959184	1.1428571429
TM-TH	1	0	0	0	1	0	0	0	0	1	1	1	1	0	1	0	0.603	2.39	1.64	0.2523012552	1.4573170732	0.3676829268
TM-WD	1	0	1	0	1	1	0	0	0	1	1	1	1	1	1	1	1.5	3.9	2.1	0.3846153846	1.8571428571	0.7142857143
TM-WG	1	1	0	1	0	1	1	0	1	1	1	1	0	0	0	0	17.16	9.79	7.91	1.7528089888	1.2376738306	2.1694058154
TYPE:albidulees	1	0	1	0	1	1	1	1	1	0	1	0	1	1	0	0	17.5	10.648	6.512	1.643501127	1.6351351351	2.6873464373

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
TYPE.bel.J.ang	1	0	0	0	1	1	1	1	0	1	1	1	1	0	1	0	2.281	6.04	2.228	0.3776490066	2.710951526	1.0237881508
TYPE.bel.v.cras	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	0	1.939	3.736	1.922	0.5190042827	1.9438085328	1.0088449532
TYPE.bel.v.glab. α	1	0	0	1	1	0	0	1	0	0	1	1	1	1	1	1	2.281	2.44	3.044	0.9348360656	0.8015768725	0.7493429698
TYPE.bel.v.glab. β	1	0	1	0	0	1	0	0	0	1	1	1	1	1	0	0	1.084	3.16	2.228	0.3430379747	1.4183123878	0.486535009
TYPE.bel.v.orbi	1	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	2.452	2.872	2.432	0.8537604457	1.1809210526	1.0082236842
TYPE.bel.v.seto	1	0	0	1	1	0	0	0	0	1	1	1	1	1	1	1	5.017	5.032	3.452	0.9970190779	1.4577056779	1.4533603708
TYPE.cochleas	1	0	1	0	1	1	1	1	1	0	1	1	1	1	1	0	9.976	15.4	3.962	0.6477922078	3.8869257951	2.5179202423
TYPE.haastii	1	0	0	1	1	1	1	0	0	0	0	0	0	1	0	0	15.79	10.792	8.348	1.4631208302	1.2927647341	1.891471011
TYPE.saxifragoides	1	1	1	0	1	1	0	0	1	1	1	0	0	1	0	0	3.478	10.648	7.838	0.3266341097	1.3585098239	0.4437356468
TYPE.southlandica	1	0	1	0	1	1	0	1	1	1	1	1	1	1	1	1	4.675	3.016	4.268	1.550066313	0.7066541706	1.0953608247
WELT-31825	1	0	0	1	1	0	0	0	0	0	1	0	1	1	1	0	1.768	4.168	2.228	0.424184261	1.8707360862	0.7935368043
WELT-31827	1	0	0	0	0	1	0	0	0	1	1	1	1	0	1	1	2.281	4.168	2.024	0.5472648752	2.0592885375	1.1269762846
WELT-31831	1	0	1	0	1	0	0	0	0	0	1	1	1	0	1	1	1.426	3.736	1.208	0.3816916488	3.0927152318	1.1804635762
WELT-31837	1	0	1	0	1	1	1	0	1	1	1	1	0	1	0	0	5.53	13.24	11	0.417673716	1.2036363636	0.5027272727
WELT-39004	1	0	1	1	1	0	0	1	0	1	1	1	0	1	1	1	3.307	2.44	2.126	1.3553278689	1.1476952023	1.5555032926
WELT-476	1	0	1	0	1	1	0	0	1	1	1	0	0	0	0	0	4.333	9.928	6.614	0.4364423852	1.5010583611	0.6551254914
WELT-482	1	0	1	0	1	1	0	0	0	1	1	0	1	0	0	0	3.478	7.048	2.84	0.4934733258	2.4816901408	1.2246478873
WELT-502	1	0	1	0	1	1	1	0	1	1	1	0	1	0	0	0	2.281	10.936	10.388	0.2085771763	1.0527531767	0.2195802849
WELT-506	1	0	0	1	1	0	0	1	0	1	1	1	1	1	0	0	1.426	2.872	2.228	0.4965181058	1.289048474	0.6400359066
WELT-507	1	0	0	0	0	1	0	0	0	1	1	0	1	0	1	0	4.333	7.48	4.676	0.5792780749	1.5996578272	0.9266467066
WELT-509	0	1	0	1	0	0	0	0	0	1	1	0	0	0	1	0	3.649	6.04	3.35	0.6041390728	1.8029850746	1.0892537313
WELT-517	1	0	1	0	0	0	0	0	0	1	1	1	1	0	1	1	2.965	4.168	1.616	0.7113723608	2.5792079208	1.8347772277
WELT-527	1	0	1	0	0	0	0	0	0	0	1	1	1	0	1	1	5.359	4.456	2.84	1.2026481149	1.5690140845	1.886971831
WELT-529	1	0	1	0	1	1	1	1	1	0	1	1	0	0	1	0	8.266	7.768	4.676	1.0641091658	1.6612489307	1.7677502139
WELT-532	1	0	1	0	1	1	0	0	0	1	1	1	1	0	1	1	1.255	3.88	1.82	0.3234536082	2.1318681319	0.6895604396
WELT-5433	1	0	1	0	1	1	0	0	0	1	1	1	1	0	1	1	1.426	3.736	1.82	0.3816916488	2.0527472527	0.7835164835
WELT-5439	1	0	1	0	0	0	0	0	0	0	1	1	1	0	1	1	2.623	2.728	0.902	0.9615102639	3.0243902439	2.9079822616
WELT-543 α	1	0	0	1	1	0	0	0	0	0	1	0	1	1	0	0	1.597	2.872	2.126	0.5560584958	1.3508936971	0.7511759172
WELT-5442	1	0	0	0	1	0	0	1	0	1	1	1	1	0	1	1	1.426	2.44	1.106	0.5844262295	2.2061482821	1.2893309222
WELT-545	1	1	1	0	1	0	0	0	0	1	1	1	1	0	0	0	0.742	3.448	2.024	0.2151972158	1.7035573123	0.3666007905
WELT-5450	1	0	1	0	1	1	1	0	0	1	1	1	1	1	1	1	13.054	9.064	3.656	1.4402030009	2.4792122538	3.5705689278
WELT-5459	1	0	1	0	1	0	0	0	0	1	1	0	1	1	0	0	1.939	4.312	3.554	0.4496753247	1.2132808104	0.5455824423
WELT-547 α	1	0	1	0	1	1	0	0	0	0	1	1	1	0	1	0	3.82	3.448	2.534	1.1078886311	1.3606945541	1.5074980268
WELT-547 β	1	0	1	0	1	0	0	0	0	0	1	1	1	1	1	1	2.623	3.592	2.228	0.730233853	1.6122082585	1.1772890485
WELT-548 α	1	0	0	1	1	0	0	0	0	0	1	1	1	1	1	1	1.255	1.864	1.31	0.6732832618	1.4229007634	0.9580152672
WELT-557	1	0	1	0	1	0	0	0	0	0	1	0	1	0	0	0	4.675	6.616	3.86	0.7066203144	1.7139896373	1.2111398964
WELT-558	1	0	0	1	1	1	0	1	0	0	1	1	1	0	0	1	2.452	3.304	1.922	0.7421307506	1.7190426639	1.2757544225

Specimen ID	01	02	03	04	05	06	07	08	09	10	11	12	13	14	15	16	19	20	21	22	23	24
WELT-559	1	0	0	1	1	0	0	0	0	1	1	0	1	0	0	0	1.426	2.44	2.024	0.5844262295	1.2055335968	0.7045454545
WELT-560	1	0	0	0	1	0	0	1	0	1	1	1	1	1	1	1	1.426	2.872	1.922	0.4965181058	1.494276795	0.7419354839
WELT-561	1	1	0	0	0	0	0	0	0	1	1	1	0	0	0	1	2.623	3.736	2.432	0.7020877944	1.5361842105	1.0785361842
WELT-563	1	0	1	0	1	0	0	1	0	1	1	1	1	1	1	0	1.426	2.872	1.82	0.4965181058	1.578021978	0.7835164835
WELT-568	1	0	1	0	1	0	0	0	0	1	1	1	1	0	1	1	1.255	2.44	1.208	0.5143442623	2.0198675497	1.0389072848
WELT-570	1	0	0	1	1	0	0	0	0	0	1	0	1	1	0	0	2.965	3.448	2.738	0.8599187935	1.2593133674	1.0829072316
WELTU-5572	1	0	0	0	1	0	0	0	0	0	1	1	1	0	1	1	3.991	4.168	2.738	0.9575335893	1.5222790358	1.457633309

Appendix B

Genetic Marker Presence/Absence Data

Allele presence/absence data from nine dominant markers for 273 specimens. Collector ID is shown for each specimen. Presence or absence of an allele is indicated by a '1' or '0' respectively. Alleles are labelled by marker name (see table 3.2) and average fragment length. Missing data indicates that a marker was not successfully amplified for that individual.

Part 1

Population	Sample	P12.184.3	P12.185.3	P12.186.3	P12.187.3	P12.188.5	Smht04.202	Smht04.204.7	Smht04.205.6	Smht04.206.6	Smht04.207.3	Smht04.208.4	Smht04.209.3	Smht04.210.6	Smht04.211.8	Smht04.213.9	Smht04.215.9	Smht04.216.7	Smht04.217.9	Smht04.218.9	Smht04.219.9	Smht04.221.2	Smht04.230.5	Smht04.258.6
1	Kwk2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
1	Kwk3	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
1	Kwk4	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
1	Kwk5	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
1	Kwk6	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
1	Kwk7	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
2	RgG1	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
2	RgG3	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
2	RgG5	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
2	RgG7	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
2	RgG10	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
2	RgG9	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
3	HpR1	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
3	HpR2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
3	HpR3							0	0	0	1	0	1	0	1	0	0	0	0	0	0	0	0	0
3	HpR4	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
3	HpR5	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
3	HpR7	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
4	RtH1	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
4	RtH2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
4	RtH3	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
4	RtH4	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
4	RtH5	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0

[illegible]

Population	Sample	P12.184.3	P12.185.3	P12.186.3	P12.187.3	P12.188.5	Smht04.202	Smht04.204.7	Smht04.205.6	Smht04.206.6	Smht04.207.3	Smht04.208.4	Smht04.209.3	Smht04.210.6	Smht04.211.8	Smht04.213.9	Smht04.215.9	Smht04.216.7	Smht04.217.9	Smht04.218.9	Smht04.219.9	Smht04.221.2	Smht04.230.5	Smht04.258.6
14	Lew698	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
14	Lew700	0	1	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
14	Lew701	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
14	Lew702	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
14	Lew703	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
14	Lew699	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
15	Den690	0	0	0	1	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	1	0	0	0
15	Den693	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
15	Den694	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
15	Den697	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
15	Den698	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
15	Den691	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
16	Cha674	0	0	0	1	0	0	0	0	1	0	1	0	1	0	0	0	0	0	0	0	0	0	0
16	Cha675	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
16	Cha676	0	0	0	1	0	0	0	0	1	0	1	0	1	0	0	0	0	0	0	0	0	0	0
16	Cha681	0	0	0	1	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0
16	Cha682	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
16	Cha677	0	0	0	1	0	0	0	0	1	0	1	0	1	0	0	0	0	0	0	0	0	0	0
16	Cha678	0	1	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
17	Swp733	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
17	Swp734	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	1	0	0
17	Swp735	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	1	0	1	0	0
17	Swp736	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	1	0	0
17	Swp738	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	1	0	0
17	Swp737	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	1	0	0	1	0	0	0	0
18	OtV2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
18	OtV3	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
18	OtV6	0	1	0	1	0	0	1	0	1	0	0	0	1	0	0	1	0	0	0	1	0	0	0
18	OtV7	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
18	OtV8	0	1	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
18	OVT10	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
19	BVTD1	0	1	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
19	BVTD2	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	1	0	0
19	BVTD3	0	1	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	1	0	0
19	BVTW1	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
19	BVTW2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
19	BVTW3	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
20	MBT1	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
20	MBT2	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	1	0	0
20	MBT3	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	1	0	0
20	MBT4	0	1	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
20	MBT5	0	0	0	1	0	0	1	0	1	0	0	0	1	0	0	0	0	0	0	1	1	0	0
20	MBT6	0	0	0	1	0	0	0	1	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
20	MBT7	0	1	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0
21	Cas498	0	0	1	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	1	0	0
21	Cas499	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	1	0	0
21	Cas500	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0
22	CvS510	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
22	CvS511	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
22	CvS512	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
22	CvS513	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
22	CvS514	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
23	PoP515	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	1	0	0
23	PoP516	0	1	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
23	PoP517	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
23	PoP518	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0

Population	Sample	P12.184.3	P12.185.3	P12.186.3	P12.187.3	P12.188.5	Smht04.202	Smht04.204.7	Smht04.205.6	Smht04.206.6	Smht04.207.3	Smht04.208.4	Smht04.209.3	Smht04.210.6	Smht04.211.8	Smht04.213.9	Smht04.215.9	Smht04.216.7	Smht04.217.9	Smht04.218.9	Smht04.219.9	Smht04.221.2	Smht04.230.5	Smht04.258.6
23	PoP519	0	0	1	0	0	0	0	0	1	0	1	0	1	0	0	0	0	0	0	1	0	0	0
23	MCV8	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
24	MSm520	0	0	0	1	0	0	0	0	1	0	0	1	1	1	0	0	0	0	0	0	0	0	0
24	Msm523	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
24	Msm521	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
24	MSm522	0	0	0	1	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	1	0	0	0
24	MSm524	0	0	0	1	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	1	0	0	0
25	PrG1	0	0	1	0	1	0	1	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
25	PrG2	0	0	1	0	0	0	1	0	1	0	0	0	1	0	0	0	0	0	0	1	1	0	0
25	PrG3	0	0	1	0	1	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
25	PrG4	0	0	0	0	1	0	1	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
25	PrG5	0	0	1	0	1	0	1	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
26	Gdy1	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
26	Gdy2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
26	Gdy3	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0
26	Gdy4	0	0	0	1	0	0	1	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
26	Gdy5	0	1	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	1	0	0
26	Gdy6	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
26	Gdy7	0	0	1	0	0	0	1	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
27	PHSL493	0	1	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	1	0	0	0	0	0
27	PHSL494	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
27	PHSL495	0	0	0	1	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0
27	PHSL496	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
27	PHSL497	0	0	0	1	0	0	0	0	0	0	0	1	1	0	0	0	0						

Population	Sample	P12.184.3	P12.185.3	P12.186.3	P12.187.3	P12.188.5	Smht04.202	Smht04.204.7	Smht04.205.6	Smht04.206.6	Smht04.207.3	Smht04.208.4	Smht04.209.3	Smht04.210.6	Smht04.211.8	Smht04.213.9	Smht04.215.9	Smht04.216.7	Smht04.217.9	Smht04.218.9	Smht04.219.9	Smht04.221.2	Smht04.230.5	Smht04.258.6
34	BLd2	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
34	BLd5	0	0	0	1	0	0	0	0	1	0	0	0	1	0	1	0	0	0	0	1	0	0	0
34	BLd6	0	0	0	1	0	0	0	0	1	0	0	0	1	0	1	0	0	0	0	1	0	0	0
34	BLd7	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
34	BLd3	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
34	BL4	0	1	0	0	0	0	0	0	1	0	0	0	1	0	0	1	0	0	0	1	0	0	0
35	Rks6	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	1	0	0	0	1	0	0	0
35	Rks1	0	0	0	1	0	0	0	0	1	1	0	0	1	0	0	0	0	0	0	1	0	0	0
35	Rks2	0	0	1	0	0	0	0	0	1	0	0	1	1	0	0	1	0	0	0	0	0	0	0
35	Rks3	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
35	Rks4	0	0	0	1	0	0	0	0	1	1	0	1	0	0	0	0	0	0	0	1	0	0	0
35	Rks5	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
36	Wye1	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
36	Wye2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
36	Wye3	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	1	0	0	0	1	0	0	0
37	KyS2	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0
37	KyS3	0	0	0	1	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	1	0	0	0
37	KyS4	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
37	KyS5	0	1	0	1	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	1	0	0	0
37	KyS6	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
37	KyS7	0	0	0	1	0	0	0	1	0	0	0	0	1	0	1	0	0	0	0	1	0	0	0
38	MBn1	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0
38	MBn6	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0
38	MBn7	0	0	0	1	0	0	0	0	0	0	0	1	1	0	1	0	0	0	0	1	0</		

Population	Sample	P12.184.3	P12.185.3	P12.186.3	P12.187.3	P12.188.5	Smht04.202	Smht04.204.7	Smht04.205.6	Smht04.206.6	Smht04.207.3	Smht04.208.4	Smht04.209.3	Smht04.210.6	Smht04.211.8	Smht04.213.9	Smht04.215.9	Smht04.216.7	Smht04.217.9	Smht04.218.9	Smht04.219.9	Smht04.221.2	Smht04.230.5	Smht04.258.6
43	Fst1	0	0	0	1	0	0	0	0	1	0	1	0	1	0	0	0	0	0	0	0	1	0	0
43	Fst2	0	0	1	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
43	Fst3	0	0	1	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
43	Fst4	0	0	1	0	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
43	Fst5	0	0	1	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
43	Fst6	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
44	RnP1	0	0	0	1	0	0	0	0	1	0	1	1	1	0	0	0	0	0	0	0	0	0	0
44	RnP2	0	0	0	1	0	0	0	0	1	1	0	1	1	0	0	0	0	0	0	1	0	0	0
44	RnP3	0	1	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
44	RnP4	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
44	RnP5	0	0	1	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	1	0	0
44	RnP6	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
45	RaS1	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
45	RaS3	0	0	0	1	0	0	0	0	1	0	1	1	0	0	1	0	0	0	0	0	0	0	0
45	RaS5	0	0	1	1	0	0	0	0	1	0	0	0	1	0	1	0	0	0	0	1	0	0	0
45	RaS7	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	0	0	0
45	RaS8	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
45	RaS2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
45	RaS4	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	1	0	0	0
46	THS1	0	0	0	1	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	1	1	0	0
46	THS2	0	0	0	1	0	0	0	0	1	0	0	1	1	0	1	0	0	0	0	1	0	0	0
46	THS3	0	0	0	1	0	0	0	0	1	0	0	1	0	0	1	0	0	0	0	0	0	0	0
46	THS4	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
46	THS5	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
46	THS6	0	0	0	1	0	0	0	0	1	0	0	1	1	0	0	0	0	0	0	0	0	0	0
46	THS7	0	0	0	1	0	0	0	0	1	0	0	0	1	0	1	0	0	0	0	1	1	0	0

Part 2

Population	Sample ID	Bra06.374.3	Bra06.377.2	Bra06.387.5	Bra06.390.3	Bra06.393.5	Bra06.394.6	Bra06.396.3	Bra06.397.5	Bra06.398.4	Bra06.399.2	Bra06.400.5	Bra06.402.2	Bra06.403.6	Bra06.405.8	Bra06.406.6	Bra06.408.5	Bra06.409.7	Bra06.411.5	Bra06.412.8	Bra06.413.8	Bra06.415.8	Bra06.416.9	Bra06.418.8
10	SaB7	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
10	SaB8	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
10	SaB2	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
10	SaB4	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
11	Art1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	1	0	0
11	Art3	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	1	0	0	0	0
11	Art5	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	1	0	0
11	Art7	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	1	0	1	0	0
11	Art9	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	1	0	0
11	Art2	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	0	0	0	0	0	0
11	Art8	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	1	0	0	0	0
11	Art10	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0
12	Mrn705	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0	1	0	0	0
12	Mrn706	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	1
12	Mrn707	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
12	Mrn708	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0
12	Mrn709	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
12	Mrn704	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0	1	0	0	0
12	Mrn710	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
13	BwR684	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	1	0	0	0	0
13	BwR685	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
13	BwR689	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0
13	BwR687	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0
13	BwR688	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0					

[illegible]

[illegible]

Population	Sample ID	Bra06.374.3	Bra06.377.2	Bra06.387.5	Bra06.390.3	Bra06.393.5	Bra06.394.6	Bra06.396.3	Bra06.397.5	Bra06.398.4	Bra06.399.2	Bra06.400.5	Bra06.402.2	Bra06.403.6	Bra06.405.8	Bra06.406.6	Bra06.408.5	Bra06.409.7	Bra06.411.5	Bra06.412.8	Bra06.413.8	Bra06.415.8	Bra06.416.9	Bra06.418.8
39	WDM7	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0
40	WhG1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0
40	WhG2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
40	WhG3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1
40	WhG4	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0
40	WhG5	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0
40	WhG6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
40	WhG7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
41	BkG1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
41	BkG2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
41	BkG3	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
41	BkG4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
41	BkG5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
41	BkG6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
42	Maa1	1	0	1	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
42	Maa2	1	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
42	Maa3	0	0	1	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
42	Maa4	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0
42	Maa5	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	1	0	1	0	0	0	0
42	Maa6	1	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
42	Maa7	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0
43	Fst1	0	0	0	0	0	0	1	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0
43	Fst2	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0
43	Fst3	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
43	Fst4	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0
43	Fst5	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
43	Fst6	0	0	1	0	0	0	0	0	0	0	0	1	1	0	0	1	0	0	0	0	0	0	0
44	RnP1	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0
44	RnP2	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
44	RnP3	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
44	RnP4	0	1	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0
44	RnP5	0	0	0	0	1	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
44	RnP6	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
45	RaS1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0
45	RaS3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
45	RaS5	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0
45	RaS7	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0
45	RaS8	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0	0	0	0	0	0	0	0
45	RaS2	0	0	1	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	1	0	0	0	0
45	RaS4	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
46	THS1	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0
46	THS2	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0
46	THS3	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
46	THS4	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	1	0	0	0	0
46	THS5	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0
46	THS6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
46	THS7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0

Population	Sample ID	Bra06.420.7	Bra06.421.9	Bra06.423.7	Bra06.425	Bra06.428	Bra06.429.9	Bra06.431.1	Bra06.434.1	Bra06.437.1	Bra06.475	Bra09.295.6	Bra09.300.7	Bra09.302	Bra09.305.1	Bra09.308.6	Bra09.312.4	Bra09.314.8	Bra09.317.7	Bra09.320.7	Bra09.324.1	Bra09.327.1	Bra09.330.3	Bra09.333.5
1	Kwk2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
1	Kwk3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
1	Kwk4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
1	Kwk5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
1	Kwk6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
1	Kwk7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
2	RgG1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
2	RgG3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
2	RgG5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
2	RgG7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
2	RgG10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
2	RgG9	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
3	HpR1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
3	HpR2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
3	HpR3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
3	HpR4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
3	HpR5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
3	HpR7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
4	RtH1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
4	RtH2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
4	RtH3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
4	RtH4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
4	RtH5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
4	RtH8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
5	RRs2	0																						

Population	Sample ID	Bra06.420.7	Bra06.421.9	Bra06.423.7	Bra06.425	Bra06.428	Bra06.429.9	Bra06.431.1	Bra06.434.1	Bra06.437.1	Bra06.475	Bra09.295.6	Bra09.300.7	Bra09.302	Bra09.305.1	Bra09.308.6	Bra09.312.4	Bra09.314.8	Bra09.317.7	Bra09.320.7	Bra09.324.1	Bra09.327.1	Bra09.330.3	Bra09.333.5
10	SaB7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0
10	SaB8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
10	SaB2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
10	SaB4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
11	Art1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
11	Art3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
11	Art5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	0
11	Art7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1
11	Art9	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0	1	1
11	Art2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	1
11	Art8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0	1	0
11	Art10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
12	Mrn705	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
12	Mrn706	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
12	Mrn707	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
12	Mrn708	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
12	Mrn709	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
12	Mrn704	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
12	Mrn710	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
13	BwR684	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
13	BwR685	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
13	BwR689	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
13	BwR687	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
13	BwR688	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
14	Lew698	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	1	1
14	Lew700	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
14	Lew701	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	1	0
14	Lew702	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	1	1
14	Lew703	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
14	Lew699	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1	0
15	Den690	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1	0
15	Den693	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
15	Den694	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1
15	Den697	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
15	Den698	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	1	1
15	Den691											0	0	0	0	0	0	0	1	0	0	1	1	0
16	Cha674	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1
16	Cha675	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	1
16	Cha676	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	1
16	Cha681	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	1	1
16	Cha682	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0
16	Cha677	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
16	Cha678	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	1
17	Swp733	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1	1
17	Swp734	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1	0
17	Swp735	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	0
17	Swp736	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
17	Swp738	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	0	1	0
17	Swp737	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0
18	OtV2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
18	OtV3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
18	OtV6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
18	OtV7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0
18	OtV8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
18	OVT10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
19	BVTD1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	1	0
19	BVTD2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0	0

Population	Sample ID	Bra06.420.7	Bra06.421.9	Bra06.423.7	Bra06.425	Bra06.428	Bra06.429.9	Bra06.431.1	Bra06.434.1	Bra06.437.1	Bra06.475	Bra09.295.6	Bra09.300.7	Bra09.302	Bra09.305.1	Bra09.308.6	Bra09.312.4	Bra09.314.8	Bra09.317.7	Bra09.320.7	Bra09.324.1	Bra09.327.1	Bra09.330.3	Bra09.333.5
19	BVTD3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	1	0
19	BVTW1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
19	BVTW2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
19	BVTW3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
20	MBT1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
20	MBT2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
20	MBT3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1
20	MBT4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1	0
20	MBT5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	0
20	MBT6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
20	MBT7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
21	Cas498	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
21	Cas499	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
21	Cas500	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
22	CvS510	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
22	CvS511	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	1	0
22	CvS512	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0
22	CvS513	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
22	CvS514	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0
23	PoP515	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
23	PoP516	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	1	0
23	PoP517	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0
23	PoP518	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
23	PoP519	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
23	MCV8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
24	Msm520	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1	0
24	Msm523	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
24	Msm521	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
24	Msm522	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
24	Msm524	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0
25	PrG1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0
25	PrG2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
25	PrG3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
25	PrG4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
25	PrG5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0
26	Gdy1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
26	Gdy2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
26	Gdy3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0
26	Gdy4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0
26	Gdy5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
26	Gdy6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
26	Gdy7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0
27	PHSL493	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1	0
27	PHSL494	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1	0
27	PHSL495	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
27	PHSL496	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1	0
27	PHSL497	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
28	PHWT488	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	1	0
28	PHWT490	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0
28	PHWT491	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0
28	PHWT489	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
28	PHWT492	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
29	BPSR471	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
29	BPSR472	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
29	BP473	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	1	1
29	BP474	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
29	BP475	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0

Population	Sample ID	Bra06.420.7	Bra06.421.9	Bra06.423.7	Bra06.425	Bra06.428	Bra06.429.9	Bra06.431.1	Bra06.434.1	Bra06.437.1	Bra06.475	Bra09.295.6	Bra09.300.7	Bra09.302	Bra09.305.1	Bra09.308.6	Bra09.312.4	Bra09.314.8	Bra09.317.7	Bra09.320.7	Bra09.324.1	Bra09.327.1	Bra09.330.3	Bra09.333.5
29	BPSR476	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
29	BPSR477	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
30	BPLR478	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
30	BPLR479	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	1	1
30	BPLR481	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
30	BPLR482	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
31	Mld753	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1
31	Mld754	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
31	Mld755	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1
31	Mld756	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
32	MCV1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
32	MCV2a	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0
32	MCV3a	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	1	0
32	MCV4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0
32	MCV5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
32	MCV6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
32	MCV7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
33	Lin728	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	1	0
33	Lin729	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0
33	Lin731	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
33	Lin730	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1	0
34	BLd1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
34	BLd2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
34	BLd5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
34	BLd6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
34	BLd7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
34	BLd3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
34	BL4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
35	Rks6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
35	Rks1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
35	Rks2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
35	Rks3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
35	Rks4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
35	Rks5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
36	Wye1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
36	Wye2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
36	Wye3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
37	KyS2	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
37	KyS3	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
37	KyS4	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
37	KyS5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
37	KyS6	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
37	KyS7	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn6	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn8	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn9	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn3	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
39	Wdm1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	1	0
39	Wdm2	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
39	Wdm3	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	1	0
39	Wdm4	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	1	1	0
39	Wdm5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0
39	Wdm6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0

Population	Sample ID	Bra06.420.7	Bra06.421.9	Bra06.423.7	Bra06.425	Bra06.428	Bra06.429.9	Bra06.431.1	Bra06.434.1	Bra06.437.1	Bra06.475	Bra09.295.6	Bra09.300.7	Bra09.302	Bra09.305.1	Bra09.308.6	Bra09.312.4	Bra09.314.8	Bra09.317.7	Bra09.320.7	Bra09.324.1	Bra09.327.1	Bra09.330.3	Bra09.333.5
39	Wdm7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
40	WhG1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
40	WhG2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
40	WhG3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
40	WhG4	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
40	WhG5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
40	WhG6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
40	WhG7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
41	BkG1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
41	BkG2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
41	BkG3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
41	BkG4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
41	BkG5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
41	BkG6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
42	Maa1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
42	Maa2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1
42	Maa3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
42	Maa4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
42	Maa5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
42	Maa6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
42	Maa7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
43	Fst1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
43	Fst2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
43	Fst3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
43	Fst4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
43	Fst5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
43	Fst6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
44	RnP1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
44	RnP2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
44	RnP3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
44	RnP4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
44	RnP5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
44	RnP6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
45	RaS1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
45	RaS3	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
45	RaS5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0
45	RaS7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1
45	RaS8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
45	RaS2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
45	RaS4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
46	THS1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
46	THS2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
46	THS3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
46	THS4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
46	THS5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
46	THS6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
46	THS7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0

Part 4

[illegible]

Population	Sample ID	Bra09.336.5	Bra09.339.4	Bra12.247.2	Bra12.250	Bra12.252.8	Bra12.255.8	Bra12.258.8	Bra12.261.8	Bra12.267.6	Bra19.203	Bra19.222.6	Bra19.225.5	Bra19.237.7	Bra19.240.9	Bra19.243.5	Bra19.246.7	Bra19.249.9	Bra19.252.8	Bra19.255.5	Bra19.258.3	Bra19.261	Bra19.263.7	Bra19.265.7
19	BVTD3	0	0	0	0	1	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
19	BVTW1	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
19	BVTW2	1	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
19	BVTW3	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
20	MBT1	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
20	MBT2	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0
20	MBT3	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
20	MBT4	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
20	MBT5	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
20	MBT6	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
20	MBT7	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
21	Cas498	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
21	Cas499	0	0	0	0	1	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
21	Cas500	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
22	CvS510	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
22	CvS511	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
22	CvS512	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
22	CvS513	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
22	CvS514	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
23	PoP515	0	0	0	0	1	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0
23	PoP516	0	0	0	0	1	1	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
23	PoP517	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
23	PoP518	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0
23	PoP519	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
23	MCV8	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
24	Msm520	0	0	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
24	Msm523	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0
24	Msm521	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
24	Msm522	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
24	Msm524	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
25	PrG1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
25	PrG2	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
25	PrG3	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
25	PrG4	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
25	PrG5	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy2	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy3	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy4	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0
26	Gdy5	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy6	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy7	1	0	0	0	1	1	1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0
27	PHSL493	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
27	PHSL494	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
27	PHSL495	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0
27	PHSL496	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
27	PHSL497	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
28	PHWT488	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
28	PHWT490	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
28	PHWT491	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
28	PHWT489	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
28	PHWT492	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
29	BPSR471	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
29	BPSR472	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
29	BP473	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0
29	BP474	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1
29	BP475	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0

Population	Sample ID	Bra09.336.5	Bra09.339.4	Bra12.247.2	Bra12.250	Bra12.252.8	Bra12.255.8	Bra12.258.8	Bra12.261.8	Bra12.267.6	Bra19.203	Bra19.222.6	Bra19.225.5	Bra19.237.7	Bra19.240.9	Bra19.243.5	Bra19.246.7	Bra19.249.9	Bra19.252.8	Bra19.255.5	Bra19.258.3	Bra19.261	Bra19.263.7	Bra19.265.7
29	BPSR476	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
29	BPSR477	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0
30	BPLR478	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
30	BPLR479	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
30	BPLR481	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
30	BPLR482	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
31	Mld753	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
31	Mld754	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
31	Mld755	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
31	Mld756	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
32	MCV1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
32	MCV2a	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
32	MCV3a	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0
32	MCV4	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
32	MCV5	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0
32	MCV6	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
32	MCV7	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0
33	Lin728	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
33	Lin729	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0
33	Lin731	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
33	Lin730	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
34	BLd1	1	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
34	BLd2	0	0	0	0	1	0	0	0	0														
34	BLd5	0	0	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
34	BLd6	0	0	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
34	BLd7	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
34	BLd3	0	1	0	0	1	0	1	0	0														
34	BL4	1	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
35	Rks6	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
35	Rks1	0	0	0	0	1	1	0	0	0														
35	Rks2	0	0	0	0	0	1	0	0	0														
35	Rks3	0	0	0	0	1	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
35	Rks4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
35	Rks5	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
36	Wye1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
36	Wye2	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0
36	Wye3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
37	KyS2	0	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
37	KyS3	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
37	KyS4	0	0	0	1	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
37	KyS5	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
37	KyS6	0	0	0	1	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
37	KyS7	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
38	MBn1	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
38	MBn6	0	0	0	1	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
38	MBn7	0	0	0	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
38	MBn8	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
38	MBn9	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
38	MBn10	0	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
38	MBn3	1	0	0	1	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
38	MBn4	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
39	WDM1	1	0	0	1	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0
39	WDM2	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
39	WDM3	0	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
39	WDM4	0	0	0	1	1	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
39	WDM5	1	0	0	1	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
39	WDM6	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0

Population	Sample ID	Bra09.336.5	Bra09.339.4	Bra12.247.2	Bra12.250	Bra12.252.8	Bra12.255.8	Bra12.258.8	Bra12.261.8	Bra12.267.6	Bra19.203	Bra19.222.6	Bra19.225.5	Bra19.237.7	Bra19.240.9	Bra19.243.5	Bra19.246.7	Bra19.249.9	Bra19.252.8	Bra19.255.5	Bra19.258.3	Bra19.261	Bra19.263.7	Bra19.265.7
39	WDm7	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
40	WhG1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
40	WhG2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
40	WhG3	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
40	WhG4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
40	WhG5	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
40	WhG6	0	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
40	WhG7	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
41	BkG1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
41	BkG2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
41	BkG3	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
41	BkG4	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
41	BkG5	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
41	BkG6	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
42	Maa1	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
42	Maa2	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
42	Maa3	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
42	Maa4	0	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
42	Maa5	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
42	Maa6	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
42	Maa7	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
43	Fst1	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
43	Fst2	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
43	Fst3	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
43	Fst4	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
43	Fst5	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
43	Fst6	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
44	RnP1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
44	RnP2	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
44	RnP3	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
44	RnP4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
44	RnP5	0	0	0	0	0	1	0	0	0														
44	RnP6	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
45	RaS1	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
45	RaS3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
45	RaS5	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
45	RaS7	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
45	RaS8	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
45	RaS2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
45	RaS4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
46	THS1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
46	THS2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
46	THS3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
46	THS4	0	0	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
46	THS5	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
46	THS6	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
46	THS7	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0

[illegible]

[illegible]

[illegible]

Population	Sample ID	Bra19.266.5	Bra19.269.3	Bra19.272.2	Bra19.274.7	Bra19.280.5	Bra19.283.4	Bra19.286.6	Bra19.289.1	Bra19.294.6	Bra19.297.6	Bra19.300.6	Bra19.302.2	Bra19.303.4	Bra19.306.4	Bra19.309.6	Bra19.312.4	Bra19.315.3	Bra19.319	Bra19.322.5	Bra19.324.8	Bra19.349.5	Bra19.357	Bra19.360
29	BPSR476	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
29	BPSR477	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
30	BPLR478	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
30	BPLR479	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
30	BPLR481	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
30	BPLR482	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
31	Mld753	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
31	Mld754	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
31	Mld755	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
31	Mld756	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
32	MCV1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
32	MCV2a	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
32	MCV3a	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
32	MCV4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
32	MCV5	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
32	MCV6	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
32	MCV7	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
33	Lin728	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
33	Lin729	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
33	Lin731	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
33	Lin730	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
34	BLd1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
34	BLd2																							
34	BLd5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
34	BLd6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
34	BLd7	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
34	BLd3																							
34	BL4	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
35	Rks6	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
35	Rks1																							
35	Rks2																							
35	Rks3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
35	Rks4	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
35	Rks5	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
36	Wye1	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
36	Wye2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
36	Wye3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
37	KyS2	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0
37	KyS3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
37	KyS4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
37	KyS5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
37	KyS6	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
37	KyS7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
38	MBn1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
38	MBn6	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0
38	MBn7	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
38	MBn8	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
38	MBn9	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
38	MBn10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
38	MBn3	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0
38	MBn4	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
39	WDm1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
39	WDm2	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0
39	WDm3	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
39	WDm4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
39	WDm5	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
39	WDm6	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0

Part 6

Population	Sample ID	Bra19.366	Bra19.371.6	Bra19.374.3	Bra19.377	Bra21.199	Bra21.205.5	Bra21.212.1	Bra21.215.6	Bra21.218.7	Bra21.222.1	Bra21.225.2	Bra21.228.7	Bra21.231.8	Bra21.235.2	Bra21.238.3	Bra21.240.7	Bra21.242.1	Bra21.244.9	Bra21.251	Bra21.254.2	Bra21.256.9	Bra21.262.9	Bra21.268.8
1	Kwk2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
1	Kwk3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
1	Kwk4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
1	Kwk5	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
1	Kwk6	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
1	Kwk7	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
2	RgG1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
2	RgG3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
2	RgG5	0	0	0	0																			
2	RgG7	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
2	RgG10	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
2	RgG9	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
3	HpR1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
3	HpR2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
3	HpR3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
3	HpR4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
3	HpR5	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
3	HpR7	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
4	RtH1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
4	RtH2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
4	RtH3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
4	RtH4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
4	RtH5	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
4	RtH8	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
5	RRs2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
5	RRs3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
5	RRs4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
5	RRs5	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
5	RRs6	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
5	RRs88	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
6	MFy1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
6	MFy2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
6	MFy3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
6	MFy10	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
6	MFy11	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
6	MFy7	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
7	Blv742	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	
7	Blv743	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	
7	Blv744	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	
7	Blv745	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0	0	
8	Rly1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
8	Rly2	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
8	Rly3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
8	Rly4	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
8	Rly5	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
9	RdH1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	
9	RdH2	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
9	RdH4	0	0	0	0																			
9	RdH6	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
9	RdH7	0	0	0	0																			
9	RdH3	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
9	RdH5	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	
10	SaB1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	
10	SaB3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	
10	SaB5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	

Population	Sample ID	Bra19.366	Bra19.371.6	Bra19.374.3	Bra19.377	Bra21.199	Bra21.205.5	Bra21.212.1	Bra21.215.6	Bra21.218.7	Bra21.222.1	Bra21.225.2	Bra21.228.7	Bra21.231.8	Bra21.235.2	Bra21.238.3	Bra21.240.7	Bra21.242.1	Bra21.244.9	Bra21.251	Bra21.254.2	Bra21.256.9	Bra21.262.9	Bra21.268.8
10	SaB7	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0
10	SaB8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0
10	SaB2	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0
10	SaB4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
11	Art1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
11	Art3	0	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
11	Art5	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
11	Art7	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0
11	Art9	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
11	Art2	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
11	Art8	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
11	Art10	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0
12	Mrn705	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0
12	Mrn706	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
12	Mrn707	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
12	Mrn708	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
12	Mrn709	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0
12	Mrn704	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0
12	Mrn710	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
13	BwR684	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
13	BwR685	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
13	BwR689	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
13	BwR687	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
13	BwR688	0	0	0	0																			
14	Lew698	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
14	Lew700	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
14	Lew701	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
14	Lew702	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
14	Lew703	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
14	Lew699	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
15	Den690	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
15	Den693	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
15	Den694	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
15	Den697	0	0	0	0																			
15	Den698	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
15	Den691	0	0	0	0																			
16	Cha674	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
16	Cha675	0	0	0	0																			
16	Cha676	0	0	0	0																			
16	Cha681	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
16	Cha682	0	0	0	0																			
16	Cha677	0	0	0	0																			
16	Cha678	0	0	0	0																			
17	Swp733	0	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
17	Swp734					0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
17	Swp735	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
17	Swp736	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
17	Swp738	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
17	Swp737	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
18	OtV2	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
18	OtV3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0
18	OtV6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1
18	OtV7	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0
18	OtV8	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0	0	0
18	OVT10	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0
19	BVTD1	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
19	BVTD2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0

Population	Sample ID	Bra19.366	Bra19.371.6	Bra19.374.3	Bra19.377	Bra21.199	Bra21.205.5	Bra21.212.1	Bra21.215.6	Bra21.218.7	Bra21.222.1	Bra21.225.2	Bra21.228.7	Bra21.231.8	Bra21.235.2	Bra21.238.3	Bra21.240.7	Bra21.242.1	Bra21.244.9	Bra21.251	Bra21.254.2	Bra21.256.9	Bra21.262.9	Bra21.268.8
19	BVTD3	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
19	BVTW1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
19	BVTW2	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0
19	BVTW3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
20	MBT1	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
20	MBT2	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1
20	MBT3	0	0	0	1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0
20	MBT4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0
20	MBT5	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0
20	MBT6	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0
20	MBT7	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
21	Cas498	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
21	Cas499	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0	0	0	0	0
21	Cas500	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0
22	CvS510	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0
22	CvS511	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0
22	CvS512	0	0	1	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
22	CvS513	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
22	CvS514	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
23	PoP515	0	0	1	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
23	PoP516	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
23	PoP517	0	1	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0
23	PoP518	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0
23	PoP519	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
23	MCV8	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
24	Msm520	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
24	Msm523	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
24	Msm521	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0
24	Msm522	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
24	Msm524	0	0	0	0																			
25	PrG1	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
25	PrG2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
25	PrG3	0	0	0	0																			
25	PrG4	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
25	PrG5	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy1	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
26	Gdy2	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy3	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0
26	Gdy4	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0
26	Gdy5	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy6	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26	Gdy7	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
27	PHSL493	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
27	PHSL494	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
27	PHSL495	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
27	PHSL496	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
27	PHSL497	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
28	PHWT488	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0
28	PHWT490	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
28	PHWT491	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0
28	PHWT489	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0
28	PHWT492	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0
29	BPSR471	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0
29	BPSR472	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0	0	0
29	BP473	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1
29	BP474	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
29	BP475	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0

[illegible]

[illegible]

Part 7

Population	Sample ID	Bra21.271.1	Bra21.274.2	Bra21.277.1	Bra21.280	Bra21.285.8	Bra21.306.6	Bra25.342.3	Bra25.348.4	Bra25.356.5	Bra25.359.7	Bra25.362.2	Bra25.364.8	Bra25.367.8	Bra25.369.9	Bra25.371	Bra25.372.9	Bra25.373.8	Bra25.376.7	Bra25.379.6	Bra25.382.4	Bra25.384	Bra25.385.4	Bra25.388.4
1	Kwk2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
1	Kwk3	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1
1	Kwk4	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1	Kwk5	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1	Kwk6	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
1	Kwk7	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1
2	RgG1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	RgG3	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	RgG5							0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	RgG7	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	RgG10	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2	RgG9	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
3	HpR1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
3	HpR2	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0
3	HpR3	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0
3	HpR4	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0
3	HpR5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
3	HpR7	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0
4	RtH1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1
4	RtH2	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0
4	RtH3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
4	RtH4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
4	RtH5	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1
4	RtH8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
5	RRs2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
5	RRs3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
5	RRs4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
5	RRs5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
5	RRs6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
5	RRs88	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
6	MFy1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
6	MFy2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
6	MFy3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
6	MFy10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
6	MFy11	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
6	MFy7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
7	Blv742	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0
7	Blv743	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	1	0
7	Blv744	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	1	0	0	0	0	0
7	Blv745	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
8	Rly1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
8	Rly2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
8	Rly3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
8	Rly4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
8	Rly5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
9	RdH1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
9	RdH2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
9	RdH4							0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0
9	RdH6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
9	RdH7							0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
9	RdH3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
9	RdH5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
10	SaB1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
10	SaB3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
10	SaB5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0

Population	Sample ID	Bra21.271.1	Bra21.274.2	Bra21.277.1	Bra21.280	Bra21.285.8	Bra21.306.6	Bra25.342.3	Bra25.348.4	Bra25.356.5	Bra25.359.7	Bra25.362.2	Bra25.364.8	Bra25.367.8	Bra25.369.9	Bra25.371	Bra25.372.9	Bra25.373.8	Bra25.376.7	Bra25.379.6	Bra25.382.4	Bra25.384	Bra25.385.4	Bra25.388.4
10	SaB7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
10	SaB8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
10	SaB2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
10	SaB4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
11	Art1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
11	Art3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
11	Art5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
11	Art7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
11	Art9	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
11	Art2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
11	Art8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
11	Art10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
12	Mrn705	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
12	Mrn706	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
12	Mrn707	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
12	Mrn708	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	1	1	0	0	0
12	Mrn709	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
12	Mrn704	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
12	Mrn710	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
13	BwR684	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
13	BwR685	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
13	BwR689	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
13	BwR687	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
13	BwR688							0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
14	Lew698	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
14	Lew700	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
14	Lew701	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
14	Lew702	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
14	Lew703	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
14	Lew699	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
15	Den690	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
15	Den693	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
15	Den694	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	1
15	Den697							0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
15	Den698	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
15	Den691							0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
16	Cha674	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0
16	Cha675							0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
16	Cha676							0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
16	Cha681	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
16	Cha682							0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
16	Cha677							0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
16	Cha678							0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
17	Swp733	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1	1	0	0	0
17	Swp734	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	1	0
17	Swp735	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
17	Swp736	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0
17	Swp738	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
17	Swp737	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	1	0
18	OtV2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
18	OtV3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
18	OtV6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
18	OtV7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
18	OtV8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0
18	OVT10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
19	BVTD1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
19	BVTD2	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	1	0	0	0

Population	Sample ID	Bra21.271.1	Bra21.274.2	Bra21.277.1	Bra21.280	Bra21.285.8	Bra21.306.6	Bra25.342.3	Bra25.348.4	Bra25.356.5	Bra25.359.7	Bra25.362.2	Bra25.364.8	Bra25.367.8	Bra25.369.9	Bra25.371	Bra25.372.9	Bra25.373.8	Bra25.376.7	Bra25.379.6	Bra25.382.4	Bra25.384	Bra25.385.4	Bra25.388.4
19	BVTD3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
19	BVTW1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
19	BVTW2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
19	BVTW3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
20	MBT1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
20	MBT2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
20	MBT3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
20	MBT4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
20	MBT5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0
20	MBT6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
20	MBT7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
21	Cas498	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0
21	Cas499	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	0	0	0	0	0
21	Cas500	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	0	0	0	0	0
22	CvS510	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
22	CvS511	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
22	CvS512	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
22	CvS513	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
22	CvS514	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
23	PoP515	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
23	PoP516	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
23	PoP517	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
23	PoP518	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
23	PoP519	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
23	MCV8	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0
24	Msm520	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
24	Msm523	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
24	Msm521	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
24	Msm522	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0
24	Msm524							0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
25	PrG1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
25	PrG2	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0
25	PrG3							0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0
25	PrG4	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
25	PrG5	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
26	Gdy1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
26	Gdy2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
26	Gdy3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
26	Gdy4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
26	Gdy5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
26	Gdy6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
26	Gdy7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
27	PHSL493	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
27	PHSL494	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
27	PHSL495	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
27	PHSL496	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
27	PHSL497	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
28	PHWT488	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
28	PHWT490	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
28	PHWT491	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
28	PHWT489	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
28	PHWT492	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
29	BPSR471	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
29	BPSR472	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
29	BP473	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
29	BP474	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
29	BP475	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0

Population	Sample ID	Bra21.271.1	Bra21.274.2	Bra21.277.1	Bra21.280	Bra21.285.8	Bra21.306.6	Bra25.342.3	Bra25.348.4	Bra25.356.5	Bra25.359.7	Bra25.362.2	Bra25.364.8	Bra25.367.8	Bra25.369.9	Bra25.371	Bra25.372.9	Bra25.373.8	Bra25.376.7	Bra25.379.6	Bra25.382.4	Bra25.384	Bra25.385.4	Bra25.388.4
29	BPSR476	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0
29	BPSR477	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0
30	BPLR478	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	1
30	BPLR479	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0
30	BPLR481	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	1
30	BPLR482	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0
31	Mld753	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	1	0	0	0
31	Mld754	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	0	0	0	0	0
31	Mld755	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	1	0	0	0
31	Mld756	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0
32	MCV1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
32	MCV2a	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
32	MCV3a	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
32	MCV4	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0
32	MCV5	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0
32	MCV6	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	0	0	0	0
32	MCV7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
33	Lin728	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1
33	Lin729	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0
33	Lin731	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	1
33	Lin730	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
34	BLd1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
34	BLd2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
34	BLd5	0	1	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	1
34	BLd6	0	1	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	1
34	BLd7	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	1
34	BLd3	0	1	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	1	0	0	0	0	0
34	BL4	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	1	0	0	0
35	Rks6	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	0	0	0	0	0	0
35	Rks1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	0	0	0	0
35	Rks2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
35	Rks3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
35	Rks4	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	0	0	0	0	0
35	Rks5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
36	Wye1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
36	Wye2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
36	Wye3	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0
37	KyS2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
37	KyS3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
37	KyS4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
37	KyS5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
37	KyS6	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0
37	KyS7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
38	MBn1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
38	MBn6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
38	MBn7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	1	0
38	MBn8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0
38	MBn9	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	1	0
38	MBn10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
38	MBn3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
38	MBn4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDM1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDM2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDM3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDM4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDM5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDM6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0

Population	Sample ID	Bra21.271.1	Bra21.274.2	Bra21.277.1	Bra21.280	Bra21.285.8	Bra21.306.6	Bra25.342.3	Bra25.348.4	Bra25.356.5	Bra25.359.7	Bra25.362.2	Bra25.364.8	Bra25.367.8	Bra25.369.9	Bra25.371	Bra25.372.9	Bra25.373.8	Bra25.376.7	Bra25.379.6	Bra25.382.4	Bra25.384	Bra25.385.4	Bra25.388.4
39	WDm7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
40	WhG1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	1	0	0	0
40	WhG2	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1	0	0	0	0	0
40	WhG3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1	0	0	0	0
40	WhG4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
40	WhG5	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
40	WhG6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
40	WhG7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
41	BkG1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
41	BkG2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
41	BkG3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
41	BkG4	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0
41	BkG5	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	1	1	0	0	0	0	0
41	BkG6	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0
42	Maa1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
42	Maa2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	1	0
42	Maa3	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
42	Maa4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
42	Maa5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
42	Maa6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
42	Maa7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
43	Fst1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
43	Fst2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
43	Fst3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
43	Fst4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
43	Fst5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
43	Fst6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
44	RnP1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
44	RnP2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
44	RnP3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
44	RnP4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
44	RnP5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
44	RnP6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
45	RaS1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
45	RaS3	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0	0	0
45	RaS5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
45	RaS7	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	1	0
45	RaS8	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1	0	1	0	0	0
45	RaS2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
45	RaS4	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	1	0
46	THS1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
46	THS2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
46	THS3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
46	THS4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	1	0
46	THS5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
46	THS6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0
46	THS7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0

Part 8

Population	Sample ID	Bra25.391.1	Bra25.393.9	Bra25.396.8	Bra25.402.8	Bra25.405.5	Bra25.408.7	Bra26.256.3	Bra26.258.9	Bra26.262	Bra26.264.5	Bra26.267.8	Bra26.270.3	Bra26.273.2	Bra26.276.1	Bra26.279.2	Bra26.281.9
1	Kwk2	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
1	Kwk3	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
1	Kwk4	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
1	Kwk5	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
1	Kwk6	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
1	Kwk7	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
2	RgG1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
2	RgG3	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
2	RgG5	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
2	RgG7	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
2	RgG10	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
2	RgG9	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
3	HpR1	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
3	HpR2	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
3	HpR3	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
3	HpR4	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
3	HpR5	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
3	HpR7	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
4	RtH1	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
4	RtH2	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
4	RtH3	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
4	RtH4	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
4	RtH5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
4	RtH8	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
5	RRs2	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
5	RRs3	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
5	RRs4	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
5	RRs5	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
5	RRs6	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
5	RRs88	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
6	MFy1	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
6	MFy2	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
6	MFy3	1	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0
6	MFy10	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
6	MFy11	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0
6	MFy7	0	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0
7	Blv742	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
7	Blv743	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0
7	Blv744	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0
7	Blv745	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
8	Rly1	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
8	Rly2	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
8	Rly3	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
8	Rly4	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
8	Rly5	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
9	RdH1	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
9	RdH2	1	1	0	0	0	0	0	0	1	0	0	0	0	0	0	0
9	RdH4	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
9	RdH6	1	1	0	0	0	0	0	0	1	0	0	0	0	0	0	0
9	RdH7	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
9	RdH3	1	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
9	RdH5	1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
10	SaB1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
10	SaB3	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0
10	SaB5	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0

Population	Sample ID	Bra25.391.1	Bra25.393.9	Bra25.396.8	Bra25.402.8	Bra25.405.5	Bra25.408.7	Bra26.256.3	Bra26.258.9	Bra26.262	Bra26.264.5	Bra26.267.8	Bra26.270.3	Bra26.273.2	Bra26.276.1	Bra26.279.2	Bra26.281.9
10	SaB7	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0
10	SaB8	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
10	SaB2	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0
10	SaB4	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0
11	Art1	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
11	Art3	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
11	Art5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
11	Art7	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
11	Art9	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
11	Art2	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
11	Art8	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
11	Art10	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0
12	Mrn705	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
12	Mrn706	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
12	Mrn707	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0
12	Mrn708	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
12	Mrn709	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
12	Mrn704	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
12	Mrn710	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0
13	BwR684	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
13	BwR685	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0
13	BwR689	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
13	BwR687	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0
13	BwR688	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
14	Lew698	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
14	Lew700	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
14	Lew701	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0
14	Lew702	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
14	Lew703	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
14	Lew699	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
15	Den690	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
15	Den693	1	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
15	Den694	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
15	Den697	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
15	Den698	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
15	Den691	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
16	Cha674	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
16	Cha675	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
16	Cha676	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
16	Cha681	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
16	Cha682	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
16	Cha677	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
16	Cha678	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
17	Swp733	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0
17	Swp734	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
17	Swp735	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
17	Swp736	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
17	Swp738	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0
17	Swp737	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
18	OtV2	0	0	1	0	0	0	0	1	0	1	0	0	0	0	0	0
18	OtV3	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
18	OtV6	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
18	OtV7	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
18	OtV8	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0
18	OVT10	1	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0
19	BVTD1	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
19	BVTD2	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0

Population	Sample ID	Bra25.391.1	Bra25.393.9	Bra25.396.8	Bra25.402.8	Bra25.405.5	Bra25.408.7	Bra26.256.3	Bra26.258.9	Bra26.262	Bra26.264.5	Bra26.267.8	Bra26.270.3	Bra26.273.2	Bra26.276.1	Bra26.279.2	Bra26.281.9
19	BVTD3	1	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0
19	BVTW1	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
19	BVTW2	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
19	BVTW3	0	0	1	0	0	0	0	0	1	0	0	1	0	0	0	0
20	MBT1	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
20	MBT2	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
20	MBT3	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
20	MBT4	1	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
20	MBT5	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
20	MBT6	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
20	MBT7	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0
21	Cas498	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
21	Cas499	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
21	Cas500	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
22	CvS510	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
22	CvS511	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0
22	CvS512	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0
22	CvS513	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
22	CvS514	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
23	PoP515	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0
23	PoP516	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
23	PoP517	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
23	PoP518	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0
23	PoP519	0	0	1	0	0	0	0	0	0	0	1	1	0	0	0	0
23	MCV8	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
24	Msm520	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
24	Msm523	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
24	Msm521	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
24	Msm522	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
24	Msm524	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
25	PrG1	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
25	PrG2	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
25	PrG3	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
25	PrG4	0	1	0	0	0	0	0	0	0	0	0	1	1	0	0	0
25	PrG5	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
26	Gdy1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0
26	Gdy2	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
26	Gdy3	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
26	Gdy4	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
26	Gdy5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
26	Gdy6	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
26	Gdy7	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
27	PHSL493	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
27	PHSL494	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
27	PHSL495	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
27	PHSL496	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
27	PHSL497	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
28	PHWT488	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
28	PHWT490	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
28	PHWT491	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
28	PHWT489	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
28	PHWT492	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
29	BPSR471	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
29	BPSR472	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
29	BP473	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0
29	BP474	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
29	BP475	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0

Population	Sample ID	Bra25.391.1	Bra25.393.9	Bra25.396.8	Bra25.402.8	Bra25.405.5	Bra25.408.7	Bra26.256.3	Bra26.258.9	Bra26.262	Bra26.264.5	Bra26.267.8	Bra26.270.3	Bra26.273.2	Bra26.276.1	Bra26.279.2	Bra26.281.9
29	BPSR476	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0
29	BPSR477	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0
30	BPLR478	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0
30	BPLR479	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
30	BPLR481	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
30	BPLR482	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
31	Mld753	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
31	Mld754	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
31	Mld755	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
31	Mld756	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
32	MCV1	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
32	MCV2a	0	0	0	0	1	0	0	0	1	0	0	1	0	0	0	0
32	MCV3a	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
32	MCV4	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0
32	MCV5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
32	MCV6	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0
32	MCV7	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1	0
33	Lin728	0	1	0	0	0	0	0	0	0	0	0	1	0	0	0	1
33	Lin729	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
33	Lin731	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0
33	Lin730	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
34	BLd1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
34	BLd2	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
34	BLd5	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
34	BLd6	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
34	BLd7	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
34	BLd3	0	0	0	1	0	0	0	1	0	0	0	1	0	0	0	0
34	BL4	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
35	Rks6	0	1	0	0	0	0	0	0	1	0	0	1	0	0	0	0
35	Rks1	0	0	0	0	0	1	0	0	0	0	0	1	1	0	0	0
35	Rks2	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
35	Rks3	0	0	0	1	0	0	0	1	0	0	0	1	0	0	0	0
35	Rks4	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
35	Rks5	1	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
36	Wye1	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
36	Wye2	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
36	Wye3	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
37	KyS2	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
37	KyS3	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
37	KyS4	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
37	KyS5	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0
37	KyS6	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
37	KyS7	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
38	MBn1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
38	MBn6	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
38	MBn7	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
38	MBn8	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
38	MBn9	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
38	MBn10	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
38	MBn3	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
38	MBn4	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDm1	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
39	WDm2	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDm3	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
39	WDm4	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
39	WDm5	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
39	WDm6	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0

Population	Sample ID	Bra25.391.1	Bra25.393.9	Bra25.396.8	Bra25.402.8	Bra25.405.5	Bra25.408.7	Bra26.256.3	Bra26.258.9	Bra26.262	Bra26.264.5	Bra26.267.8	Bra26.270.3	Bra26.273.2	Bra26.276.1	Bra26.279.2	Bra26.281.9
39	Wdm7	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
40	WhG1	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
40	WhG2	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
40	WhG3	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0
40	WhG4	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
40	WhG5	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
40	WhG6	0	0	0	1	0	0	0	1	0	0	0	1	0	0	0	0
40	WhG7	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0
41	BkG1	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
41	BkG2	0	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0
41	BkG3	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
41	BkG4	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0
41	BkG5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
41	BkG6	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
42	Maa1	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
42	Maa2	0	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0
42	Maa3	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
42	Maa4	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
42	Maa5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
42	Maa6	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
42	Maa7	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
43	Fst1	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
43	Fst2	0	0	1	0	0	0	0	0	0	0	0	1	0	0	0	0
43	Fst3	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
43	Fst4	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0
43	Fst5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
43	Fst6	0	0	0	0	0	0	1	0	1	0	0	0	0	0	0	0
44	RnP1	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
44	RnP2	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0
44	RnP3	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0
44	RnP4	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
44	RnP5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
44	RnP6	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
45	RaS1	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
45	RaS3	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
45	RaS5	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
45	RaS7	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0
45	RaS8	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
45	RaS2	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
45	RaS4	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
46	THS1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
46	THS2	0	1	0	0	0	0	0	0	0	0	1	1	0	0	0	0
46	THS3	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
46	THS4	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0
46	THS5	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0
46	THS6	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0
46	THS7	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0